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(54) Title: MULTIVALENT ANTIGEN-BINDING PROTEINS

(57) Abstract

Compositions of, genetic constructions coding for, and methods for producing multivalent antigen-binding proteins are described and claimed. The methods include purification of compositions containing both monomeric and multivalent forms of single polypeptide chain molecules, and production of multivalent proteins from purified monomers. Production of multivalent proteins may occur by a concentration-dependent association of monomeric proteins, or by rearrangement of regions involving dissociation followed by reassociation of different regions. Bivalent proteins, including homobivalent and heterobivalent proteins, are made in the present invention. Genetic sequences coding for bivalent single-chain antigen-binding proteins are disclosed. Uses include all those appropriate for monoclonal and polyclonal antibodies and fragments thereof, including use as a bispecific antigen-binding molecule.

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Multivalent Antigen-Binding Proteins

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Cross-Reference to Related Applications

This application is a continuation-in-part of U.S. Patent Application Serial Number 07/796,936, filed Nov. 25, 1991, which is a continuation-in-part of U.S. Patent Application Serial No. 07/512,910 filed April 25, 1990, which is a continuation-in-part of Serial No. 07/299,617, filed Jan. 1, 1989, issued as U.S. Patent No. 4,946,778 (Ladner *et al.*), which was a continuation-in-part of Serial No. 092,110, filed Sept. 2, 1987, and Serial No. 902,971, filed Sept. 2, 1986, now abandoned, the contents of all of which are fully incorporated herein by reference.

Background of the Invention

1. Field of the Invention

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The present invention relates generally to the production of antigenbinding molecules. More specifically, the invention relates to multivalent forms of antigen-binding proteins. Compositions of, genetic constructions for, methods of use, and methods for producing these multivalent antigen-binding proteins are disclosed.

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2. Description of the Background Art

Antibodies are proteins generated by the immune system to provide a specific molecule capable of complexing with an invading molecule, termed an antigen. Figure 14 shows the structure of a typical antibody molecule. Natural antibodies have two identical antigen-binding sites, both of which are specific to a particular antigen. The antibody molecule "recognizes" the antigen by complexing its antigen-binding sites with areas of the antigen termed epitopes. The epitopes fit into the conformational architecture of the antigen-binding sites of the antibody, enabling the antibody to bind to the antigen.

The antibody molecule is composed of two identical heavy and two identical light polypeptide chains, held together by interchain disulfide bonds (see Fig. 14). The remainder of this discussion will refer only to one light/heavy pair of chains, as each light/heavy pair is identical. Each individual light and heavy chain folds into regions of approximately 110 amino acids, assuming a conserved three-dimensional conformation. The light chain comprises one variable region (termed V_L) and one constant region (C_L), while the heavy chain comprises one variable region (V_H) and three constant regions (V_H). Pairs of regions associate to form discrete structures as shown in Figure 14. In particular, the light and heavy chain variable regions, V_L and V_H , associate to form an " V_L " area which contains the antigen-binding site.

The variable regions of both heavy and light chains show considerable variability in structure and amino acid composition from one antibody molecule to another, whereas the constant regions show little variability. The term "variable" as used in this specification refers to the diverse nature of the amino acid sequences of the antibody heavy and light chain variable regions. Each antibody recognizes and binds antigen through the binding site defined by the association of the heavy and light chain variable regions into an F_V area. The light-chain variable region V_L and the heavy-chain variable region V_R of a particular antibody molecule have specific amino acid sequences that

allow the antigen-binding site to assume a conformation that binds to the antigen epitope recognized by that particular antibody.

Within the variable regions are found regions in which the amino acid sequence is extremely variable from one antibody to another. Three of these so-called "hypervariable" regions or "complementarity-determining regions" (CDR's) are found in each of the light and heavy chains. The three CDR's from a light chain and the three CDR's from a corresponding heavy chain form the antigen-binding site.

Cleavage of the naturally-occurring antibody molecule with the proteolytic enzyme papain generates fragments which retain their antigenbinding site. These fragments, commonly known as Fab's (for Fragment, antigen binding site) are composed of the C_L , V_L , C_H1 and V_H regions of the antibody. In the Fab the light chain and the fragment of the heavy chain are covalently linked by a disulfide linkage.

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Recent advances in immunobiology, recombinant DNA technology, and computer science have allowed the creation of single polypeptide chain molecules that bind antigen. These single-chain antigen-binding molecules incorporate a linker polypeptide to bridge the individual variable regions, V_L and V_H, into a single polypeptide chain. A computer-assisted method for linker design is described more particularly in U.S. Patent No. 4,704,692, issued to Ladner *et al.* in November, 1987, and incorporated herein by reference. A description of the theory and production of single-chain antigen-binding proteins is found in U.S. Patent No. 4,946,778 (Ladner *et al.*), issued August 7, 1990, and incorporated herein by reference. The single-chain antigen-binding proteins produced under the process recited in U.S. Patent 4,946,778 have binding specificity and affinity substantially similar to that of the corresponding Fab fragment.

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Bifunctional, or bispecific, antibodies have antigen binding sites of different specificities. Bispecific antibodies have been generated to deliver cells, cytotoxins, or drugs to specific sites. An important use has been to deliver host cytotoxic cells, such as natural killer or cytotoxic T cells, to specific cellular targets. (U.D. Staerz, O. Kanagawa, M.J. Bevan, *Nature*

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314:628 (1985); S. Songilvilai, P.J. Lachmann, Clin. Exp. Immunol. 79: 315 (1990)). Another important use has been to deliver cytotoxic proteins to specific cellular targets. (V. Raso, T. Griffin, Cancer Res. 41:2073 (1981); S. Honda, Y. Ichimori, S. Iwasa, Cytotechnology 4:59 (1990)). Another important use has been to deliver anti-cancer non-protein drugs to specific cellular targets (J. Corvalan, W. Smith, V. Gore, Intl. J. Cancer Suppl. 2:22 (1988); M. Pimm et al., British J. of Cancer 61:508 (1990)). Such bispecific antibodies have been prepared by chemical cross-linking (M. Brennan et al., Science 229:81 (1985)), disulfide exchange, or the production of hybrid-hybridomas (quadromas). Quadromas are constructed by fusing hybridomas that secrete two different types of antibodies against two different antigens (Kurokawa, T. et al., Biotechnology 7:1163 (1989)).

Summary of the Invention

This invention relates to the discovery that multivalent forms of singlechain antigen-binding proteins have significant utility beyond that of the monovalent single-chain antigen-binding proteins. A multivalent antigenbinding protein has more than one antigen-binding site. Enhanced binding activity, di- and multi-specific binding, and other novel uses of multivalent antigen-binding proteins have been demonstrated or are envisioned here. Accordingly, the invention is directed to multivalent forms of single-chain antigen-binding proteins, compositions of multivalent and single-chain antigenbinding proteins, methods of making and purifying multivalent forms of singlechain antigen-binding proteins, and uses for multivalent forms of single-chain antigen-binding proteins. The invention provides a multivalent antigen-binding protein comprising two or more single-chain protein molecules, each singlechain molecule comprising a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; and a peptide linker linking the first and second polypeptides into a single-chain protein.

Also provided is a composition comprising a multivalent antigenbinding protein substantially free of single-chain molecules.

Also provided is an aqueous composition comprising an excess of multivalent antigen-binding protein over single-chain molecules.

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A method of producing a multivalent antigen-binding protein is provided, comprising the steps of producing a composition comprising multivalent antigen-binding protein and single-chain molecules, each single-chain molecule comprising a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; and a peptide linker linking the first and second polypeptides into a single-chain molecule; separating the multivalent protein from the single-chain molecules; and recovering the multivalent protein.

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Also provided is a method of producing multivalent antigen-binding protein, comprising the steps of producing a composition comprising single-chain molecules as previously defined; dissociating the single-chain molecules; reassociating the single-chain molecules; separating the resulting multivalent antigen-binding proteins from the single-chain molecules; and recovering the multivalent proteins.

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Also provided is another method of producing a multivalent antigenbinding protein, comprising the step of chemically cross-linking at least two single-chain antigen-binding molecules.

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Also provided is another method of producing a multivalent antigenbinding protein, comprising the steps of producing a composition comprising single-chain molecules as previously defined; concentrating said single-chain molecules; separating said multivalent protein from said single-chain molecules; and finally recovering said multivalent protein.

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Also provided is another method of producing a multivalent antigenbinding protein comprising two or more single-chain molecules, each singlechain molecule as previously defined, said method comprising: providing a genetic sequence coding for said single-chain molecule; transforming a host cell or cells with said sequence; expressing said sequence in said host or hosts; and recovering said multivalent protein.

Another aspect of the invention includes a method of detecting an antigen in or suspected of being in a sample, which comprises contacting said sample with the multivalent antigen-binding protein of claim 1 and detecting whether said multivalent antigen-binding protein has bound to said antigen.

Another aspect of the invention includes a method of imaging the internal structure of an animal, comprising administering to said animal an effective amount of a labeled form of the multivalent antigen-binding protein of claim 1 and measuring detectable radiation associated with said animal.

Another aspect of the invention includes a composition comprising an association of a multivalent antigen-binding protein with a therapeutically or diagnostically effective agent.

Another aspect of this invention is a single-chain protein comprising: a first polypeptide comprising the binding portion of the variable region of an antibody light chain; a second polypeptide comprising the binding portion of the variable region of an antibody light chain; a peptide linker linking said first and second polypeptides (a) and (b) into said single-chain protein.

Another aspect of the present invention includes the genetic constructions encoding the combinations of regions V_L - V_L and V_H - V_H for single-chain molecules, and encoding multivalent antigen-binding proteins.

Another part of this invention is a multivalent single-chain antigen-binding protein comprising: a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; a peptide linker linking said first and second polypeptides (a) and (b) into said multivalent protein; a third polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; a fourth polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; a peptide linker linking said third and fourth polypeptides (d) and (e) into said multivalent protein; and a peptide linker linking said second and third polypeptides (b) and (d) into said

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multivalent protein. Also included are gentic constructions coding for this multivalent single-chain antigen-binding protein.

Also included are replicable cloning or expression vehicles including plasmids, hosts transformed with the aforementioned genetic sequences, and methods of producing multivalent proteins with the sequences, transformed hosts, and expression vehicles.

Methods of use are provided, such as a method of using the multivalent antigen-binding protein to diagnose a medical condition; a method of using the multivalent protein as a carrier to image the specific bodily organs of an animal; a therapeutic method of using the multivalent protein to treat a medical condition; and an immunotherapeutic method of conjugating a multivalent protein with a therapeutically or diagnostically effective agent. Also included are labelled multivalent proteins, improved immunoassays using them, and improved immunoaffinity purifications.

An advantage of using multivalent antigen-binding proteins instead of single-chain antigen-binding molecules or Fab fragments lies in the enhanced binding ability of the multivalent form. Enhanced binding occurs because the multivalent form has more binding sites per molecule. Another advantage of the present invention is the ability to use multivalent antigen-binding proteins as multi-specific binding molecules.

An advantage of using multivalent antigen-binding proteins instead of whole antibodies, is the enhanced clearing of the multivalent antigen-binding proteins from the serum due to their smaller size as compared to whole antibodies which may afford lower background in imaging applications. Multivalent antigen-binding proteins may penetrate solid tumors better than monoclonals, resulting in better tumor-fighting ability. Also, because they are smaller and lack the Fc component of intact antibodies, the multivalent antigen-binding proteins of the present invention may be less immunogenic than whole antibodies. The Fc component of whole antibodies also contains binding sites for liver, spleen and certain other cells and its absence should thus reduce accumulation in non-target tissues.

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Another advantage of multivalent antigen-binding proteins is the ease with which they may be produced and engineered, as compared to the myeloma-fusing technique pioneered by Kohler and Milstein that is used to produce whole antibodies.

Brief Description of the Drawings.

The present invention as defined in the claims can be better understood with reference to the text and to the following drawings:

FIG. 1A is a schematic two-dimensional representation of two identical single-chain antigen-binding protein molecules, each comprising a variable light chain region (V_L) , a variable heavy chain region (V_H) , and a polypeptide linker joining the two regions. The single-chain antigen-binding protein molecules are shown binding antigen in their antigen-binding sites.

- FIG. 1B depicts a hypothetical homodivalent antigen-binding protein formed by association of the polypeptide linkers of two monovalent single-chain antigen-binding proteins from Fig. 1A (the Association model). The divalent antigen-binding protein is formed by the concentration-driven association of two identical single-chain antigen-binding protein molecules.
- FIG. 1C depicts the hypothetical divalent protein of FIG. 1B with bound antigen molecules occupying both antigen-binding sites.
 - FIG. 2A depicts the hypothetical homodivalent protein of Figure 1B.
- FIG. 2B depicts three single-chain antigen-binding protein molecules associated in a hypothetical trimer.
- FIG. 2C depicts a hypothetical tetramer of four single-chain antigenbinding protein molecules.
- FIG. 3A depicts two separate and distinct monovalent single-chain antigen-binding proteins, Anti-A single-chain antigen-binding protein and Anti-B single-chain antigen-binding protein, with different antigen specificities, each individually binding either Antigen A or Antigen B.

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- FIG. 3B depicts a hypothetical bispecific heterodivalent antigen-binding protein formed from the single-chain antigen-binding proteins of Fig. 3A according to the Association model.
- FIG. 3C depicts the hypothetical heterodivalent antigen-binding protein of FIG. 3B binding bispecifically, i.e., binding the two different antigens, A and B.
- FIG. 4A depicts two identical single-chain antigen-binding protein molecules, each having a variable light chain region (V_L) , a variable heavy chain region (V_H) , and a polypeptide linker joining the two regions. The single-chain antigen-binding protein molecules are shown binding identical antigen molecules in their antigen-binding sites.
- FIG. 4B depicts a hypothetical homodivalent protein formed by the rearrangement of the V_L and V_H regions shown in FIG. 4A (the Rearrangement model). Also shown is bound antigen.
- FIG. 5A depicts two single-chain protein molecules, the first having an anti-B V_L and an anti-A V_H , and the second having an anti-A V_L and an anti-B V_H . The figure shows the non-complementary nature of the V_L and V_H regions in each single-chain protein molecule.
- FIG. 5B shows a hypothetical bispecific heterodivalent antigen-binding protein formed by rearrangement of the two single-chain proteins of Figure 5A.
- FIG. 5C depicts the hypothetical heterodivalent antigen-binding protein of FIG. 5B with different antigens A and B occupying their respective antigenbinding sites.
- FIG. 6A is a schematic depiction of a hypothetical trivalent antigenbinding protein according to the Rearrangement model.
- FIG. 6B is a schematic depiction of a hypothetical tetravalent antigenbinding protein according to the Rearrangement model.
- FIG. 7 is a chromatogram depicting the separation of CC49/212 antigen-binding protein monomer from dimer on a cation exchange high performance liquid chromatographic column. The column is a PolyCAT A

aspartic acid column (Poly WC, Columbia, MD). Monomer is shown as Peak 1, eluting at 27.32 min., and dimer is shown as Peak 2, eluting at 55.52 min.

FIG. 8 is a chromatogram of the purified monomer from Fig. 7. Monomer elutes at 21.94 min., preceded by dimer (20.135 min.) and trimer (18.640 min.). Gel filtration column, Protein-Pak 300SW (Waters Associates, Milford, MA).

FIG. 9 is a similar chromatogram of purified dimer (20.14 min.) from Fig. 7, run on the gel filtration HPLC column of Fig. 8.

FIG. 10A is an amino acid (SEQ ID NO. 11) and nucleotide (SEQ ID NO. 10) sequence of the single-chain protein comprising the 4-4-20 V_L region connected through the 212 linker polypeptide to the CC49 V_H region.

FIG. 10B is an amino acid (SEQ ID NO. 13) and nucleotide (SEQ ID NO. 12) sequence of the single-chain protein comprising the CC49 V_L region connected through the 212 linker polypeptide to the 4-4-20 V_H region.

FIG. 11 is a chromatogram depicting the separation of the monomer (27.83 min.) and dimer (50.47 min.) forms of the CC49/212 antigen-binding protein by cation exchange, on a PolyCAT A cation exchange column (Poly LC, Columbia, MD).

Fig. 12 shows the separation of monomer (17.65 min.), dimer (15.79 min.), trimer (14.19 min.), and higher oligomers (shoulder at about 13.09 min.) of the B6.2/212 antigen-binding protein. This separation depicts the results of a 24-hour treatment of a 1.0 mg/ml B6.2/212 single-chain antigen-binding protein sample. A TSK G2000SW gel filtration HPLC column was used, Toyo Soda, Tokyo, Japan.

Fig. 13 shows the results of a 24-hour treatment of a 4.0 mg/ml CC49/212 antigen-binding protein sample, generating monomer, dimer, and trimer at 16.91, 14.9, and 13.42 min., respectively. The same TSK gel filtration column was used as in Fig. 12.

Fig. 14 shows a schematic view of the four-chain structure of a human IgG molecule.

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Fig. 15A is an amino acid (SEQ ID NO. 15) and nucleotide (SEQ ID NO. 14) sequence of the 4-4-20/212 single-chain antigen-binding protein with a single cysteine hinge.

Fig. 15B is an amino acid (SEQ ID NO. 17) and nucleotide (SEQ. ID NO. 16) sequence of the 4-4-20/212 single-chain antigen-binding protein with the two-cysteine hinge.

Fig. 16 shows the amino acid (SEQ ID NO. 19) and nucleotide (SEQ ID NO. 18) sequence of a divalent CC49/212 single-chain antigen-binding protein.

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Fig. 17 shows the expression of the divalent CC49/212 single-chain antigen-binding protein of Fig. 16 at 42°C, on an SDS-PAGE gel containing total *E. coli* protein. Lane 1 contains the molecular weight standards. Lane 2 is the uninduced *E. coli* production strain grown at 30°C. Lane 3 is divalent CC49/212 single-chain antigen-binding protein induced by growth at 42°C. The arrow shows the band of expressed divalent CC49/212 single-chain antigen-binding protein.

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Fig. 18 is a graphical representation of four competition radioimmunoassays (RIA) in which unlabeled CC49 IgG (open circles) CC49/212 single-chain antigen-binding protein (closed circles) and CC49/212 divalent antigen-binding protein (closed squares) and anti-fluorescein 4-4-20/212 single-chain antigen-binding protein (open squares) competed against a CC49 IgG radiolabeled with ¹²⁵I for binding to the TAG-72 antigen on a human breast carcinoma extract.

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Figure 19A is an amino acid (SEQ ID NO. 21) and nucleotide (SEQ ID NO. 20) sequence of the single-chain polypeptide comprising the 4-4-20 V_L region connected through the 217 linker polypeptide to the CC49 V_H region.

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Figure 19B is an amino acid (SEQ ID NO. 23) and nucleotide (SEQ ID NO. 22) sequence of the single-chain polypeptide comprising the CC49 $\rm V_L$ region connected through the 217 linker polypeptide to the 4-4-20 $\rm V_H$ region.

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Figure 20 is a chromatogram depicting the purification of CC49/4-4-20 heterodimer Fv on a cation exchange high performance liquid chromatographic column. The column is a PolyCAT A aspartic acid column (Poly LC,

Columbia, MD). The heterodimer Fv is shown as peak 5, eluting at 30.10 min.

Figure 21 is a coomassie-blue stained 4-20% SDS-PAGE gel showing the proteins separated in Figure 20. Lane 1 contains the molecular weight standards. Lane 3 contains the starting material before separation. Lanes 4-8 contain fractions 2, 3, 5, 6 and 7 respectively. Lane 9 contains purified CC49/212.

Figure 22A is a chromatogram used to determine the molecular size of fraction 2 from Figure 20. A TSK G3000SW gel filtration HPLC column was used (Toyo Soda, Tokyo, Japan).

Figure 22B is a chromatogram used to determine the molecular size of fraction 5 from Figure 20. A TSK G3000SW gel filtration HPLC column was used (Toyo Soda, Tokyo, Japan).

Figure 22C is a chromatogram used to determine the molecular size of fraction 6 from Figure 20. A TSK G30005W gel filtration HPLC column was used (Toyo Soda, Tokyo, Japan).

Figure 23 shows a Scatchard analysis of the fluorescein binding affinity of the CC49 4-4-20 heterodimer Fv (fraction 5 in Figure 20).

Figure 24 is a graphical representation of three competition enzymelinked immunosorbent assays (ELISA) in which unlabeled CC49 4-4-20 Fv (closed squares) CC49/212 single-chain Fv (open squares) and MOPC-21 IgG (+) competed against a biotin-labeled CC49 IgG for binding to the TAG-72 antigen on a human breast carcinoma extract. MOPC-21 is a control antibody that does not bind to TAG-72 antigen.

Figure 25 shows a coomassie-blue stained non-reducing 4-20% SDS-PAGE gel. Lanes 1 and 9 contain the molecular weight standards. Lane 3 contains the 4-4-20/212 CPPC single-chain antigen-binding protein after purification. Lane 4, 5 and 6 contain the 4-4-20/212 CPPC single-chain antigen-binding protein after treatment with DTT and air oxidation. Lane 7 contains 4-4-20/212 single-chain antigen-binding protein.

Figure 26 shows a coomassie-blue stained reducing 4-20% SDS-PAGE gel (samples were treated with β -mercaptoethanol prior to being loaded on the

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gel). Lanes 1 and 8 contain the molecular weight standards. Lane 3 contains the 4-4-20/212 CPPC single-chain antigen-binding protein after treatment with bis-maleimidehexane. Lane 5 contains peak 1 of bis-maleimidehexane treated 4-4-20/212 CPCC single-chain antigen-binding protein. Lane 6 contains peak 3 of bis-maleimidehexane treated 4-4-20/212 CPPC single-chain antigen-binding protein.

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Detailed Description of the Preferred Embodiments

This invention relates to the discovery that multivalent forms of singlechain antigen-binding proteins have significant utility beyond that of the monovalent single-chain antigen-binding proteins. A multivalent antigenbinding protein has more than one antigen-binding site. For the purposes of this application, "valent" refers to the numerosity of antigen binding sites. Thus, a bivalent protein refers to a protein with two binding sites. Enhanced binding activity, bi- and multi-specific binding, and other novel uses of multivalent antigen-binding proteins have been demonstrated or are envisioned here. Accordingly, the invention is directed to multivalent forms of singlechain antigen-binding proteins, compositions of multivalent and single-chain antigen-binding proteins, methods of making and purifying multivalent forms of single-chain antigen-binding proteins, and new and improved uses for multivalent forms of single-chain antigen-binding proteins. The invention provides a multivalent antigen-binding protein comprising two or more singlechain protein molecules, each single-chain molecule comprising a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; and a peptide linker linking the first and second polypeptides into a single-chain protein.

The term "multivalent" means any assemblage, covalently or non-covalently joined, of two or more single-chain proteins, the assemblage having more than one antigen-binding site. The single-chain proteins composing the

assemblage may have antigen-binding activity, or they may lack antigen-binding activity individually but be capable of assembly into active multivalent antigen-binding proteins. The term "multivalent" encompasses bivalent, trivalent, tetravalent, etc. It is envisioned that multivalent forms above bivalent may be useful for certain applications.

A preferred form of the multivalent antigen-binding protein comprises bivalent proteins, including heterobivalent and homobivalent forms. The term "bivalent" means an assemblage of single-chain proteins associated with each other to form two antigen-binding sites. The term "heterobivalent" indicates multivalent antigen-binding proteins that are bispecific molecules capable of binding to two different antigenic determinants. Therefore, heterobivalent proteins have two antigen-binding sites that have different binding specificities. The term "homobivalent" indicates that the two binding sites are for the same antigenic determinant.

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The terms "single-chain molecule" or "single-chain protein" are used interchangeably here. They are structurally defined as comprising the binding portion of a first polypeptide from the variable region of an antibody, associated with the binding portion of a second polypeptide from the variable region of an antibody, the two polypeptides being joined by a peptide linker linking the first and second polypeptides into a single polypeptide chain. The single polypeptide chain thus comprises a pair of variable regions connected by a polypeptide linker. The regions may associate to form a functional antigen-binding site, as in the case wherein the regions comprise a light-chain and a heavy-chain variable region pair with appropriately paired complementarity determining regions (CDRs). In this case, the single-chain protein is referred to as a "single-chain antigen-binding protein" or "single-chain antigen-binding molecule."

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Alternatively, the variable regions may have unnaturally paired CDRs or may both be derived from the same kind of antibody chain, either heavy or light, in which case the resulting single-chain molecule may not display a functional antigen-binding site. The single-chain antigen-binding protein

molecule is more fully described in U.S. Patent No. 4,946,778 (Ladner et al.), and incorporated herein by reference.

Without being bound by any particular theory, the inventors speculate on several models which can equally explain the phenomenon of multivalence. The inventors' models are presented herein for the purpose of illustration only, and are not to be construed as limitations upon the scope of the invention. The invention is useful and operable regardless of the precise mechanism of multivalence.

Figure 1 depicts the first hypothetical model for the creation of a multivalent protein, the "Association" model. Fig. 1A shows two monovalent single-chain antigen-binding proteins, each composed of a V_L, a V_H, and a linker polypeptide covalently bridging the two. Each monovalent single-chain antigen-binding protein is depicted having an identical antigen-binding site containing antigen. Figure 1B shows the simple association of the two single-chain antigen-binding proteins to create the bivalent form of the multivalent protein. It is hypothesized that simple hydrophobic forces between the monovalent proteins are responsible for their association in this manner. The origin of the multivalent proteins may be traceable to their concentration dependence. The monovalent units retain their original association between the V_H and V_L regions. Figure 1C shows the newly-formed homobivalent protein binding two identical antigen molecules simultaneously. Homobivalent antigen-binding proteins are necessarily monospecific for antigen.

Homovalent proteins are depicted in Figs. 2A through 2C formed according to the Association model. Fig. 1A depicts a homobivalent protein, Fig. 2B a trivalent protein, and Fig. 2C a tetravalent protein. Of course, the limitations of two-dimensional images of three-dimensional objects must be taken into account. Thus, the actual spatial arrangement of multivalent proteins can be expected to vary somewhat from these figures.

A heterobivalent antigen-binding protein has two different binding sites, the sites having different binding specificities. Figures 3A through C depict the Association model pathway to the creation of a heterobivalent protein. Figure 3A shows two monovalent single-chain antigen-binding proteins, Anti-

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A single-chain antigen-binding protein and Anti-B single-chain antigen-binding protein, with antigen types A and B occupying the respective binding sites. Figure 3B depicts the heterobivalent protein formed by the simple association of the original monovalent proteins. Figure 3C shows the heterobivalent protein having bound antigens A and B into the antigen-binding sites. Figure 3C therefore shows the heterobivalent protein binding in a bispecific manner.

An alternative model for the formation of multivalent antigen-binding proteins is shown in Figures 4 through 6. This "Rearrangement" model hypothesizes the dissociation of the variable region interface by contact with dissociating agents such as guanidine hydrochloride, urea, or alcohols such as ethanol, either alone or in combination. Combinations and relevant concentration ranges of dissociating agents are recited in the discussion concerning dissociating agents, and in Example 2. Subsequent re-association of dissociated regions allows variable region recombination differing from the starting single-chain proteins, as depicted in Fig. 4B. The homobivalent antigen-binding protein of Figure 4B is formed from the parent single-chain antigen-binding proteins shown in Figure 4A, the recombined bivalent protein having V_L and V_H from the parent monovalent single-chain proteins. The homobivalent protein of Figure 4B is a fully functional monospecific bivalent protein, shown actively binding two antigen molecules.

proteins via the Rearrangement model. Figure 5A shows a pair of single-chain proteins, each having a V_L with complementarity determining regions (CDRs) that do not match those of the associated V_H. These single-chain proteins have reduced or no ability to bind antigen because of the mixed nature of their antigen-binding sites, and thus are made specifically to be assembled into multivalent proteins through this route. Figure 5B shows the heterobivalent antigen-binding protein formed whereby the V_H and V_L regions of the parent proteins are shared between the separate halves of the heterobivalent protein. Figure 5C shows the binding of two different antigen molecules to the resultant functional bispecific heterobivalent protein. The Rearrangement model also explains the generation of multivalent proteins of

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a higher order than bivalent, as it can be appreciated that more than a pair of single-chain proteins can be reassembled in this manner. These are depicted in Figures 6A and 6B.

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One of the major utilities of the multivalent antigen-binding protein is in the heterobivalent form, in which one specificity is for one type of hapten or antigen, and the second specificity is for a second type of hapten or antigen. A multivalent molecule having two distinct binding specificities has many potential uses. For instance, one antigen binding site may be specific for a cell-surface epitope of a target cell, such as a tumor cell or other undesirable cell. The other antigen-binding site may be specific for a cell-surface epitope of an effector cell, such as the CD3 protein of a cytotoxic T-cell. In this way, the heterobivalent antigen-binding protein may guide a cytotoxic cell to a particular class of cells that are to be preferentially attacked.

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Other uses of heterobivalent antigen-binding proteins are the specific targeting and destruction of blood clots by a bispecific molecule with specificity for tissue plasminogen activator (tPA) and fibrin; the specific targeting of pro-drug activating enzymes to tumor cells by a bispecific molecule with specificity for tumor cells and enzyme; and specific targeting of cytotoxic proteins to tumor cells by a bispecific molecule with specificity for turnor cells and a cytotoxic protein. This list is illustrative only, and any use for which a multivalent specificity is appropriate comes within the scope of this invention.

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The invention also extends to uses for the multivalent antigen-binding proteins in purification and biosensors. Affinity purification is made possible by affixing the multivalent antigen-binding protein to a support, with the antigen-binding sites exposed to and in contact with the ligand molecule to be separated, and thus purified. Biosensors generate a detectable signal upon binding of a specific antigen to an antigen-binding molecule, with subsequent processing of the signal. Multivalent antigen-binding proteins, when used as the antigen-binding molecule in biosensors, may change conformation upon binding, thus generating a signal that may be detected.

Essentially all of the uses for which monoclonal or polyclonal antibodies, or fragments thereof, have been envisioned by the prior art, can be addressed by the multivalent proteins of the present invention. These uses include detectably-labelled forms of the multivalent protein. Types of labels are well-known to those of ordinary skill in the art. They include radiolabelling, chemiluminescent labeling, fluorochromic labelling, and chromophoric labeling. Other uses include imaging the internal structure of an animal (including a human) by administering an effective amount of a labelled form of the multivalent protein and measuring detectable radiation associated with the animal. They also include improved immunoassays, including sandwich immunoassay, competitive immunoassay, and other immunoassays wherein the labelled antibody can be replaced by the multivalent antigen-binding protein of this invention.

A first preferred method of producing multivalent antigen-binding proteins involves separating the multivalent proteins from a production composition that comprises both multivalent and single-chain proteins, as represented in Example 1. The method comprises producing a composition of multivalent and single-chain proteins, separating the multivalent proteins from the single-chain proteins, and recovering the multivalent proteins.

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A second preferred method of producing multivalent antigen-binding proteins comprises the steps of producing single-chain protein molecules, dissociating said single-chain molecules, reassociating the single-chain molecules such that a significant fraction of the resulting composition includes multivalent forms of the single-chain antigen-binding proteins, separating multivalent antigen-binding proteins from single-chain molecules, and recovering the multivalent proteins. This process is illustrated with more detail in Example 2. For the purposes of this method, the term "producing a composition comprising single-chain molecules" may indicate the actual production of these molecules. The term may also include procuring them from whatever commercial or institutional source makes them available. Use of the term "producing single-chain proteins" means production of single-chain proteins by any process, but preferably according to the process set forth in

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U.S. Patent No. 4,946,778 (Ladner *et al.*). Briefly, that patent pertains to a single polypeptide chain antigen-binding molecule which has binding specificity and affinity substantially similar to the binding specificity and affinity of the aggregate light and heavy chain variable regions of an antibody, to genetic sequences coding therefore, and to recombinant DNA methods of producing such molecules, and uses for such molecules. The single-chain protein produced by the Ladner *et al.* methodology comprises two regions linked by a linker polypeptide. The two regions are termed the V_H and V_L regions, each region comprising one half of a functional antigen-binding site.

The term "dissociating said single-chain molecules" means to cause the physical separation of the two variable regions of the single-chain protein without causing denaturation of the variable regions.

"Dissociating agents" are defined herein to include all agents capable of dissociating the variable regions, as defined above. In the context of this invention, the term includes the well-known agents alcohol (including ethanol), guanidine hydrochloride (GuHCl), and urea. Others will be apparent to those of ordinary skill in the art, including detergents and similar agents capable of interrupting the interactions that maintain protein conformation. In the preferred embodiment, a combination of GuHCl and ethanol (EtOH) is used as the dissociating agent. A preferred range for ethanol and GuHCl is from 0 to 50% EtOH, vol/vol, 0 to 2.0 moles per liter (M) GuHCl. A more preferred range is from 10-30% EtOH and 0.5-1.0 M GuHCl, and a most preferred range is 20% EtOH, 0.5 M GuHCl. A preferred dissociation buffer contains 0.5 M guanidine hydrochloride, 20% ethanol, 0.05 M TRIS, and 0.01 M CaCl₂, pH 8.0.

Use of the term "re-associating said single-chain molecules" is meant to describe the reassociation of the variable regions by contacting them with a buffer solution that allows reassociation. Such a buffer is preferably used in the present invention and is characterized as being composed of 0.04 M MOPS, 0.10 M calcium acetate, pH 7.5. Other buffers allowing the reassociation of the V_L and V_H regions are well within the expertise of one of ordinary skill in the art.

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The separation of the multivalent protein from the single-chain molecules occurs by use of standard techniques known in the art, particularly including cation exchange or gel filtration chromatography.

Cation exchange chromatography is the general liquid chromatographic technique of ion-exchange chromatography utilizing anion columns well-known to those of ordinary skill in the art. In this invention, the cations exchanged are the single-chain and multivalent protein molecules. Since multivalent proteins will have some multiple of the net charge of the single-chain molecule, the multivalent proteins are retained more strongly and are thus separated from the single-chain molecules. The preferred cationic exchanger of the present invention is a polyaspartic acid column, as shown in Figure 7. Figure 7 depicts the separation of single-chain protein (Peak 1, 27.32 min.) from bivalent protein (Peak 2, 55.54 min.) Those of ordinary skill in the art will realize that the invention is not limited to any particular type of chromatography column, so long as it is capable of separating the two forms of protein molecules.

Gel filtration chromatography is the use of a gel-like material to separate proteins on the basis of their molecular weight. A "gel" is a matrix of water and a polymer, such as agarose or polymerized acrylamide. The present invention encompasses the use of gel filtration HPLC (high performance liquid chromatography), as will be appreciated by one of ordinary skill in the art. Figure 8 is a chromatogram depicting the use of a Waters Associates' Protein-Pak 300 SW gel filtration column to separate monovalent single-chain protein from multivalent protein, including the monomer (21.940 min.), bivalent protein (20.135 min.), and trivalent protein (18.640 min.).

Recovering the multivalent antigen-binding proteins is accomplished by standard collection procedures well known in the chemical and biochemical arts. In the context of the present invention recovering the multivalent protein preferably comprises collection of eluate fractions containing the peak of interest from either the cation exchange column, or the gel filtration HPLC column. Manual and automated fraction collection are well-known to one of

ordinary skill in the art. Subsequent processing may involve lyophilization of the eluate to produce a stable solid, or further purification.

A third preferred method of producing multivalent antigen-binding proteins is to start with purified single-chain proteins at a lower concentration, and then increase the concentration until some significant fraction of multivalent proteins is formed. The multivalent proteins are then separated and recovered. The concentrations conducive to formation of multivalent proteins in this manner are from about 0.5 milligram per milliliter (mg/ml) to the concentration at which precipitates begin to form.

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The use of the term "substantially free" when used to describe a composition of multivalent and single-chain antigen-binding protein molecules means the lack of a significant peak corresponding to the single-chain molecule, when the composition is analyzed by cation exchange chromatography, as disclosed in Example 1 or by gel filtration chromatography as disclosed in Example 2.

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By use of the term "aqueous composition" is meant any composition of single-chain molecules and multivalent proteins including a portion of water. In the same context, the phrase "an excess of multivalent antigenbinding protein over single-chain molecules" indicates that the composition comprises more than 50% of multivalent antigen-binding protein.

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The use of the term "cross-linking" refers to chemical means by which one can produce multivalent antigen-binding proteins from monovalent single-chain protein molecules. For example, the incorporation of a cross-linkable sulfhydryl chemical group as a cysteine residue in the single-chain proteins allows cross-linking by mild reduction of the sulfhydryl group. Both monospecific and multispecific multivalent proteins can be produced from single-chain proteins by cross-linking the free cysteine groups from two or more single-chain proteins, causing a covalent chemical linkage to form between the individual proteins. Free cysteines have been engineered into the C-terminal portion of the 4-4-20/212 single-chain antigen-binding protein, as discussed in Example 5 and Example 8. These free cysteines may then be cross-linked to form multivalent antigen-binding proteins.

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The invention also comprises single-chain proteins, comprising: (a) a first polypeptide comprising the binding portion of the variable region of an antibody light chain; (b) a second polypeptide comprising the binding portion of the variable region of an antibody light chain; and (c) a peptide linker linking said first and second polypeptides (a) and (b) into said single-chain protein. A similar single-chain protein comprising the heavy chain variable regions is also a part of this invention. Genetic sequences encoding these molecules are also included in the scope of this invention. Since these proteins are comprised of two similar variable regions, they do not necessarily have any antigen-binding capability.

The invention also includes a DNA sequence encoding a bispecific bivalent antigen-binding protein. Example 4 and Example 7 discusses in detail the sequences that appear in Figs. 10A and 10B that allow one of ordinary skill to construct a heterobivalent antigen-binding molecule. Figure 10A is an amino acid and nucleotide sequence listing of the single-chain protein comprising the 4-4-20 V_L region connected through the 212 linker polypeptide to the CC49 V_H region. Figure 10B is a similar listing of the single-chain protein comprising the CC49 V_L region connected through the 212 linker polypeptide to the 4-4-20 V_H region. Subjecting a composition including these single-chain molecules to dissociating and subsequent re-associating conditions results in the production of a bivalent protein with two different binding specificities.

Synthesis of DNA sequences is well know in the art, and possible through at least two routes. First, it is well-known that DNA sequences may be synthesized through the use of automated DNA synthesizers *de novo*, once the primary sequence information is known. Alternatively, it is possible to obtain a DNA sequence coding for a multivalent single-chain antigen-binding protein by removing the stop codons from the end of a gene encoding a single-chain antigen-binding protein, and then inserting a linker and a gene encoding a second single-chain antigen-binding protein. Example 6 demonstrates the construction of a DNA sequence coding for a bivalent single-chain antigen-binding protein. Other methods of genetically constructing multivalent single-

chain antigen-binding proteins come within the spirit and scope of the present invention.

Having now generally described this invention the same will better be understood by reference to certain specific examples which are included for purposes of illustration and are not intended to limit it unless otherwise specified.

Example 1

Production of Multivalent Antigen-Binding Proteins During Purification

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In the production of multivalent antigen-binding proteins, the same recombinant E. coli production system that was used for prior single-chain antigen-binding protein production was used. See Bird, et al., Science 242:423 (1988). This production system produced between 2 and 20% of the total E. coli protein as antigen-binding protein. For protein recovery, the frozen cell paste from three 10-liter fermentations (600-900 g) was thawed overnight at 4°C and gently resuspended at 4°C in 50 mM Tris-Hcl, 1.0 mM EDTA, 100 mM KCl, 0.1 mM PMSF, pH 8.0 (lysis buffer), using 10 liters of lysis buffer for every kilogram of wet cell paste. When thoroughly resuspended, the chilled mixture was passed three times through a Manton-Gaulin cell homogenizer to totally lyse the cells. Because the cell homogenizer raised the temperature of the cell lysate to 25 \pm 5°C, the cell lysate was cooled to $5\pm$ 2°C with a Lauda/Brinkman chilling coil after each pass. Complete lysis was verified by visual inspection under a microscope.

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The cell lysate was centrifuged at 24,300g for 30 min. at 6°C using a Sorvall RC-5B centrifuge. The pellet containing the insoluble antigen-binding protein was retained, and the supernatant was discarded. The pellet was washed by gently scraping it from the centrifuge bottles and resuspending it in 5 liters of lysis buffer/kg of wet cell paste. The resulting 3.0- to 4.5-liter suspension was again centrifuged at 24,300g for 30 min at 6°C, and the

supernatant was discarded. This washing of the pellet removes soluble E. coli proteins and can be repeated as many as five times. At any time during this washing procedure the material can be stored as a frozen pellet at -20°C. A substantial time saving in the washing steps can be accomplished by utilizing a Pellicon tangential flow apparatus equipped with 0.22- μ m microporous filters, in place of centrifugation.

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The washed pellet was solubilized at 4°C in freshly prepared 6 M guanidine hydrochloride, 50 mM Tris-HCl, 10 mM CaCl₂, 50 mM KCl, pH 8.0 (dissociating buffer), using 9 ml/g of pellet. If necessary, a few quick pulses from a Heat Systems Ultrasonics tissue homogenizer can be used to complete the solubilization. The resulting suspension was centrifuged at 24,300g for 45 min at 6°C and the pellet was discarded. The optical density of the supernatant was determined at 280 nm and if the OD₂₈₀ was above 30, additional dissociating buffer was added to obtain an OD₂₈₀ of approximately 25.

The supernatant was slowly diluted into cold (4-7°C) refolding buffer (50 mM Tris-HCl, 10 mM CaCl₂, 50 mM KCl, pH 8.0) until a 1:10 dilution was reached (final volume 10-20 liters). Re-folding occurs over approximately eighteen hours under these conditions. The best results are obtained when the GuHCl extract is slowly added to the refolding buffer over a 2-h period, with gentle mixing. The solution was left undisturbed for at least a 20-h period, and 95% ethanol was added to this solution such that the final ethanol concentration was approximately 20%. This solution was left undisturbed until the flocculated material settled to the bottom, usually not less than sixty minutes. The solution was filtered through a 0.2 um Millipore Millipak 200. This filtration step may be optionally preceded by a centrifugation step. The filtrate was concentrated to 1 to 2 liters using an Amicon spiral cartridge with a 10,000 MWCO cartridge, again at 4°C.

The concentrated crude antigen-binding protein sample was dialyzed against Buffer A (60 mM MOPS, 0.5 mM Ca acetate, pH 6.0-6.4) until the conductivity was lowered to that of Buffer A. The sample was then loaded on a 21.5 x 250-mm polyaspartic acid PolyCAT A column, manufactured by Poly

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LC of Columbia, Maryland. If more than 60 mg of protein is loaded on this column, the resolution begins to deteriorate; thus, the concentrated crude sample often must be divided into several PolyCAT A runs. Most antigenbinding proteins have an extinction coefficient of about 2.0 ml mg⁻¹ cm⁻¹ at 280 nm and this can be used to determine protein concentration. The antigenbinding protein sample was eluted from the PolyCAT A column with a 50-min linear gradient from Buffer A to Buffer B (see Table 1). Most of the single-chain proteins elute between 20 and 26 minutes when this gradient is used. This corresponds to an eluting solvent composition of approximately 70% Buffer A and 30% Buffer B. Most of the bivalent antigen-binding proteins elute later than 45 minutes, which correspond to over 90% Buffer B.

Figure 7 is a chromatogram depicting the separation of single-chain protein from bivalent CC49/212 protein, using the cation-exchange method just described. Peak 1, 27.32 minutes, represents the monomeric single-chain fraction. Peak 2, 55.52 minutes, represents the bivalent protein fraction.

Figure 8 is a chromatogram of the purified monomeric single-chain antigen-binding protein CC49/212 (Fraction 7 from Fig. 7) run on a Waters Protein-Pak 300SW gel filtration column. Monomer, with minor contaminates of dimer and trimer, is shown. Figure 9 is a chromatogram of the purified bivalent antigen-binding protein CC49/212 (Fraction 15 from Fig. 7) run on the same Waters Protein-Pak 300SW gel filtration column as used in Fig. 8.

	TABLE 1	-						
PolyCAT A Cation-Exchange HPLC Gradients								
	Buffers ^b							
Flow (ml/min)	Α	В	С					
15.0	100	0	0					
15.0	0:	100	0					
15.0	0	100	0					
15.0	0	0	100					
15.0	0	0	100					
15.0	100	0	0					
15.0	100	0	0					
	Flow (ml/min) 15.0 15.0 15.0 15.0 15.0 15.0	Flow (ml/min) A 15.0 100 15.0 0 15.0 0 15.0 0 15.0 0 15.0 0 15.0 100	PolyCAT A Cation-Exchange HPLC Gradients Buffersb					

^{*}Linear gradients are run between each time point.

^bBuffer A, 60 mM MOPS, 0.5 mM Ca acetate, pH 6.0-6.4; Buffer B, 60 mM MOPS, 20mM Ca acetate, pH 7.5-8.0; Buffer C, 40 mM MOPS, 100 mM CaCl₂, pH 7.5.

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This purification procedure yielded multivalent antigen-binding proteins that are more than 95% pure as examined by SDS-PAGE and size exclusion HPLC. Modifications of the above procedure may be dictated by the isoelectric point of the particular multivalent antigen-binding protein being purified. Of the monomeric single-chain proteins that have been purified to date, all have had an isoelectric point (pI) between 8.0 and 9.5. However, it is possible that a multivalent antigen-binding protein may be produced with a pI of less than 7.0. In that case, an anion exchange column may be required for purification.

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The CC49 monoclonal antibody was developed by Dr. Jeffrey Schlom's group, Laboratory of Tumor Immunology and Biology, National Cancer Institute. It binds specifically to the pan-carcinoma tumor antigen TAG-72. See Muraro, R. et al., Cancer Research 48:4588-4596 (1988).

To determine the binding properties of the bivalent and monomeric CC49/212 antigen-binding proteins, a competition radioimmunoassay (RIA)

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was set up in which a CC49 IgG (with two antigen binding sites) radiolabeled with ¹²⁵I was competed against unlabeled CC49 IgG, or monovalent (fraction 7 in Figure 7) or bivalent (fraction 15 in Figure 7) CC49/212 antigen-binding protein for binding to the TAG-72 antigen on a human breast carcinoma extract. (See Figure 18). This competition RIA showed that the bivalent antigen-binding protein competed equally well for the antigen as did IgG, whereas the monovalent single-chain antigen-binding protein needed a ten-fold higher protein concentration to displace the IgG. Thus, the monovalent antigen-binding protein competes with about a ten-fold lower affinity for the antigen than does the bivalent IgG or bivalent antigen-binding protein. Figure 18 also shows the result of the competition RIA of a non-TAG-72 specific single-chain antigen-binding protein, the antifluorescein 4-4-20/212, which does not compete for binding.

Example 2

Process of Making Multivalent Antigen-Binding Proteins Using Dissociating Agents

A. Process Using Guanidine HCl and Ethanol

Multivalent antigen-binding proteins were produced from purified single-chain proteins in the following way. First the purified single-chain protein at a concentration of 0.25-4 mg/ml was dialyzed against 0.5 moles/liter (M) guanidine hydrochloride (GuHCl), 20% ethanol (EtOH), in 0.05 M TRIS, 0.05 M KCl, 0.01 M CaCl₂ buffer pH 8.0. This combination of dissociating agents is thought to disrupt the V_L/V_H interface, allowing the V_H of a first single-chain molecule to come into contact with a V_L from a second single-chain molecule. Other dissociating agents such as urea, and alcohols such as isopropanol or methanol should be substitutable for GuHCl and EtOH. Following the initial dialysis, the protein was dialyzed against the load buffer for the final HPLC purification step. Two separate purification protocols,

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cation exchange and gel filtration chromatography, can be used to separate the single-chain protein monomer from the multivalent antigen-binding proteins. In the first method, monomeric and multivalent antigen-binding proteins were separated by using cation exchange HPLC chromography, using a polyaspartate column (PolyCAT A). This was a similar procedure to that used in the final purification of the antigen-binding proteins as described in Example 1. The load buffer was 0.06 M MOPS, 0.001 M Calcium Acetate pH 6.4. In the second method, the monomeric and multivalent antigen-binding proteins were separated by gel filtration HPLC chromatography using as a load buffer 0.04 M MOPS, 0.10 M Calcium Acetate pH 7.5. Gel filtration chromatography separates proteins based on their molecular size.

Once the antigen-binding protein sample was loaded on the cation exchange HPLC column, a linear gradient was run between the load buffer (0.04 to 0.06 M MOPS, 0.000 to 0.001 M calcium acetate, 0 to 10% glycerol pH 6.0-6.4) and a second buffer (0.04 to 0.06 M MOPS, 0.01 to 0.02 M calcium acetate, 0 to 10% glycerol pH 7.5). It was important to have extensively dialyze the antigen-binding protein sample before loading it on the column. Normally, the conductivity of the sample is monitored against the dialysis buffer. Dialysis is continued until the conductivity drops below 600 μ S. Figure 11 shows the separation of the monomeric (27.83 min.) and bivalent (50.47 min.) forms of the CC49/212 antigen-binding protein by cation The chromatographic conditions for this separation were as exchange. follows: PolyCAT A column, 200 x 4.6mm, operated at 0.62 ml/min.; load buffer and second buffer as in Example 1; gradient program from 100 percent load buffer A to 0 percent load buffer A over 48 mins; sample was CC49/212, 1.66 mg/ml; injection volume 0.2 ml. Fractions were collected from the two peaks from a similar chromatogram and identified as monomeric and bivalent proteins using gel filtration HPLC chromatography as described below.

Gel filtration HPLC chromatography (TSK G2000SW column from Toyo Soda, Tokyo, Japan) was used to identify and separate monomeric single-chain and multivalent antigen-binding proteins. This procedure has been described by Fukano, et al., J. Chromatography 166:47 (1978).

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Multimerization (creation of multivalent protein from monomeric single-chain protein) was by treatment with 0.5 M GuHCl and 20% EtOH for the times indicated in Table 2A followed by dialysis into the chromatography buffer. Figure 12 shows the separation of monomeric (17.65 min.), bivalent (15.79 min.), trivalent (14.19 min.), and higher oligomers (shoulder at about 13.09 min.) of the B6.2/212 antigen-binding protein. The B6.2/212 single-chain antigen-binding protein is described in Colcher, D., et al., J. Nat. Cancer Inst. 82:1191-1197 (1990)). This separation depicts the results of a 24-hour multimerization treatment of a 1.0 mg/ml B6.2/212 antigen-binding protein sample. The HPLC buffer used was 0.04 M MOPS, 0.10 M calcium acetate, 0.04% sodium azide, pH 7.5.

Figure 13 shows the results of a 24-hour treatment of a 4.0 mg/ml CC49/212 antigen-binding protein sample, generating monomeric, bivalent and trivalent proteins at 16.91, 14.9, and 13.42 min., respectively. The HPLC buffer was 40 mM MOPS, 100 mM calcium acetate, pH 7.35. Multimerization treatment was for the times indicated in Table 2.

The results of Example 2A are shown in Table 2A. Table 2A shows the percentage of bivalent and other multivalent forms before and after treatment with 20% ethanol and 0.5M GuHCl. Unless otherwise indicated, percentages were determined using a automatic data integration software package.

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Table 2A
Summary of the generation of bivalent and higher multivalent forms of B6.2/212 and CC49/212 proteins using guanidine hydrochloride and ethanol

	Time	Concentration		%		
protein	(hours)	(mg/ml)	monomer	dimer	trimer	multimers
CC49/212	0	0.25	86.7	11.6	1.7	0.0
	0	1.0 ²	84.0	10.6	5.5	0.0
	0	4.0	70.0	17.1	12.91	0.0
	2	0.252	62.9	33.2	4.2	0.0
	2	1.0	24.2	70.6	5.1	0.0
	2	4.0	9.3	81.3	9.5	0.0
	26	0.25	16.0	77.6	6.4	0.0
	26	1.0	9.2	82.8	7.9	0.0
	26	4.0	3.7	78.2	18.1	0.0
B6.2/212	0	0.25	100.0	0.0	0.0	0.0
	0	1.0	100.0	0.0	0.0	0.0
	0	4.0	100.0	0.0	0.0	0.0
	2	0.25 ²	98.1	1.9	0.0	0:0
	2	1.0	100.0	0.0	0.0	0.0
	2	4.0	90.0	5.5	1.0	0.0
	24	0.25	45.6	37.5	10.2	6.7
	24	1.0	50.8	21.4	12.3	15.0
	24	4.0	5 . 9	37.2	25.7	29.9

Based on cut out peaks that were weighted.

² Average of two experiments.

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B. Process Using Urea and Ethanol

Multivalent antigen-binding proteins were produced from purified single-chain proteins in the following way. First the purified single-chain protein at a concentration of 0.25-1 mg/ml was dialyzed against 2M urea, 20% ethanol (EtOH), and 50mM Tris buffer pH 8.0, for the times indicated in Table 2B. This combination of dissociating agents is thought to disrupt the V_L/V_H interface, allowing the V_H of a first single-chain molecule to come into contact with a V_L from a second single-chain molecule. Other dissociating agents such as isopropanol or methanol should be substitutable for EtOH.

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Following the initial dialysis, the protein was dialyzed against the load buffer for the final HPLC purification step.

Gel filtration HPLC chromatography (TSK G2000SW column from Toyo Soda, Tokyo, Japan) was used to identify and separate monomeric single-chain and multivalent antigen-binding proteins. This procedure has been described by Fukano, et al., J. Chromatography 166:47 (1978).

The results of Example 2B are shown in Table 2B. Table 2B shows the percentage of bivalent and other multivalent forms before and after treatment with 20% ethanol and urea. Percentages were determined using an automatic data integration software package.

Table 2B

Summary of the generation of bivalent and higher multivalent forms of B6.2/212 and CC49/212 proteins using urea and ethanol

protein	Time (hours)	Concentration (mg/ml)	monomer	% dimer	trimer	multimers
B6.2	. 0	0.25	44.1	37.6	15.9	2.4
	0	1.0	37.7	33.7	19.4	9.4
	3	0.25	22.2	66.5	11.3	0.0
	3	1.0	13.7	69.9	16.4	0.0

Example 3

Determination of Binding Constants

Three anti-fluorescein single-chain antigen-binding proteins have been constructed based on the anti-fluorescein monoclonal antibody 4-4-20. The three 4-4-20 single-chain antigen-binding proteins differ in the polypeptide linker connecting the V_H and V_L regions of the protein. The three linkers used were 202', 212 and 216 (see Table 3). Bivalent and higher forms of the 4-4-20 antigen-binding protein were produced by concentrating the purified monomeric single-chain antigen-binding protein in the cation exchange load buffer (0.06 M MOPS, 0.001 M calcium acetate pH 6.4) to 5 mg/ml. The

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bivalent and monomeric forms of the 4-4-20 antigen-binding proteins were separated by cation exchange HPLC (polyaspartate column) using a 50 min. linear gradient between the load buffer (0.06 M MOPS, 0.001 M calcium acetate pH 6.4) and a second buffer (0.06 M MOPS, 0.02 M calcium acetate pH 7.5). Two 0.02 ml samples were separated, and fractions of the bivalent and monomeric protein peaks were collected on each run. The amount of protein contained in each fraction was determined from the absorbance at 278 nm from the first separation. Before collecting the fractions from the second separation run, each fraction tube had a sufficient quantity of 1.03 x 10⁵ M fluorescein added to it, such that after the fractions were collected a 1-to-1 molar ratio of protein-to-fluorescein existed. Addition of fluorescein stabilized the bivalent form of the 4-4-20 antigen-binding proteins. These samples were kept at 2°C (on ice).

The fluorescein dissociation rates were determined for each of these samples following the procedures described by Herron, J.N., in *Fluorescence Hapten: An Immunological Probe*, E.W. Voss, Ed., CRC Press, Boca Raton, FL (1984). A sample was first diluted with 20 mM HEPES buffer pH 8.0 to 5.0×10^8 M 4-4-20 antigen-binding protein. $560 \mu l$ of the 5.0×10^8 M 4-4-20 antigen-binding protein sample was added to a cuvette in a fluorescence spectrophotometer equilibrated at 2°C and the fluorescence was read. $140 \mu l$ of 1.02×10^{-5} M fluoresceinamine was added to the cuvette, and the fluorescence was read every 1 minute for up to 25 minutes (see Table 4).

The binding constants (K₂) for the 4-4-20 single-chain antigen-binding protein monomers diluted in 20 mM HEPES buffer pH 8.0 in the absence of fluorescein were also determined (see Table 4).

The three polypeptide linkers in these experiments differ in length. The 202', 212 and 216 linkers are 12, 14 and 18 residues long, respectively. These experiments show that there are two effects of linker length on the 4-4-20 antigen-binding proteins: first, the shorter the linker length the higher the fraction of bivalent protein formed; second, the fluorescein dissociation rates of the monomeric single-chain antigen-binding proteins are effected more by the linker length than are the dissociation rates of the bivalent antigen-binding

proteins. With the shorter linkers 202' and 212, the bivalent antigen-binding proteins have slower dissociation rates than the monomers. Thus, the linkers providing optimum production and binding affinities for monomeric and bivalent antigen-binding proteins may be different. Longer linkers may be more suitable for monomeric single-chain antigen-binding proteins, and shorter linkers may be more suitable for multivalent antigen-binding proteins.

Table 3 Linker Designs							
-KLEIE	GKSSGSGSESKS'	TQKLD-	202'	Bird et al.			
-KLEIK	GSTSGSGKSSEGKG ²	EVKLD-	212	Bedzyk et al.			
-KLEIK	GSTSGSGKSSEGSGSTKG ³	EVKLD-	216	This application			
-KLVLK	GSTSGKPSEGKG ⁴	EVKLD-	217	This application			

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- (1) SEQ ID NO. 1
- (2) SEQ ID NO. 2
- (3) SEQ ID NO. 3
- (4) SEQ ID NO. 4

	Table 4						
Effects of Linke	ers on the SCA Prote	ein Monomers and	Dimers				
	Linker						
202' 212 216							
Monomer Fraction Ka Dissociation rate	0.47 0.5 x 10 ⁹ M ⁻¹ 8.2 x 10 ⁻³ s ⁻¹	0.66 1.0 x 10 ⁹ M ⁻¹ 4.9 x 10 ⁻³ s ⁻¹	0.90 1.3 x 10 ⁹ M ⁻¹ 3.3 x 10 ⁻³ s ⁻¹				
Dimer Fraction Dissociation rate	0.53 4.6 x 10 ⁻³ s ⁻¹	0.34 3.5 x 10 ⁻³ s ⁻¹	0.10 3.5 x 10 ⁻³ s ⁻¹				
Monomer/Dimer Dissociation rate ratio	1.8	1.4	0.9				

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Genetic Construction of a Mixed-Fragment Bivalent Antigen-Binding Protein

The genetic constructions for one particular heterobivalent antigen-binding protein according to the Rearrangement model are shown in Figures 10A and 10B. Figure 10A is an amino acid and nucleotide sequence listing of the 4-4-20 V_L/212/CC49 V_H construct, coding for a single-chain protein with a 4-4-20 V_L, linked via a 212 polypeptide linker to a CC49 V_H. Figure 10B is a similar listing showing the CC49 V_L/212/4-4-20 V_H construct, coding for a single-chain protein with a CC49 V_L, linked via a 212 linker to a 4-4-20 V_H. These single-chain proteins may recombine according to the Rearrangement model to generate a heterobivalent protein comprising a CC49 antigen-binding site linked to a 4-4-20 antigen-binding site, as shown in Figure 5B.

"4-4-20 V_L" means the variable region of the light chain of the 4-4-20 mouse monoclonal antibody (Bird, R.E. et al., Science 242:423 (1988)). The number "212" refers to a specific 14-residue polypeptide linker that links the 4-4-20 V_L and the CC49 V_H. See Bedzyk, W.D. et al., J. Biol. Chem. 265:18615-18620 (1990). "CC49 V_H" is the variable region of the heavy chain of the CC49 antibody, which binds to the TAG-72 antigen. The CC49 antibody was developed at The National Institutes of Health by Schlom, et al. Generation and Characterization of B72.3 Second Generation Monoclonal Antibodies Reactive With The Tumor-associated Glycoprotein 72 Antigen, Cancer Research 48:4588-4596 (1988).

Insertion of the sequences shown in FIGS. 10A and 10B, by standard recombinant DNA methodology, into a suitable plasmid vector will enable one of ordinary skill in the art to transform a suitable host for subsequent expression of the single-chain proteins. See Maniatis et al., Molecular Cloning, A Laboratory Manual, p. 104, Cold Spring Harbor Laboratory (1982), for general recombinant techniques for accomplishing the aforesaid goals; see also U.S. Patent 4,946,778 (Ladner et al.) for a complete

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description of methods of producing single-chain protein molecules by recombinant DNA technology.

To produce multivalent antigen-binding proteins from the two single-chain proteins, 4-4-20V_L-212/CC49V_H and CC49V_L/212/4-4-20V_H, the two single-chain proteins are dialyzed into 0.5 M GuHCl/20% EtOH being combined in a single solution either before or after dialysis. The multivalent proteins are then produced and separated as described in Example 2.

Example 5

Preparation of Multivalent Antigen-Binding Proteins by Chemical Cross-Linking

Free cysteines were engineered into the C-terminal of the 4-4-20/212 single-chain antigen-binding protein, in order to chemically crosslink the protein. The design was based on the hinge region found in antibodies between the C_H1 and C_H2 regions. In order to try to reduce antigenicity in humans, the hinge sequence of the most common IgG class, IgG1, was chosen. The 4-4-20 Fab structure was examined and it was determined that the C-terminal sequence GluH216-ProH217-ArgH218, was part of the C_H1 region and that the hinge between C_H1 and C_H2 starts with ArgH218 or GlyH219 in the mouse 4-4-20 IgG2A antibody. Figure 14 shows the structure of a human IgG. The hinge region is indicated generally. Thus the hinge from human IgG1 would start with LysH218 or SerH219. (See Table 5).

The C-terminal residue in most of the single-chain antigen-binding proteins described to date is the amino acid serine. In the design for the hinge region, the C-terminal serine in the 4-4-20/212 single-chain antigen-binding protein was made the first serine of the hinge and the second residue of the hinge was changed from a cysteine to a serine. This hinge cysteine normally forms a disulfide bridge to the C-terminal cysteine in the light chain.

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TABLE 5

5	'SCA+3'	A - -	E - -	P V V V	K T T	G V V V	s s	c	D D	K K	T T	Н	T T	C	P	P	С	L -	С	-	
10	(1) SEO ID NO. 5		, 1																		

10 (1) SEQ ID NO. 5 (2) SEQ ID NO. 6 (3) SEQ ID NO. 7 (4) SEQ ID NO. 8

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(5) SEQ ID NO. 9

There are possible advantages to having two C-terminal cysteines, for they might form an intramolecular disulfide bond, making the protein recovery easier by protecting the sulfurs from oxidation. The hinge regions were added by introduction of a BstE II restriction site in the 3'-terminus of the gene encoding the 4-4-20/212 single-chain antigen-binding protein (see Figures 15A-15B).

The monomeric single-chain antigen-binding protein containing the Cterminal cysteine can be purified using the normal methods of purifying a single-chain antigen-binding proteins, with minor modifications to protect the free sulfhydryls. The cross-linking could be accomplished in one of two ways. First, the purified single-chain antigen-binding protein could be treated with a mild reducing agent, such as dithiothreitol, then allowed to air oxidize to form a disulfide-bond between the individual single-chain antigen-binding This type of chemistry has been successful in producing proteins. heterodimers from whole antibodies (Nisonoff et al., Quantitative Estimation of the Hybridization of Rabbit Antibodies, Nature 4826:355-359 (1962); Brennan et al., Preparation of Bispecific Antibodies by Chemical Recombination of Monoclonal Immunoglobulin G₁ Fragments, Science 229:81-83 (1985)). Second, chemical crosslinking agents such as bismaleimidehexane could be used to cross-link two single-chain antigen-binding proteins by their C-terminal cysteines. See Partis et al., J. Prot. Chem. 2:263-277 (1983).

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Example 6

Genetic Construction of Bivalent Antigen-Binding Proteins

Bivalent antigen-binding proteins can be constructed genetically and subsequently expressed in E. coli or other known expression systems. This can be accomplished by genetically removing the stop codons at the end of a gene encoding a monomeric single-chain antigen-binding protein and inserting a linker and a gene encoding a second single-chain antigen-binding protein. We have constructed a gene for a bivalent CC49/212 antigen-binding protein in this manner (see Figure 16). The CC49/212 gene in the starting expression plasmid is in an Aat II to Bam H1 restriction fragment (see Bird et al., Single-Chain Antigen-Binding Proteins, Science 242:423-426 (1988); and Whitlow et al., Single-Chain Fv Proteins and Their Fusion Proteins, Methods 2:97-105 (1991)). The two stop codons and the Bam H1 site at the C-terminal end of the CC49/212 antigen-binding protein gene were replaced by a single residue linker (Ser) and an Aat II restriction site. The resulting plasmid was cut with Aat II and the purified Aat II to Aat II restriction fragment was ligated into Aat II cut CC49/212 single-chain antigen-binding protein expression plasmid. The resulting bivalent CC49/212 single-chain antigen-binding protein expression plasmid was transfected into an E. coli expression host that contained the gene for the cI857 temperature-sensitive repressor. Expression of single-chain antigen-binding protein in this system is induced by raising the temperature from 30°C to 42°C. Fig. 17 shows the expression of the divalent CC49/212 single-chain antigen-binding protein of Fig. 16 at 42°C, on an SDS-PAGE gel containing total E. coli protein. Lane 1 contains the molecular weight standards. Lane 2 is the uninduced E. coli production strain grown at 30°C. Lane 3 is divalent CC49/212 single-chain antigen-binding protein induced by growth at 42°C. The arrow shows the band of expressed divalent CC49/212 single-chain antigen-binding protein.

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Example 7

Construction, Purification, and Testing of 4-4-20/CC49 Heterodimer F_v With 217 Linkers.

The goals of this experiment were to produce, purify and analyze for activity a new heterodimer Fv that would bind to both fluorescein and the pancarcinoma antigen TAG-72. The design consisted of two polypeptide chains, which associated to form the active heterodimer Fv. Each polypeptide chain can be described as a mixed single-chain Fv (mixed sFv). The first mixed sFv (GX 8952) comprised a 4-4-20 variable light chain (V_L) and a CC-49 variable heavy chain (V_H) connected by a 217 polypeptide linker (Figure 19A). The second mixed sFv (GX 8953) comprised a CC-49 V_L and a 4-4-20 V_H connected by a 217 polypeptide linker (Figure 19B). The sequence of the 217 polypeptide linker is shown in Table 3. Construction of analogous CC49/4-4-20 heterodimers connected by a 212 polypeptide linker as described in Example 4.

Results

A. Purification

One 10-liter fermentation of each mixed sFv was grown on casein digest-glucose-salts medium at 32°C to an optical density at 600 nm of 15 to 20. The mixed sFv expression was induced by raising the temperature of the fermentation to 42°C for one hour. 277gm (wet cell weight) of *E. coli* strain GX 8952 and 233gm (wet cell weight) of *E. coli* strain GX 8953 were harvested in a centrifuge at 7000g for 10 minutes. The cell pellets were kept and the supernate discarded. The cell pellets were frozen at -20°0C for storage.

2.55 liters of "lysis/wash buffer" (50mM Tris/ 200mM NaCl/ 1 mM EDTA, pH 8.0) was added to both of the mixed sFv's cell pellets, which were previously thawed and combined to give 510gm of total wet cell weight. After complete suspension of the cells they were then passed through a Gaulin homogenizer at 9000psi and 4°C. After this first pass the temperature increased to 23°C. The temperature was immediately brought down to 0°C using dry ice and methanol. The cell suspension was passed through the Gaulin homogenizer a second time and centrifuged at 8000 rpm with a Dupont GS-3 rotor for 60 minutes. The supernatant was discarded after centrifugation and the pellets resuspended in 2.5 liters of "lysis/wash buffer" at 4°C. This suspension was centrifuged for 45 minutes at 8000 rpm with the Dupont GS-3 rotor. The supernatant was again discarded and the pellet weighed. The pellet weight was 136.1 gm.

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1300ml of 6M Guanidine Hydrochloride/50mM Tris/50mM KCl/10mM CaCl₂pH 8.0 at 4°C was added to the washed pellet. An overhead mixer was used to speed solubilization. After one hour of mixing, the heterodimer GuHCl extract was centrifuged for 45 minutes at 8000 rpm and the pellet was discarded. The 1425ml of heterodimer Fv 6M GuHCl extract was slowly added (16 ml/min) to 14.1 liters of "Refold Buffer" (50mM Tris/50mM KCl/10mM CaCl₂, pH 8.0) under constant mixing at 4°C to give an approximate dilution of 1:10. Refolding took place overnight at 4°C.

After 17 hours of refolding the anti-fluorescein activity was checked by a 40% quenching assay, and the amount of active protein calculated. 150mg total active heterodimer Fv was found by the 40% quench assay, assuming a 54,000 molecular weight.

4 liters of prechilled (4°C) 190 proof ethanol was added to the 15 liters of refolded heterodimer with mixing for 3 hours. The mixture sat overnight at 4°C. A flocculent precipitate had settled to the bottom after this overnight treatment. The nearly clear solution was filtered through a Millipak-200 (0.22μ) filter so as to not disturb the precipitate. A 40% quench assay showed that 10% of the anti-fluorescein activity was recovered in the filtrate.

The filtered sample of heterodimer was dialyzed, using a Pellicon system containing 10,000 dalton MWCO membranes, with "dialysis buffer" 40mM MOPS/0.5mM Calcium Acetate (CaAc), pH 6.4 at 4°C. 20 liters of dialysis buffer was required before the conductivity of the retentate was equal to that of the dialysis buffer ($\sim 500 \mu S$). After dialysis the heterodimer sample was filtered through a Millipak-20 filter, O.22 μ . After this step a 40% quench assay showed there was 8.8 mg of active protein.

The crude heterodimer sample was loaded on a Poly CAT A cation exchange column at 20ml/min. The column was previously equilibrated with 60mM MOPS, 1 mM CaAc pH 6.4, at 4°C, (Buffer A). After loading, the column was washed with 150ml of "Buffer A" at 15ml/min. A 50min linear gradient was performed at 15ml/min using "Buffer A" and "Buffer B" (60mM MOPS, 20mM CaAc pH 7.5 at 4°C). The gradient conditions are presented in Table 6. "Buffer C" comprises 60mM MOPS, 100mM CaCl₂, pH 7.5.

		Table 6		
Time	%A	%B	%C	Flow
0:00	100.0	0.0	0.0	15ml/min
50:00	0.0	100.0	0.0	15ml/min
52:00	0.0	100.0	0.0	15ml/min
54:00	0.0	0.0	100.0	15ml/min
58:00	0.0	0.0	100.0	15ml/min
60:00	100.0	0.0	0.0	15ml/min

Approximately 50ml fractions were collected and analyzed for activity, purity, and molecular weight by size-exclusion chromatography. The fractions were not collected by peaks, so contamination between peaks is likely. Fractions 3 through 7 were pooled (total volume - 218ml), concentrated to 50ml and dialyzed against 4 liters of 60mM MOPS, 0.5mM CaAc pH 6.4 at 4°C overnight. The dialyzed pool was filtered through a 0.22μ filter and

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checked for absorbance at 280nm. The filtrate was loaded onto the PolyCAT A column, equilibrated with 60mM MOPS, 1 mM CaAc pH 6.4 at 4°C, at a flow rate of 10ml/min. Buffer B was changed to 60mM MOPS, 10mM CaAc pH 7.5 at 4°C. The gradient was run as in Table 6. The fractions were collected by peak and analyzed for activity, purity, and molecular weight. The chromatogram is shown in Figure 20. Fraction identification and analysis is presented in Table 7.

		Table 7											
Fraction Analysis of the Heterodimer Fv protein													
Fraction No.	A ₂₈₀ reading	Total Volume (ml)	HPLC-SE Elution Time (min)										
2	0.161	36	20.525										
3	0.067	40											
4	0.033	40											
5	0.178	45	19.133										
6	0.234	50	19.163										
7	0.069	50											
8	0.055	40											

Fractions 2 to 7 and the starting material were analyzed by SDS gel electrophoresis, 4-20%. A picture and description of the gel is presented in Figure 21.

B. HPLC Size Exclusion Results

Fractions 2, 5, and 6 correspond to the three main peaks in Figure 20 and therefore were chosen to be analyzed by HPLC size exclusion. Fraction 2 corresponds to the peak that runs at 21.775 minutes in the preparative purification (Figure 20), and runs on the HPLC sizing column at 20.525 minutes, which is in the monomeric position (Figure 22A). Fractions 5 and 6 (30.1 and 33.455 minutes, respectively, in Figure 20) run on the HPLC sizing column (Figures 22B and 22C) at 19.133 and 19.163 minutes,

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respectively (see Table 7). Therefore, both of these peaks could be considered dimers. 40% Quenching assays were performed on all fractions of this purification. Only fraction 5 gave significant activity. 2.4 mg of active CC49 4-4-20 heterodimer Fv was recovered in fraction 5, based on the Scatchard analysis described below.

C. N-terminal sequencing of the fractions

The active heterodimer Fv fraction should contain both polypeptide chains. N-terminal sequence analysis showed that fractions 5 and 6 displayed N-terminal sequences consistent with the prescence of both CC49 and 4-4-20 polypeptides and fraction 2 displayed a single sequence corresponding to the CC49/212/4-4-20 polypeptide only. We believe that fraction 6 was contaminated by fraction 5 (see Figure 20), since only fraction 5 had significant activity.

D. Anti-fluorescein activity by Scatchard analysis

The fluorescein association constants (Ka) were determined for fractions 5 and 6 using the fluorescence quenching assay described by Herron, J.N., in Fluorescence Hapten: An Immunological Probe, E.W. Voss, ed., Each sample was diluted to CRC Press, Boca Raton, FL (1984). approximately 5.0 x 10^8 M with 20 mM HEPES buffer pH 8.0. 590 μ l of the 5.0 x 10-8 M sample was added to a cuvette in a fluorescence spectrophotometer equilibrated at room temperature. In a second cuvette 590 μl of 20 mM HEPES buffer pH 8.0 was added. To each cuvette was added 10 μ l of 3.0 x 10⁻⁷ M fluorescein in 20 mM HEPES buffer pH 8.0, and the fluorescence recorded. This is repeated until 140 μ l of fluorescein had been The resulting Scatchard analysis for fraction 5 shows a binding constant of 1.16 x 109 M-1 for fraction #5 (see Figure 23). This is very close to the 4-4-20/212 sFv constant of 1.1 x 109 M⁻¹ (see Pantoliano et al., Biochemistry 30:10117-10125 (1991)). The R intercept on the Scatchard analysis represents the fraction of active material. For fraction 5, 61% of the material was active. The graph of the Scatchard analysis on fraction 6 shows a binding constant of $3.3 \times 10^8 \text{ M}^{-1}$ and 14% active. The activity that is present in fraction 6 is most likely contaminants from fraction 5.

E. Anti-TAG-72 activity by competition ELISA

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The CC49 monoclonal antibody was developed by Dr. Jeffrey Schlom's group, Laboratory of Tumor Immunology and Biology, National Cancer Institute. It binds specifically to the pan-carcinoma tumor antigen TAG-72. See Muraro, R., et al., Cancer Research 48:4588-4596 (1988).

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To determine the binding properties of the bivalent CC49/4-4-20 Fv (fraction 5) and the CC49/212 sFv, a competition enzyme-linked immunosorbent assay (ELISA) was set up in which a CC49 IgG labeled with biotin was competed against unlabeled CC49/4-4-20 Fv and the CC49/212 sFv for binding to TAG-72 on a human breast carcinoma extract (see Figure 24). The amount of biotin-labeled CC49 IgG was determined using a preformed complex with avidin and biotin coupled to horse radish peroxidase and O-phenylenediamine dihydrochloride (OPD). The reaction was stopped with 4N H₂SO₄ (sulfuric acid), after 10 min. and the optical density read at 490nm. This competition ELISA showed that the bivalent CC49/4-4-20 Fv binds to the TAG-72 antigen. The CC49/4-4-20 Fv needed a two hundred-fold higher protein concentration to displace the IgG than the single-chain Fv.

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Example 8

Cross-Linking Antigen-Binding Dimers

We have chemically crosslinked dimers of 4-4-20/212 antigen-binding protein with the two cysteine C-terminal extension (4-4-20/212 CPPC single-chain antigen-binding protein) in two ways. In Example 5 we describe the design and genetic construction of the 4-4-20/212 CPPC single-chain antigen-binding protein (hinge design 2 in Table 5). Figure 15B shows the nucleic

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acid and protein sequences of this protein. After purifying the 4-4-20/212 CPPC single-chain antigen-binding protein, using the methods described in Whitlow and Filpula, *Meth. Enzymol.* 2:97 (1991), dimers were formed by two methods. First, the free cysteines were mildly reduced with dithiothreitol (DTT) and then the disulfide-bonds between the two molecules were allowed to form by air oxidation. Second, the chemical crosslinker *bis*-maleimidehexane was used to produce dimers by crosslinking the free cysteines from two 4-4-20/212 CPPC single-chain antigen-binding proteins.

A 0.1 mg/ml solution of the 4-4-20/212 CPPC single-chain antigen-binding protein was mildly reduced using 1 mM DTT, 50 mM HEPES, 50mM NaCl, 1 mM EDTA buffer pH 8.0 at 4°C. The samples were dialyzed against 50mM HEPES, 50 mM NaCl, 1 mM EDTA buffer pH 8.0 at 4°C overnight, to allow the oxidation of free sulfhydrals to intermolecular disulfide-bonds. Figure 25 shows a non-reducing SDS-PAGE gel after the air oxidation; it shows that approximately 10% of the 4-4-20/212 CPPC protein formed dimers with molecular weights around 55,000 Daltons.

A 0.1 mg/ml solution of the 4-4-20/212 CPPC single-chain antigen-binding protein was treated with 2 mM bis-maleimidehexane. Unlike forming a disulfide-bond between two free cysteines in the previous example, the bis-maleimidehexane crosslinker material should be stable to reducing agents such as β -mercaptoethanol. Figure 26 shows that approximately 5% of the treated material produced dimer with a molecular weight of 55,000 Daltons on a reducing SDS-PAGE gel (samples were treated with β -mercaptalethanol prior to being loaded on the gel). We further purified the bis-maleimidehexane treated 4-4-20/212 CPPC protein on PolyCAT A cation exchange column after the protein had been extensively dialyzed against buffer A. Figure 26 shows that we were able to enhance the fraction containing the dimer to approximately 15%.

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Conclusions

We have produced a heterodimer Fv from two complementary mixed sFv's which has been shown to have the size of a dimer of the sFv's. The N-terminal analysis has shown that the active heterodimer Fv contains two polypeptide chains. The heterodimer Fv has been shown to be active for both fluorescein and TAG-72 binding.

All publications cited herein are incorporated fully into this disclosure by reference.

From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention and the following claims. As examples, the steps of the preferred embodiment constitute only one form of carrying out the process in which the invention may be embodied.

-46-SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Whitlow, Marc Wood, James F. Hardman, Karl Bird, Robert Filpula, David Rollence, Michele
- (ii) TITLE OF INVENTION: Multivalent Antigen-Binding Proteins
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 - (E) COUNTRY: U.S.A.
 - (F) ZIP: 20036
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: (to be assigned)
 - (B) FILING DATE: Herewith
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 07/796,936
 - (B) FILING DATE: 25-NOV-1991
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Goldstein, Jorge A.
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 - (B) TELEFAX: (202) 833-8716
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 12 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
 - Gly Lys Ser Ser Gly Ser Gly Ser Glu Ser Lys Ser
- (2) INFORMATION FOR SEQ ID NO:2:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 14 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
 - Gly Ser Thr Ser Gly Ser Gly Lys Ser Ser Glu Gly Lys Gly

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-47-

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 18 amino acids
- (B) TYPE: amino acid (D) TOPOLOGY: both
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Gly Ser Thr Ser Gly Ser Gly Lys Ser Ser Glu Gly Ser Gly Ser Thr

Lys Gly

- (2) INFORMATION FOR SEQ ID NO:4:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 12 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Gly Ser Thr Ser Gly Lys Pro Ser Glu Gly Lys Gly

- (2) INFORMATION FOR SEQ ID NO:5:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids (B) TYPE: amino acid (D) TOPOLOGY: both
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Glu Pro Arg Gly Pro Thr Ile Lys Pro Cys Pro Pro Cys Leu Cys

- (2) INFORMATION FOR SEQ ID NO:6:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid (D) TOPOLOGY: both
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Ala Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys

- (2) INFORMATION FOR SEQ ID NO:7:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 4 amino acids(B) TYPE: amino acid(D) TOPOLOGY: both
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Val Thr Val Ser

121	INFORMATION	FOR	SEO	TD	NO - 8 :
(2)	INFURMATION	FUK	SEQ	ıv	MOTO

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 11 amino acids (B) TYPE: amino acid

 - (D) TOPOLOGY: both
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Val Thr Val Ser Ser Asp Lys Thr His Thr Cys

- (2) INFORMATION FOR SEQ ID NO:9:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 14 amino acids
 (B) TYPE: amino acid

 - (D) TOPOLOGY: both
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Val Thr Val Ser Ser Asp Lys Thr His Thr Cys Pro Pro Cys

- (2) INFORMATION FOR SEQ ID NO:10:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 731 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: both
 - (D) TOPOLOGY: both
 - (ix) FEATURE:

 - (A) NAME/KEY: CDS (B) LOCATION: 1..729
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

GAC GTC Asp Val	GTT ATG A	CT CAG ACA (hr Gln Thr I 5	CCA CTA TCA CTT CCT Pro Leu Ser Leu Pro 10	GTT AGT CTA GGT Val Ser Leu Gly 15	48
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GAT CAA GCC TCC ATC TCT TGC AGA TCT AGT CAG AGC CTT GTA CAC AGT Asp Gln Ala Ser Ile Ser Cys Arg Ser Ser Gln Ser Leu Val His Ser 20 25 30 96

AAT GGA AAC ACC TAT TTA CGT TGG TAC CTG CAG AAG CCA GGC CAG TCT Asn Gly Asn Thr Tyr Leu Arg Trp Tyr Leu Gln Lys Pro Gly Gln Ser

CCA AAG GTC CTG ATC TAC AAA GTT TCC AAC CGA TTT TCT GGG GTC CCA
Pro Lys Val Leu Ile Tyr Lys Val Ser Asn Arg Phe Ser Gly Val Pro
50 60

GAC AGG TTC AGT GGC AGT GGA TCA GGG ACA GAT TTC ACA CTC AAG ATC ABB Arg Phe Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile 240

AGC AGA GTG GAG GCT GAG GAT CTG GGA GTT TAT TTC TGC TCT CAA AGT Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Phe Cys Ser Gln Ser 288

ACA CAT GTT CCG TGG ACG TTC GGT GGA GGC ACC AAG CTT GAA ATC AAA Thr His Val Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys

GGT TCT ACC TCT GGT TCT GGT AAA TCC TCT GAA GGC AAA GGT CAG GTT Gly Ser Thr Ser Gly Ser Gly Lys Ser Ser Glu Gly Lys Gly Gln Val 384

CAG Gln	CTG Leu 130	CAG Gln	CAG Gln	TCT Ser	GAC Asp	GCT Ala 135	GAG Glu	TTG Leu	GTG Val	AAA Lys	CCT Pro 140	GGG Gly	GCT Ala	TCA Ser	GTG Val	4	132
AAG Lys 145	ATT Ile	TCC Ser	TGC Cys	AAG Lys	GCT Ala 150	TCT Ser	GGC Gly	TAC Tyr	ACC Thr	TTC Phe 155	ACT Thr	GAC Asp	CAT His	GCA Ala	ATT Ile 160	4	180
CAC His	TGG Trp	GTG Val	AAA Lys	CAG Gln 165	AAC Asn	CCT Pro	GAA Glu	CAG Gln	GGC Gly 170	CTG Leu	GAA Glu	TGG Trp	ATT Ile	GGA Gly 175	TAT Tyr	5	528
TTT Phe	TCT Ser	CCC Pro	GGA Gly 180	AAT Asn	GAT Asp	GAT Asp	TTT Phe	AAA Lys 185	TAC Tyr	AAT Asn	GAG Glu	AGG Arg	TTC Phe 190	F F F F F F F F F F F F F F F F F F F	GGC Gly	5	576
AAG Lys	GCC Ala	ACA Thr 195	CTG Leu	ACT Thr	GCA Ala	GAC Asp	AAA Lys 200	TCC Ser	TCC Ser	AGC Ser	ACT Thr	GCC Ala 205	TAC Tyr	GTG Val	CAG Gln	•	524
CTC Leu	AAC Asn 210	AGC Ser	CTG Leu	ACA Thr	TCT Ser	GAG Glu 215	GAT Asp	TCT Ser	GCA Ala	GTG Val	TAT Tyr 220	TTC Phe	TGT Cys	ACA Thr	AGA Arg	•	672
TCC Ser 225	CTG Leu	aat Asd	ATG Met	GCC Al _a	TAC Tyr 230	TGG Trp	GGT Gly	CAA Gln	GGA Gly	ACC Thr 235	TCA Ser	GTC Val	ACC Thr	GTC Val	TCC Ser 240		720
TAA	TAG	GAT Asp	CC													•	731

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 243 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
- Asp Val Val Met Thr Gln Thr Pro Leu Ser Leu Pro Val Ser Leu Gly
 1 5 10 15
- Asp Gln Ala Ser Ile Ser Cys Arg Ser Ser Gln Ser Leu Val His Ser 20 25 30
- Asn Gly Asn Thr Tyr Leu Arg Trp Tyr Leu Gln Lys Pro Gly Gln Ser 35 40 45
- Pro Lys Val Leu Ile Tyr Lys Val Ser Asn Arg Phe Ser Gly Val Pro 50 60
- Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile 65 70 75 80
- Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Phe Cys Ser Gln Ser 85 90 95
- Thr His Val Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys 100 105 110
- Gly Ser Thr Ser Gly Ser Gly Lys Ser Ser Glu Gly Lys Gly Gln Val
- Gln Leu Gln Gln Ser Asp Ala Glu Leu Val Lys Pro Gly Ala Ser Val 130 135 140
- Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asp His Ala Ile 145 155 160
- His Trp Val Lys Gln Asn Pro Glu Gln Gly Leu Glu Trp Ile Gly Tyr 165 170 175

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Phe Ser Pro Gly Asn Asp Asp Phe Lys Tyr Asn Glu Arg Phe Lys Gly 185 Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser Ser Thr Ala Tyr Val Gln
195 200 205 Leu Asn Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Phe Cys Thr Arg Ser Leu Asn Met Ala Tyr Trp Gly Gln Gly Thr Ser Val Thr Val Ser 225 230 235

* Asp

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 744 base pairs (B) TYPE: nucleic acid

 - (C) STRANDEDNESS: both
 - (D) TOPOLOGY: both

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 1..744

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

GAC Asp	GTC Val	GTG Val	ATG Met	TCA Ser 5	CAG Gln	TCT Ser	CCA Pro	TCC Ser	TCC Ser 10	CTA Leu	CCT Pro	GTG Val	TCA Ser	GTT Val 15	GGC Gly	48
GAG Glu	AAG Lys	GTT Val	ACT Thr 20	TTG Leu	AGC Ser	TGC Cys	AAG Lys	TCC Ser 25	AGT Ser	CAG Gln	AGC Ser	CTT	TTA Leu 30	TAT Tyr	AGT Ser	96
GGT Gly	AAT Asn	CAA Gln 35	AAG Lys	AAC Asn	TAC Tyr	TTG Leu	GCC Ala 40	TGG Trp	TAC Tyr	CAG Gln	CAG Gln	AAA Lys 45	CCA Pro	GGG Gly	CAG Gln	144
TCT Ser	CCT Pro 50	AAA Lys	CTG Leu	CTG Leu	ATT Ile	TAC Tyr 55	TGG Trp	GCA Ala	TCC Ser	GCT Ala	AGG Arg 60	GAA Glu	TCT Ser	GGG Gly	GTC Val	· 192
CCT Pro 65	GAT Asp	CGC Arg	TTC Phe	ACA Thr	GGC Gly 70	AGT Ser	GGA Gly	TCT Ser	GGG Gly	ACA Thr 75	GAT Asp	TTC Phe	ACT Thr	CTC Leu	TCC Ser 80	240
ATC Ile	AGC Ser	AGT Ser	GTG Val	AAG Lys 85	ACT Thr	GAA Glu	GAC Asp	CTG Leu	GCA Ala 90	GTT Val	TAT Tyr	TAC Tyr	TGT Cys	CAG Gln 95	CAG Gln	288
TAT Tyr	TAT Tyr	AGC Ser	TAT Tyr 100	CCC Pro	CTC Leu	ACG Thr	TTC Phe	GGT Gly 105	GCT Ala	GGG Gly	ACC Thr	AAG Lys	CTT Leu 110	GTG Val	CTG Leu	336
Lys	GGC Gly	Ser 115	Thr	Ser	GIY	ser	120	гÀв	Ser	Ser	GIG	125	-1-	2		384
Val	AAA Lys 130	Leu	Asp	GIU	Thr	135	GIY	GIY	Dea		140		•	_		432
Met 145	AAA Lys	Leu	Ser	Сув	150	Ala	Ser	GIY	FITC	155			•	•	160	480
ATG Met	AAC Asn	TGG Trp	GTC Val	CGC Arg 165	CAG Gln	TCT Ser	CCA Pro	GAG Glu	AAA Lys 170	GGA Gly	CTG Leu	GAG Glu	TGG Trp	GTA Val 175	GCA Ala	528
CAA Gln	ATT Ile	AGA Arg	AAC Asn 180	AAA Lys	CCT Pro	TAT Tyr	AAT Asn	TAT Tyr 185	GAA Glu	ACA Thr	TAT Tyr	TAT Tyr	TCA Ser 190	GAT Asp	TCT Ser	576

									-5	1 <u>_</u>						
GTG Val	AAA Lys	GGC Gly 195	Arg	TTC Phe	ACC Thr	ATC Ile	TCA Ser 200	AGA Arg	GAT	GAT	TCC Ser	AAA Lys 205	AGT Ser	AGT Ser	GTC Val	624
TAC Tyr	CTG Leu 210	CAA Gln	ATG Met	AAC Asn	AAC Asn	TTA Leu 215	AGA Arg	GTT Val	GAA Glu	GAC Asp	ATG Met 220	GGT Gly	ATC Ile	TAT Tyr	TAC Tyr	672
TGT Cys 225	ACG Thr	GGT Gly	TCT Ser	TAC Tyr	TAT Tyr 230	GGT Gly	ATG Met	GAC Asp	TAC Tyr	TGG Trp 235	GGT Gly	CAA Gln	GGA Gly	ACC Thr	TCA Ser 240	720
	ACC Thr															744

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 248 amino acids
 - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13: Asp Val Val Met Ser Gln Ser Pro Ser Ser Leu Pro Val Ser Val Gly
1 5 10 15 Glu Lys Val Thr Leu Ser Cys Lys Ser Ser Gln Ser Leu Leu Tyr Ser 20 25 30 Gly Asn Gln Lys Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln 35 40 45 Ser Pro Lys Leu Leu Ile Tyr Trp Ala Ser Ala Arg Glu Ser Gly Val Pro Asp Arg Phe Thr Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Ser 65 70 75 80 Ile Ser Ser Val Lys Thr Glu Asp Leu Ala Val Tyr Tyr Cys Gln Gln 85 90 95 Tyr Tyr Ser Tyr Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Val Leu 100 105 110 Lys Gly Ser Thr Ser Gly Ser Gly Lys Ser Ser Glu Gly Lys Gly Glu 115 120 125 Val Lys Leu Asp Glu Thr Gly Gly Gly Leu Val Gln Pro Gly Arg Pro 130 140 Met Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asp Tyr Trp 145 150 155 160 Met Asn Trp Val Arg Gln Ser Pro Glu Lys Gly Leu Glu Trp Val Ala 165 170 175 Gln Ile Arg Asn Lys Pro Tyr Asn Tyr Glu Thr Tyr Tyr Ser Asp Ser 180 185 Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys Ser Ser Val 195 200 205 Tyr Leu Gln Met Asn Asn Leu Arg Val Glu Asp Met Gly Ile Tyr Tyr 210 215 220

Cys Thr Gly Ser Tyr Tyr Gly Met Asp Tyr Trp Gly Gln Gly Thr Ser 235 240

Val Thr Val Ser * * Gly Ser 245

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(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 761 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: both
(D) TOPOLOGY: both

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 1..756

(xi) 5	SEOUENCE	DESCRIPTION:	SEQ	ID	NO:14:
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	(xi)	SE	QUEN	CE DI	ESCRI	PTIC	ON: S	SEQ :	ID NO	14:	:					
Asp 1	Val	Val	ATG Met	Thr 5	Gin	Thr	Pro	Leu	10	Deu				15	-	48
Asp	Gln	Ala	TCC Ser 20	IIe	ser	Сув	Arg	25	-				30			96
AAT Asn	GGA Gly	AAC Asn 35	ACC Thr	TAT Tyr	TTA Leu	CGT Arg	TGG Trp 40	TAC Tyr	CTG Leu	CAG Gln	AAG. Lys	CCA Pro- 45	GGC Gly	CAG Gln	TCT Ser	144
Pro	Lys 50	Val	CTG Leu	Ile	Tyr	ьув 55	vai	Ser	YPII	nrg	60					192
Asp 65	Arg	Phe	AGT Ser	Gly	70	GIÅ	Ser	GIŞ	1111	75				-•	80	240
Ser	Arg	Val	GAG Glu	85	GIU	Asp	Pen	GIY	90	-1-		-1-		95		288
Thr	His	Val	CCG Pro 100	Trp	Thr	Pne	GIY	105	GLY	****			110	-	_	336
Gly	Ser	Thr 115	TCT Ser	Gly	Ser	GIĀ	120	Ser	Ser	GIG	GLY	125	1			384
Lys	Leu 130	Asp	GAG Glu	Thr	GIÀ	135	GIY	Leu	Val	GIII	140	,				432
Lys 145	Leu	Ser	TGT Cys	Val	150	Ser	GIY	Pile	1111	155				•	160	480
Asn	Trp	Val	Arg	GIN 165	ser	PIO	GIU	пåв	170	200				175		528
Ile	Arg	Asn	Lys 180	Pro	туг	ASII	TÄT	185		-1-	-2-		190			576
AAA Lys	GGC Gly	AGA Arg 195	TTC Phe	ACC Thr	ATC Ile	TCA Ser	AGA Arg 200	GAT Asp	GAT Asp	TCC Ser	AAA Lys	AGT Ser 205	AGT Ser	GTC Val	TAC Tyr	624
Leu	Gln 210	Met	AAC Asn	Asn	Leu	215	Val	GIU	vob		220		•	-		672
ACG Thr 225	GGT Gly	TCT Ser	TAC Tyr	TAT Tyr	GGT Gly 230	ATG Met	GAC Asp	TAC Tyr	TGG Trp	GGT Gly 235	CAA Gln	GGA Gly	ACC Thr	TCG Ser	GTC Val 240	720

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ACC GTC TCC AGT GAT AAG ACC CAT ACA TGC TAA TAGGATCC Thr Val Ser Ser Asp Lys Thr His Thr Cys

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- (2) INFORMATION FOR SEQ ID NO:15:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 251 amino acids (B) TYPE: amino acid

 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Asp Val Val Met Thr Gln Thr Pro Leu Ser Leu Pro Val Ser Leu Gly

Asp Gln Ala Ser Ile Ser Cys Arg Ser Ser Gln Ser Leu Val His Ser

Asn Gly Asn Thr Tyr Leu Arg Trp Tyr Leu Gln Lys Pro Gly Gln Ser

Pro Lys Val Leu Ile Tyr Lys Val Ser Asn Arg Phe Ser Gly Val Pro 50 60

Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile 65 70 75 80

Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Phe Cys Ser Gln Ser 85 90 95

Thr His Val Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys
100 105 110

Gly Ser Thr Ser Gly Ser Gly Lys Ser Ser Glu Gly Lys Gly Glu Val 115 120 125

Lys Leu Asp Glu Thr Gly Gly Gly Leu Val Gln Pro Gly Arg Pro Met 130 140

Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asp Tyr Trp Met

Asn Trp Val Arg Gln Ser Pro Glu Lys Gly Leu Glu Trp Val Ala Gln

Ile Arg Asn Lys Pro Tyr Asn Tyr Glu Thr Tyr Tyr Ser Asp Ser Val

Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys Ser Ser Val Tyr

Leu Gln Met Asn Asn Leu Arg Val Glu Asp Met Gly Ile Tyr Tyr Cys 210 220

Thr Gly Ser Tyr Tyr Gly Met Asp Tyr Trp Gly Gln Gly Thr Ser Val 225 230 240

Thr Val Ser Ser Asp Lys Thr His Thr Cys

- (2) INFORMATION FOR SEQ ID NO:16:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 770 base pairs (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: both
 - (D) TOPOLOGY: both

(ix) FEATURE: (A)-NAME/KEY: CDS (B) LOCATION: 1..765

	(xi)	SEC	UENC	E DE	SCRI	PTIC	N: 5	SEQ 1	D NC	:16:	:					
GAC Asp 1							CC3	CTA	TCA Ser 10	CTT	CCT	GTT Val	AGT Ser	CTA Leu 15	GGT Gly	48
Asp	Gln	Ala	Ser 20	He	ser	Cys	Arg	25	AGT Ser				30			96
Asn	Gly	Asn 35	Thr	Tyr	Leu	Arg	40	+1-	CTG Leu		-2-	45	•			144
Pro	Lys 50	Val	Leu	IIe	туг	55 55	Val	261	AAC Asn	••••	60		•			192
Asp 65	Arg	Phe	Ser	GIÀ	70	GIŸ	Ser	GLY	ACA Thr	75				•	80	240
Ser	Arg	Val	Glu	85	GIU	мвр	TIEG	GIŞ	GTT Val 90	-1-		•		95		288
Thr	His	Val	Pro 100	Trp	THE	Pne	GLY	105	GGC		-,-		110			336
Gly	Ser	Thr 115	Ser	GIĀ	ser	GIY	120	Ser	TCT Ser		1	125	•			384
AAA Lys	CTG Leu 130	GAT Asp	GAG Glu	ACT Thr	GGA Gly	GGA Gly 135	GGC Gly	TTG Leu	GTG Val	CAA Gln	CCT Pro 140	GGG Gly	AGG Arg	Pro	ATG Met	432
Lys 145	Leu	Ser	Cys	Val	150	Ser	GIY	FIIC	ACT	155		•	•		160	480
Asn	Trp	Val	Arg	165	Ser	PLO	سيرى	טעט	GGA Gly 170			•		175		528
Ile	Arg	Asn	Lys 180	Pro	lyr	WRII	171	185		-,-	-1-		190	•		576
Lys	Gly	Arg 195	Phe	Tnr	TTE	SEL	200	vob	GAT Asp		-,	205				624
Leu	Gln 210	Met	Asn	ABN	Leu	215	Val	0.10		•••	220		_			672
Thr 225	Gly	Ser	Tyr	lyr	230	Mer	WPP	-1-	TGG Trp	235					240	720
		TCC Ser	AGT Ser	GAT Asp 245	TAR	ACC Thr	CAT	ACA Thr	TGC Cys 250		CCA Pro	TGC	TAA	TAG 255	GATCC	770

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 254 amino acids (B) TYPE: amino acid

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Asp Val Val Met Thr Gln Thr Pro Leu Ser Leu Pro Val Ser Leu Gly

Asp Gln Ala Ser Ile Ser Cys Arg Ser Ser Gln Ser Leu Val His Ser

Asn Gly Asn Thr Tyr Leu Arg Trp Tyr Leu Gln Lys Pro Gly Gln Ser 35 40

Pro Lys Val Leu Ile Tyr Lys Val Ser Asn Arg Phe Ser Gly Val Pro 50 55 60

Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile 65 70 75 80

Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Phe Cys Ser Gln Ser 85 90 95

Thr His Val Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys

Gly Ser Thr Ser Gly Ser Gly Lys Ser Ser Glu Gly Lys Gly Glu Val

Lys Leu Asp Glu Thr Gly Gly Gly Leu Val Gln Pro Gly Arg Pro Met 130 135 140

Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asp Tyr Trp Met

. Asn Trp Val Arg Gln Ser Pro Glu Lys Gly Leu Glu Trp Val Ala Gln

Ile Arg Asn Lys Pro Tyr Asn Tyr Glu Thr Tyr Tyr Ser Asp Ser Val

Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys Ser Ser Val Tyr 195 200 205

Leu Gln Met Asn Asn Leu Arg Val Glu Asp Met Gly Ile Tyr Tyr Cys

Thr Gly Ser Tyr Tyr Gly Met Asp Tyr Trp Gly Gln Gly Thr Ser Val 225 230 240

Thr Val Ser Ser Asp Lys Thr His Thr Cys Pro Pro Cys *

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1460 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: both
 - (D) TOPOLOGY: both

(ix) FEATURE:

- (A) NAME/KEY: CDS (B) LOCATION: 1..1398
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GAC Asp	GTC Val	GTG Val	ATG M <u>e</u> t	TCA Ser 5	CAG Gln	TCT Ser	CCA Pro	TCC Ser	TCC Ser 10	CTA Leu	CCT Pro	GTG Val	TCA Ser	GTT Val 15	GGC Gly	48
GAG Glu	AAG Lys	GTT Val	ACT Thr 20	TTG Leu	AGC Ser	TGC Cys	AAG Lys	TCC Ser 25	AGT Ser	CAG Gln	AGC Ser	CTT Leu	TTA Leu 30	TAT Tyr	AGT Ser	96
Gly	Asn	Gln 35	Lys	Asn	Tyr	Leu	40	Trp	TYE	GIN	GIII	45	710	G ₂ ,	CAG Gln	144
Ser	Pro 50	Lys	Leu	CTG Leu	Ile	Tyr 55	Trp	Ala	Ser	Ala	60	-		CII		192
Pro 65	yeb	Arg	Phe	ACA Thr	70	Ser	GIY	Ser	GLY	75	Nop	1			80	240
Ile	Ser	Ser	Val	AAG Lys 85	Thr	Glu	Asb	ren	90	Val	TYL	ıyı	Cy.	95		 288
Tyr	Tyr	Ser	Tyr 100	Pro	Leu	Thr	Pne	105	Ala		1111	<i>- y</i> -	110			336
Lys	Gly	Ser 115	Thr	Ser	GIA	Ser	120	пув	261	J-12		125	-,-			384
Val	Gln 130	Leu	Gln	Gln	Ser	135	Ala	GIU	Pen	AGT	140	110	- 2			432
Val 145	Lys	Ile	Ser	TGC Cys	150	Ala	Ser	GIY	171	155					160	480
Ile	His	Trp	Val	AAA Lys 165	Gln _.	Asn	PLO	GIU	170	GTY	пец	314		175		528
Tyr	Phe	Ser	Pro 180	Gly	Asn	Asp	Asp	185	гув	IYE	ABII	GIU	190			576
Gly	Lys	Ala 195	Thr	CTG Leu	Thr	Ala	200	гув	Ser	Ser	361	205		-1-		624
CAG Gln	CTC Leu 210	AAC Asn	AGC Ser	CTG Leu	ACA Thr	TCT Ser 215	GAG Glu	GAT Asp	TCT Ser	GCA Ala	GTG Val 220	TAT Tyr	TTC Phe	Cys	ACA Thr	672
AGA Arg 225	TCC Ser	CTG Leu	AAT Asn	ATG Met	GCC Ala 230	TAC Tyr	TGG Trp	GGT Gly	CAA Gln	GGA Gly 235	ACC Thr	TCA Ser	GTC Val	ACC Thr	GTC Val 240	720
Ser	Ser	Asp	Val	GTG Val 245	Met	ser	GIII	SEL	250	Jer	J.,			255		768
Val	Gly	Glu	Lys 260	GTT Val	Thr	Leu	Ser	265	пув	361	501		270			816
Tyr	Ser	Gly 275	Asn	CAA Gln	Lys	Asn	280	ьeu	ATA	IIP	-11-	285	U_	-,-	-	864
GGG Gly	CAG Gln 290	TCT Ser	CCT Pro	AAA Lys	CTG Leu	CTG Leu 295	ATT Ile	TAC Tyr	TGG Trp	GCA Ala	TCC Ser 300	GCT Ala	AGG Arg	GAA Glu	TCT Ser	912

GGG Gly 305	GTC Val	CCT Pro	GAT Asp	CGC Arg	TTC Phe 310	ACA Thr	GGC Gly	AGT Ser	GGA Gly	TCT Ser 315	GGG Gly	ACA Thr	GAT Asp	TTC Phe	ACT Thr 320	960
CTC Leu	TCC Ser	ATC Ile	AGC Ser	AGT Ser 325	GTG Val	AAG Lys	ACT Thr	GAA Glu	GAC Asp 330	CTG Leu	GCA Ala	GTT Val	TAT Tyr	TAC Tyr 335	TGT Cys	1008
CAG Gln	CAG Gln	TAT Tyr	TAT Tyr 340	AGC Ser	TAT Tyr	CCC Pro	CTC Leu	ACG Thr 345	TTC Phe	GGT Gly	GCT Ala	GGG Gly	ACC Thr 350	AAG Lys	CTT Leu	1056
GTG Val	CTG Leu	AAA Lys 355	GGC Gly	TCT Ser	ACT Thr	TCC Ser	GGT Gly 360	AGC Ser	GGC Gly	AAA Lys	TCC Ser	TCT Ser 365	GAA Glu	GGC Gly	AAA Lys	1104
GGT Gly	CAG Gln 370	GTT Val	CAG Gln	CTG Leu	CAG Gln	CAG Gln 375	TCT Ser	GAC Asp	GCT Ala	GAG Glu	TTG Leu 380	GTG Val	AAA Lys	CCT Pro	GGG Gly	1152
GCT Ala 385	TCA Ser	GTG Val	AAG Lys	ATT Ile	TCC Ser 390	TGC Cys	AAG Lys	GCT Ala	TCT Ser	GGC Gly 395	TAC Tyr	ACC Thr	TTC Phe	ACT Thr	GAC Asp 400	1200
CAT His	GCA Ala	ATT Ile	CAC His	TGG Trp 405	GTG Val	AAA Lys	CAG Gln	AAC Asn	CCT Pro 410	GAA Glu	CAG Gln	GGC Gly	CTG Leu	GAA Glu 415	TGG Trp	1248
ATT Ile	GGA Gly	TAT	TTT Phe 420	TCT Ser	CCC Pro	GGA Gly	AAT Asn	GAT Asp 425	GAT Asp	TTT Phe	AAA Lys	TAC Tyr	AAT Asn 430	GAG Glu	AGG Arg	1296
TTC Phe	AAG Lys	GGC Gly 435	AAG Lys	GCC Ala	ACA Thr	CTG Leu	ACT Thr 440	GCA Ala	GAC Asp	AAA Lys	TCC Ser	TCC Ser 445	AGC Ser	ACT Thr	GCC Ala	1344
	GTG Val 450	CAG Gln	CTC Leu	AAC Asn	AGC Ser	CTG Leu 455	ACA Thr	TCT Ser	GAG Glu	GAT Asp	TCT Ser 460	GCA Ala	GTG Val	TAT Tyr	TTC Phe	1392
TGT Cys 465	ACA Thr	AGA Arg	TCC Ser	CTG Leu	AAT Asn 470	ATG Met	GCC Ala	TAC Tyr	TGG Trp	GGT Gly 475	CAA Gln	GGA Gly	ACC Thr	TCA Ser	GTC Val 480	1440
ACC Thr			TAA	TAG * 485	GAT Asp	cc										1460

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 486 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Asp Val Val Met Ser Gln Ser Pro Ser Ser Leu Pro Val Ser Val Gly
1 5 10 15

Glu Lys Val Thr Leu Ser Cys Lys Ser Ser Gln Ser Leu Leu Tyr Ser 20 25 30

Gly Asn Gln Lys Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln 35 40 45

Ser Pro Lys Leu Leu Ile Tyr Trp Ala Ser Ala Arg Glu Ser Gly Val 50 60

Pro Asp Arg Phe Thr Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Ser 65 70 80

Ile Ser Ser Val Lys Thr Glu Asp Leu Ala Val Tyr Ser Cys Gln Gln 85 90 95 Tyr Tyr Ser Tyr Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Val Leu 100 105 110 Lys Gly Ser Thr Ser Gly Ser Gly Lys Ser Ser Glu Gly Lys Gly Gln
115 120 125 Val Gln Leu Gln Gln Ser Asp Ala Glu Leu Val Lys Pro Gly Ala Ser 130 140 Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asp His Ala 145 150 150 160 Ile His Trp Val Lys Gln Asn Pro Glu Gln Gly Leu Glu Trp Ile Gly 165 170 175 Tyr Phe Ser Pro Gly Asn Asp Asp Phe Lys Tyr Asn Glu Arg Phe Lys Gly Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser Ser Thr Ala Tyr Val 195 200 205 Gln Leu Asn Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Phe Cys Thr 210 215 220 Arg Ser Leu Asn Met Ala Tyr Trp Gly Gln Gly Thr Ser Val Thr Val 225 230 235 Ser Ser Asp Val Val Met Ser Gln Ser Pro Ser Ser Leu Pro Val Ser Val Gly Glu Lys Val Thr Leu Ser Cys Lys Ser Ser Gln Ser Leu Leu 260 265 270 Tyr Ser Gly Asn Gln Lys Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Pro 275 280 285 Gly Gln Ser Pro Lys Leu Leu Ile Tyr Trp Ala Ser Ala Arg Glu Ser 290 295 Gly Val Pro Asp Arg Phe Thr Gly Ser Gly Ser Gly Thr Asp Phe Thr 320 Leu Ser Ile Ser Ser Val Lys Thr Glu Asp Leu Ala Val Tyr Cys 325 330 335 Gln Gln Tyr Tyr Ser Tyr Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu 340 Val Leu Lys Gly Ser Thr Ser Gly Ser Gly Lys Ser Ser Glu Gly Lys 355 360 365 Gly Gln Val Gln Leu Gln Gln Ser Asp Ala Glu Leu Val Lys Pro Gly 370 380 Ala Ser Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asp 385 390 395 His Ala Ile His Trp Val Lys Gln Asn Pro Glu Gln Gly Leu Glu Trp Ile Gly Tyr Phe Ser Pro Gly Asn Asp Asp Phe Lys Tyr Asn Glu Arg 420 425 430 Phe Lys Gly Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser Thr Ala Tyr Val Gln Leu Asn Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Phe 450 455 Cys Thr Arg Ser Leu Asn Met Ala Tyr Trp Gly Gln Gly Thr Ser Val Thr Val Ser 485

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(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 725 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: both

(D) TOPOLOGY: both

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 1..723

COLOTION, SEC ID NO.20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:																	
GAC Asp 1	GTC Val	GTT Val	ATG Met	ACT Thr 5	CAG Gln	ACA Thr	CCA Pro	CTA Leu	TCA Ser 10	CTT Leu	CCT Pro	GTT Val	AGT Ser	CTA Leu 15	GGT Gly		48
GAT Asp	CAA Gln	GCC Ala	TCC Ser 20	ATC Ile	TCT Ser	TGC	AGA Arg	TCT Ser 25	AGT Ser	CAG Gln	AGC Ser	CTT Leu	GTA Val 30	CAC His	AGT Ser		96
AAT Asn	GGA Gly	AAC Asn 35	ACC Thr	TAT Tyr	TTA Leu	CGT Arg	TGG Trp 40	TAC Tyr	CTG Leu	CAG Gln	AAG Lys	CCA Pro 45	GGC Gly	CAG Gln	TCT Ser	1	.44
CCA Pro	AAG Lys 50	GTC Val	CTG Leu	ATC Ile	TAC Tyr	AAA Lys 55	GTT Val	TCC Ser	AAC Asn	CGA Arg	TTT Phe 60	TCT Ser	GGG Gly	GTC Val	CCA Pro	1	.92
GAC Asp 65	AGG Arg	TTC Phe	AGT Ser	GGC Gly	AGT Ser 70	GGA Gly	TCA Ser	GGG Gly	ACA Thr	GAT Asp 75	TTC Phe	ACA Thr	CTC Leu	AAG Lys	ATC Ile 80	2	40
AGC Ser	AGA Arg	GTG Val	GAG Glu	GCT Ala 85	GAG Glu	GAT Asp	CTG Leu	GGA Gly	GTT Val 90	TAT Tyr	TTC Phe	TGC	TCT Ser	CAA Gln 95	AGT Ser	2	88
ACA Thr	CAT His	GTT Val	CCG Pro 100	TGG Trp	ACG Thr	TTC Phe	GGT Gly	GGA Gly 105	GGC Gly	ACC Thr	AAG Lys	CTT Leu	GAA Glu 110	ATC Ile	AAA Lys	3	36
GGT Gly	TCT Ser	ACC Thr 115	TCT Ser	GGT Gly	AAA Lys	CCA Pro	TCT Ser 120	GAA Glu	GGC Gly	AAA Lys	GGT Gly	CAG Gln 125	GTT Val	CAG Gln	CTG Leu	3	84
CAG Gln	CAG Gln 130	TCT Ser	GAC Asp	GCT Ala	GAG Glu	TTG Leu 135	GTG Val	AAA Lys	CCT Pro	GGG Gly	GCT Ala 140	TCA Ser	GTG Val	AAG Lys	ATT Ile	4	32
TCC Ser 145	TGC Cys	AAG Lys	GCT Ala	TCT Ser	GGC Gly 150	TAC Tyr	ACC Thr	TTC Phe	ACT Thr	GAC Asp 155	CAT His	GCA Ala	ATT Ile	CAC His	TGG Trp 160	4	80
GTG Val	TA8	CAG Gln	AAC Asn	CCT Pro 165	GAA Glu	CAG Gln	GGC Gly	CTG Leu	GAA Glu 170	TGG Trp	ATT Ile	GGA Gly	TAT Tyr	TTT Phe 175	TCT Ser	5	28
CCC Pro	GGA Gly	AAT Asn	GAT Asp 180	GAT Asp	TTT Phe	AAA Lys	TAC Tyr	AAT Asn 185	GAG Glu	agg Arg	TTC Phe	AAG Lys	GGC Gly 190	AAG Lys	GCC Ala	5	76
ACA Thr	CTG Leu	ACT Thr 195	GCA Ala	GAC Asp	AAA Lys	TCC Ser	TCC Ser 200	AGC Ser	ACT Thr	GCC Ala	TAC Tyr	GTG Val 205	CAG Gln	CTC Leu	AAC Asn	6	24
AGC Ser	CTG Leu 210	ACA Thr	TCT Ser	GAG Glu	GAT Asp	TCT Ser 215	GCA Ala	GTG Val	TAT Tyr	TTC Phe	TGT Cys 220	ACA Thr	AGA Arg	TCC Ser	CTG Leu	6	72

AAT ATG GCC TAC TGG GGT CAA GGA ACC TCA GTC ACC GTC TCC TAA TAG 7.20 Asn Met Ala Tyr Trp Gly Gln Gly Thr Ser Val Thr Val Ser . 725 GAT CC Asp

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 241 amino acids
 - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Asp Val Val Met Thr Gln Thr Pro Leu Ser Leu Pro Val Ser Leu Gly

Asp Gln Ala Ser Ile Ser Cys Arg Ser Ser Gln Ser Leu Val His Ser 20 25 30

Asn Gly Asn Thr Tyr Leu Arg Trp Tyr Leu Gln Lys Pro Gly Gln Ser 35

Pro Lys Val Leu Ile Tyr Lys Val Ser Asn Arg Phe Ser Gly Val Pro
50 60

Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile 65 70 80

Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Phe Cys Ser Gln Ser

Thr His Val Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys 100 105 110

Gly Ser Thr Ser Gly Lys Pro Ser Glu Gly Lys Gly Gln Val Gln Leu

Gln Gln Ser Asp Ala Glu Leu Val Lys Pro Gly Ala Ser Val Lys Ile

Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asp His Ala Ile His Trp

Val Lys Gln Asn Pro Glu Gln Gly Leu Glu Trp Ile Gly Tyr Phe Ser

Pro Gly Asn Asp Asp Phe Lys Tyr Asn Glu Arg Phe Lys Gly Lys Ala 180 185

Thr Leu Thr Ala Asp Lys Ser Ser Ser Thr Ala Tyr Val Gln Leu Asn

Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Phe Cys Thr Arg Ser Leu

Asn Met Ala Tyr Trp Gly Gln Gly Thr Ser Val Thr Val Ser

Asp

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 738 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: both
 - (D) TOPOLOGY: both

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(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 1..738

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

	(X1)	SE	SOEM	- DI	SCKI	PIIC										
GAC Asp 1	GTC Val	GTG Val	ATG Met	TCA Ser 5	CAG Gln	TCT Ser	CCA Pro	TCC Ser	TCC Ser 10	CTA Leu	CCT Pro	GTG Val	TCA Ser	GTT Val 15	GGC Gly	48
Glu	Lys	Val	Thr 20	Leu	Ser	Сув	Lys	Ser 25	Ser	Gln	Ser	Leu	Jeu 30		ser	96
GGT Gly	AAT Asn	CAA Gln 35	AAG Lys	AAC Asn	TAC Tyr	TTG Leu	GCC Ala 40	TGG Trp	TAC Tyr	CAG Gln	CAG Gln	AAA Lys 45	CCA Pro	GGG Gly	CAG Gln	144
TCT Ser	CCT Pro 50	ray Yyy	CTG Leu	CTG Leu	ATT Ile	TAC Tyr 55	TGG Trp	GCA Ala	TCC Ser	GCT Ala	AGG Arg 60	GAA Glu	TCT Ser	GGG Gly	GTC Val	192
CCT Pro 65	GAT Asp	CGC Arg	TTC Phe	ACA Thr	GGC Gly 70	AGT Ser	GGA Gly	TCT Ser	GGG Gly	ACA Thr 75	GAT Asp	TTC Phe	ACT Thr	CTC Leu	TCC Ser 80	240
ATC Ile	AGC Ser	AGT Ser	GTG Val	AAG Lys 85	ACT Thr	GAA Glu	GAC Asp	CTG Leu	GCA Ala 90	GTT Val	TAT Tyr	TAC Tyr	TGT Cys	CAG Gln 95	CAG Gln	288
TAT Tyr	TAT Tyr	AGC Ser	TAT Tyr 100	CCC Pro	CTC Leu	ACG Thr	TTC Phe	GGT Gly 105	GCT Ala	GGG Gly	ACC Thr	AAG Lys	CTT Leu 110	GTG Val	CTG Leu	336
AAA Lys	GGC Gly	TCT Ser 115	ACT Thr	TCC Ser	GGT Gly	AAA Lys	CCA Pro 120	TCT Ser	GAA Glu	GGT Gly	AAA Lys	GGT Gly 125	GAA Glu	GTT Val	AAA Lys	384
CTG Leu	GAT Asp 130	GAG Glu	ACT Thr	GGA Gly	GGA Gly	GGC Gly 135	TTG Leu	GTG Val	CAA Gln	CCT Pro	GGG Gly 140	AGG Arg	CCC Pro	ATG Met	AAA Lys	432
CTC Leu 145	TCC Ser	TGT Cys	GTT Val	GCC Ala	TCT Ser 150	GGA Gly	TTC Phe	ACT Thr	TTT Phe	AGT Ser 155	GAC Asp	TAC Tyr	TGG Trp	ATG Met	AAC Asn 160	480
TGG Trp	GTC Val	CGC	CAG Gln	TCT Ser 165	CCA Pro	GAG Glu	AAA Lys	GGA Gly	CTG Leu 170	GAG Glu	TGG Trp	GTA Val	GCA Ala	CAA Gln 175	ATT Ile	528
AGA Arg	AAC Asn	AAA Lys	CCT Pro 180	TAT Tyr	AAT Asn	TAT Tyr	GAA Glu	ACA Thr 185	TAT Tyr	TAT Tyr	TCA Ser	GAT Asp	TCT Ser 190	GTG Val	AAA Lys	576
GGC Gly	AGA Arg	TTC Phe 195	ACC Thr	ATC Ile	TCA Ser	Ara	ABP	Asp	Ser	AAA Lys	Ser	Ser	vaı	TAC Tyr	CTG Leu	624
CAA Gln	ATG Met 210	AAC Asn	AAC Asn	TTA Leu	AGA Arg	GTT Val 215	GAA Glu	GAC Asp	ATG Met	GGT Gly	ATC Ile 220	TAT Tyr	TAC Tyr	TGT Cys	ACG Thr	672
GGT Gly 225	TCT Ser	TÀC Tyr	TAT Tyr	GGT Gly	ATG Met 230	GAC Asp	TAC Tyr	TGG Trp	GGT Gly	CAA Gln 235	GGA Gly	ACC Thr	TCA Ser	GTC Val	ACC Thr 240	720
	TCC Ser		TAA	GGA Gly 245	TCC Ser											738

(2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 246 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23: Asp Val Val Met Ser Gln Ser Pro Ser Ser Leu Pro Val Ser Val Gly Glu Lys Val Thr Leu Ser Cys Lys Ser Ser Gln Ser Leu Leu Tyr Ser 20 25 30 Gly Ash Gln Lys Ash Tyr Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln 35 40 Ser Pro Lys Leu Leu Ile Tyr Trp Ala Ser Ala Arg Glu Ser Gly Val
50 60 Pro Asp Arg Phe Thr Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Ser 65 70 75 Ile Ser Ser Val Lys Thr Glu Asp Leu Ala Val Tyr Tyr Cys Gln Gln 85 90 95 Tyr Tyr Ser Tyr Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Val Leu 100 105 110 Lys Gly Ser Thr Ser Gly Lys Pro Ser Glu Gly Lys Gly Glu Val Lys 115 120 125 Leu Asp Glu Thr Gly Gly Gly Leu Val Gln Pro Gly Arg Pro Met Lys 130 135 Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asp Tyr Trp Met Asn 145 150 155 160 Trp Val Arg Gln Ser Pro Glu Lys Gly Leu Glu Trp Val Ala Gln Ile 165 170 175 Arg Asn Lys Pro Tyr Asn Tyr Glu Thr Tyr Tyr Ser Asp Ser Val Lys 180 185 Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys Ser Ser Val Tyr Leu 195 200 205 Gln Met Asn Asn Leu Arg Val Glu Asp Met Gly Ile Tyr Tyr Cys Thr 210 220 Gly Ser Tyr Tyr Gly Met Asp Tyr Trp Gly Gln Gly Thr Ser Val Thr 225 230 235

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What Is Claimed Is:

- 1. A multivalent antigen-binding protein comprising two or more single-chain molecules, each single-chain molecule comprising:
- (a) a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (b) a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; and
- (c) a peptide linker linking said first and second polypeptides(a) and (b) into said single-chain molecule.
- 2. The multivalent protein of claim 1 wherein said first polypeptide comprises the binding portion of the variable region of an antibody light chain, and said second polypeptide comprises the binding portion of the variable region of an antibody heavy chain.
- 3. The multivalent protein of claim 1 wherein said first polypeptide comprises the binding portion of the variable region of an antibody light chain, and said second polypeptide comprises the binding portion of the variable region of an antibody light chain.
- 4. The multivalent protein of claim 1 wherein said first polypeptide comprises the binding portion of the variable region of an antibody heavy chain, and said second polypeptide comprises the binding portion of the variable region of an antibody heavy chain.
- 5. The multivalent protein of claims 1, 2, 3, or 4 comprising a bivalent antigen-binding protein.
- 6. The multivalent protein of claim 5 comprising a heterobivalent antigen-binding protein.

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- 7. The multivalent protein of claim 5 comprising a homobivalent antigen-binding protein.
- 8. A composition comprising a multivalent antigen-binding protein substantially free of single-chain molecules, wherein said multivalent protein comprises two or more single-chain molecules, each single-chain molecule comprising:
- (a) a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (b) a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; and
- (c) a peptide linker linking said first and second polypeptides(a) and (b) into said single-chain molecule.
- 9. The composition of claim 8 wherein said first polypeptide comprises the binding portion of the variable region of an antibody light chain, and said second polypeptide comprises the binding portion of the variable region of an antibody heavy chain.
- 10. The composition of claim 8 wherein said first polypeptide comprises the binding portion of the variable region of an antibody light chain, and said second polypeptide comprises the binding portion of the variable region of an antibody light chain.
- 11. The composition of claim 8 wherein said first polypeptide comprises the binding portion of the variable region of an antibody heavy chain, and said second polypeptide comprises the binding portion of the variable region of an antibody heavy chain.
- 12. The composition of claims 8, 9, 10, or 11, comprising a bivalent antigen-binding protein substantially free of single-chain molecules.

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- 13. The composition of claim 12 wherein said bivalent protein is heterobivalent.
- 14. The composition of claim 12 wherein said bivalent protein is homobivalent.

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15. An aqueous composition comprising an excess of multivalent antigen-binding protein over single-chain molecules, said multivalent protein comprising two or more single-chain molecules, each single-chain molecule comprising:

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- (a) a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (b) a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; and
- (c) a peptide linker linking said first and second polypeptides
 (a) and (b) into said single-chain protein.

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- 16. The aqueous composition of claim 15 wherein at least one of said single-chain molecules comprises:
- (a) a first polypeptide comprising the binding portion of the variable region of an antibody light chain;
- (b) a second polypeptide comprising the binding portion of the variable region of an antibody heavy chain; and
- (c) a peptide linker linking said first and second polypeptides
 (a) and (b) into said single-chain protein.
- 17. The aqueous composition of claim 15 wherein at least one of said single-chain molecules comprises:

- (a) a first polypeptide comprising the binding portion of the variable region of an antibody light chain;
- (b) a second polypeptide comprising the binding portion of the variable region of an antibody light chain; and

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a peptide linker linking said first and second polypeptides (c) (a) and (b) into said single-chain protein. The composition of claim 15 wherein at least one of said single-18. chain molecules comprises: a first polypeptide comprising the binding portion of the (a) variable region of an antibody heavy chain; a second polypeptide comprising the binding portion of (b) the variable region of an antibody heavy chain; and a peptide linker linking said first and second polypeptides (c) (a) and (b) into said single-chain protein. A method of producing a multivalent antigen-binding protein, 19. comprising the steps of: producing a composition comprising multivalent antigen-(a) binding protein and single-chain molecules, each single-chain molecule comprising: a first polypeptide comprising the binding portion (i) of the variable region of an antibody heavy or light chain; a second polypeptide comprising the binding (ii) portion of the variable region of an antibody heavy or light chain; and a peptide linker linking said first and second (iii) polypeptides (a) and (b) into said single-chain molecule; separating said multivalent protein from said single-chain (b) molecules; and recovering said multivalent protein. (c) The method of claim 19 wherein separating said multivalent 20. protein from said single-chain molecules comprises utilizing cation exchange chromatography.

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- 21. The method of claim 19 wherein separating said multivalent protein from said single-chain molecules comprises utilizing gel filtration chromatography.
- 22. A method of producing a multivalent antigen-binding protein comprising the steps of:
- (a) producing a composition comprising single-chain molecules, each single-chain molecule comprising:
- (i) a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (ii) a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; and
- (iii) a peptide linker linking said first and second polypeptides (a) and (b) into said single-chain molecule;
 - (b) dissociating said single-chain molecules;
 - (c) re-associating said single-chain molecules;
- (d) separating multivalent antigen-binding proteins from said single-chain molecules; and
 - (e) recovering said multivalent proteins.
- 23. The method of claim 22 wherein said dissociation is caused by dialysis against a dissociating solution.
 - 24. The method of claim 22 wherein said reassociation is caused by dialysis against a refolding solution or a refolding agent.
 - 25. A method of producing a multivalent antigen-binding protein, comprising the step of cross-linking at least two single-chain molecules to each other, each single-chain molecule comprising:
 - (a) a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;

comprising the steps of:

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- (b) a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; and
 (c) a peptide linker linking said first and second polypeptides

 (a) and (b) into said single-chain molecule.

 26. The method of claim 25 wherein said cross-linking is effected by chemical means.
- 27. A method of producing a multivalent antigen-binding protein,
- (a) producing a composition comprising single-chain molecules, each single-chain molecule comprising:
- (i) a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (ii) a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; and
- (iii) a peptide linker linking said first and second polypeptides (a) and (b) into said single-chain molecule;
 - (b) concentrating said single-chain molecules;
- (c) separating said multivalent protein from said single-chain molecules; and
 - (d) recovering said multivalent protein.
- 28. The method of claim 27 wherein said concentrating step occurs from approximately 0.5 mg/ml single-chain molecule to the concentration at which precipitation starts.
- 29. A method of detecting an antigen in or suspected of being in a sample, which comprises:
 - (a) contacting said sample with the multivalent antigenbinding protein of claim 1; and

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- detecting whether said multivalent antigen-binding · (b) protein has bound to said antigen.
- A method of imaging the internal structure of an animal, 30. comprising administering to said animal an effective amount of a labeled form of the multivalent antigen-binding protein of claim 1 and measuring detectable radiation associated with said animal.
- A composition comprising an association of a multivalent 31. antigen-binding protein as claimed in any one of claims 1-4, 8-11, or 15-18 with a therapeutically or diagnostically effective agent.

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- A single-chain protein comprising: 32.
- a first polypeptide comprising the binding portion of the (a) variable region of an antibody light chain;
- a second polypeptide comprising the binding portion of (b) the variable region of an antibody light chain;

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- a peptide linker linking said first and second polypeptides (c) (a) and (b) into said single-chain protein.
 - A single-chain protein comprising: 33.
- a first polypeptide comprising the binding portion of the variable region of an antibody heavy chain;

(b)

- a second polypeptide comprising the binding portion of the variable region of an antibody heavy chain;
- a peptide linker linking said first and second polypeptides (c) (a) and (b) into said single-chain protein.
 - A single-chain protein comprising: 34.

a first polypeptide comprising the V_L or V_H of a CC49 25 (a) monoclonal antibody;

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- (b) a second polypeptide comprising the V_L or V_H of a CC49 monoclonal antibody; and
- (c) a peptide linker linking said first and second polypeptides

 (a) and (b) into said single-chain protein.
- 35. The single-chain protein of claim 34 wherein said linker is selected from the group consisting of the 202', 212, 216, and 217 linkers.
 - 36. A single-chain protein comprising:
 - (a) a first polypeptide comprising the V_L or V_H of a CC49 monoclonal antibody;
 - (b) a second polypeptide comprising the V_L or V_H of a 4-4-20 monoclonal antibody; and
 - (c) a peptide linker linking said first and second polypeptides(a) and (b) into said single-chain protein.
 - 37. The single-chain protein of claim 36 wherein said linker is selected from the group consisting of the 202', 212, 216, and 217 linkers.
 - 38. A genetic sequence which codes for the single-chain protein of claim 32, comprising:
 - (a) a DNA sequence coding for a first polypeptide comprising the binding portion of the variable region of an antibody light chain;
 - (b) a DNA sequence coding for a second polypeptide comprising the binding portion of the variable region of an antibody light chain;
 - (c) a DNA sequence coding for a peptide linker linking said first and second polypeptides (a) and (b) into said single-chain protein.
 - 39. A genetic sequence which codes for the single-chain protein of claim 33, comprising:

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- (a) a DNA sequence coding for a first polypeptide comprising the binding portion of the variable region of an antibody heavy chain;
- (b) a DNA sequence coding for a second polypeptide comprising the binding portion of the variable region of an antibody heavy chain;
- (c) a DNA sequence coding for a peptide linker linking said first and second polypeptides (a) and (b) into said single-chain protein.
- 40. A genetic sequence which codes for the single-chain protein of claim 34, comprising:
- (a) a DNA sequence coding for the V_L or V_H of a CC49 monoclonal antibody;
- (b) a DNA sequence coding for the V_L or V_H of a CC49 monoclonal antibody;
- (c) a DNA sequence coding for a peptide linker linking said first and second polypeptides (a) and (b) into said single-chain protein.
- 41. The genetic sequence of claim 40 wherein said DNA sequence codes for a peptide linker selected from the group consisting of the 202', 212, 216, and 217 linkers.
- 42. A genetic sequence which codes for the single-chain protein of claim 36, comprising:
- (a) a DNA sequence coding for the V_L or V_H of a CC49 monoclonal antibody;
- (b) a DNA sequence coding for the $V_{\text{\tiny L}}$ or $V_{\text{\tiny H}}$ of a 4-4-20 monoclonal antibody;
- (c) a DNA sequence coding for a peptide linker linking said first and moond polypeptides (a) and (b) into said single-chain protein.

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- 43. The genetic sequence of claim 42 wherein said DNA sequence codes for a peptide linker selected from the group consisting of the 202', 212, 216, and 217 linkers.
 - 44. A multivalent single-chain antigen-binding protein comprising:

(a) a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;

- (b) a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (c) a peptide linker linking said first and second polypeptides
 (a) and (b) into said multivalent protein;
- (d) a third polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (e) a fourth polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (f) a peptide linker linking said third and fourth polypeptides(d) and (e) into said multivalent protein; and
- (g) a peptide linker linking said second and third polypeptides (b) and (d) into said multivalent protein.
 - 45. A multivalent single-chain antigen-binding protein comprising:
- (a) a first polypeptide comprising the binding portion of the variable region of an antibody light chain;
- (b) a second polypeptide comprising the binding portion of the variable region of an antibody heavy chain;
- (c) a peptide linker linking said first and second polypeptides

 (a) and (b) into said multivalent protein;
- (d) a third polypeptide comprising the binding portion of the variable region of an antibody light chain;
- (e) a fourth polypeptide comprising the binding portion of the variable region of an antibody heavy chain;

- a peptide linker linking said third and fourth polypeptides (f) (d) and (e) into said multivalent protein; and
- a peptide linker linking said second and third (g) polypeptides (b) and (d) into said multivalent protein.

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- A genetic sequence which codes for the multivalent antigen-46. binding protein of claim 44 or 45, comprising:
- a DNA sequence coding for a first polypeptide (a) comprising the binding portion of the variable region of an antibody heavy or light chain;

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- a DNA sequence coding for a second polypeptide (b) comprising the binding portion of the variable region of an antibody heavy or light chain;
- a DNA sequence coding for a peptide linker linking said (c) first and second polypeptides (a) and (b) into said multivalent protein

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a DNA sequence coding for a third polypeptide (d) comprising the binding portion of the variable region of an antibody heavy or light chain;

a DNA sequence coding for a fourth polypeptide (e) comprising the binding portion of the variable region of an antibody heavy or light chain;

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- a DNA sequence coding for a peptide linker linking said **(f)** third and fourth polypeptides (d) and (e) into said multivalent protein; and
- a DNA sequence coding for a peptide linker linking said (g) second and third polypeptides (b) and (d) into said multivalent protein.

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- A replicable cloning or expression vehicle comprising the DNA 47. sequence of any one of claims 38-43.
 - The vehicle of claim 47 which is a plasmid. 48.
 - A host cell transformed with the vehicle of claim 47. 49.

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sequence;

- 50. The host cell of claim 49 which is a bacterial cell, a yeast cell or other fungal cell, or a mammalian cell line.
- 51. A method of producing a multivalent antigen-binding protein comprising two or more single-chain molecules, each single-chain molecule comprising:
- (a) a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (b) a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain; and
- (c) a peptide linker linking said first and second polypeptides
 (a) and (b) into said single-chain molecule, said method comprising:
- (i) providing a genetic sequence coding for said single-chain molecule;
 - (ii) transforming one or more host cells with said
 - (iii) expressing said sequence in said host or hosts;
- (iv) recovering a multivalent protein from said host or hosts.
- 52. A method of producing a multivalent single-chain antigenbinding protein comprising two or more single-chain molecules, each singlechain molecule comprising:
- (a) a first polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (b) a second polypeptide comprising the binding portion of the variable region of an antibody heavy or light chain;
- (c) a peptide linker linking said first and second polypeptides

 (a) and (b) into said multivalent protein;
- (d) a third polypeptide comprising the binding portion of the
 variable region of an antibody heavy or light chain;

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- a fourth polypeptide comprising the binding portion of (e) the variable region of an antibody heavy or light chain; a peptide linker linking said third and fourth polypeptides (f) (d) and (e) into said multivalent protein; and a peptide linker linking said second and third (g) polypeptides (b) and (d) into said multivalent protein, said method comprising: providing a genetic sequence coding for said (i) single-chain molecule; transforming one or more host cells with said (ii) sequence; expressing said sequence in said host or hosts; (iii) and recovering a multivalent protein from said host (iv) or hosts. The method of claim 51 or 52 wherein recovering said 53. multivalent protein comprises separating said multivalent protein from said single-chain molecules. The method of claim 51 or 52 wherein recovering said 54. multivalent protein comprises: dissociating said single-chain molecules; (a) re-associating said single-chain molecules; (b) separating multivalent antigen-binding proteins from said (c) single-chain molecules; and recovering said multivalent proteins. (d)
 - 55. The method of claim 51 or 52 which further comprises purifying said recovered multivalent protein.
 - 56. The method of claim 51 or 52 wherein said host cell is a bacterial cell, a yeast cell or other fungal cell, or a mammalian cell line.

- 57. The method of claim 56 wherein said host cell is E. coli or Bacillus subtilis.
- 58. The multivalent antigen-binding protein of claim 1 in detectably-labelled form.

- 59. In an immunoassay method which utilizes an antibody in detectably-labelled form, the improvement comprising using the multivalent protein of claim 58 instead of said antibody.
- 60. The immunoassay of claim 59 wherein said immunoassay is a competitive immunoassay.

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- 61. The immunoassay of claim 59 wherein said immunoassay is a sandwich immunoassay.
- 62. In an immunotherapeutic method which utilizes an antibody conjugated to a therapeutic agent, the improvement comprising using the multivalent protein of claim 1 instead of said antibody.

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63. In a method of immunoaffinity purification which utilizes an antibody therefor, the improvement which comprises using the molecule of claim 1 instead of said antibody.

WO 93/11161 PCT/US92/09965

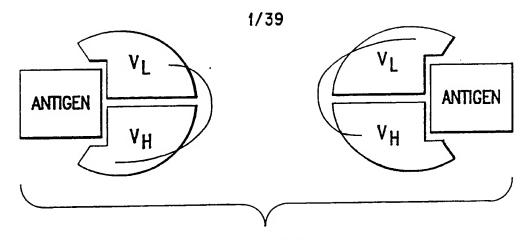


FIG.1A

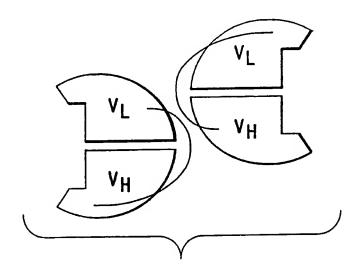


FIG.1B

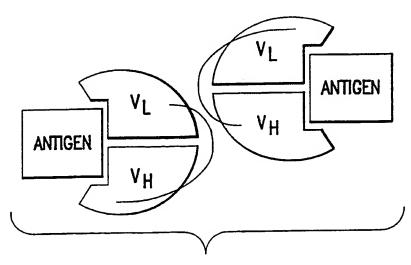
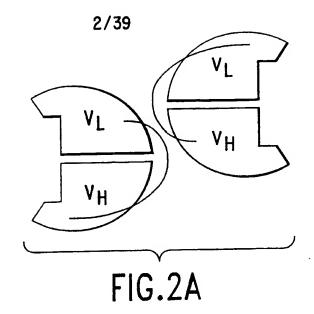
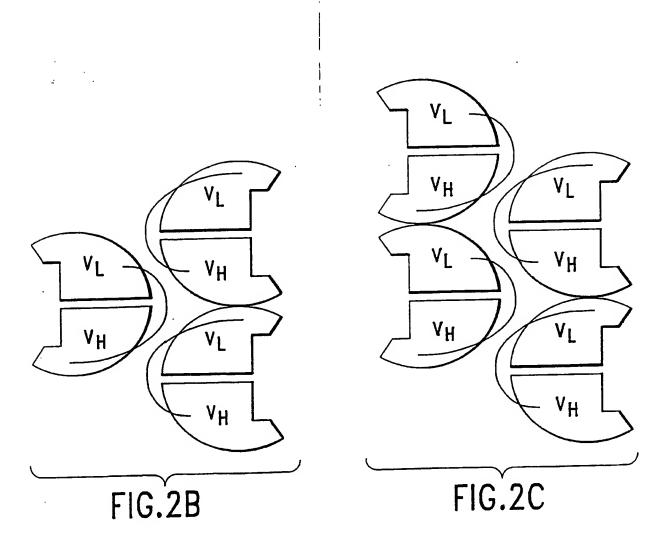
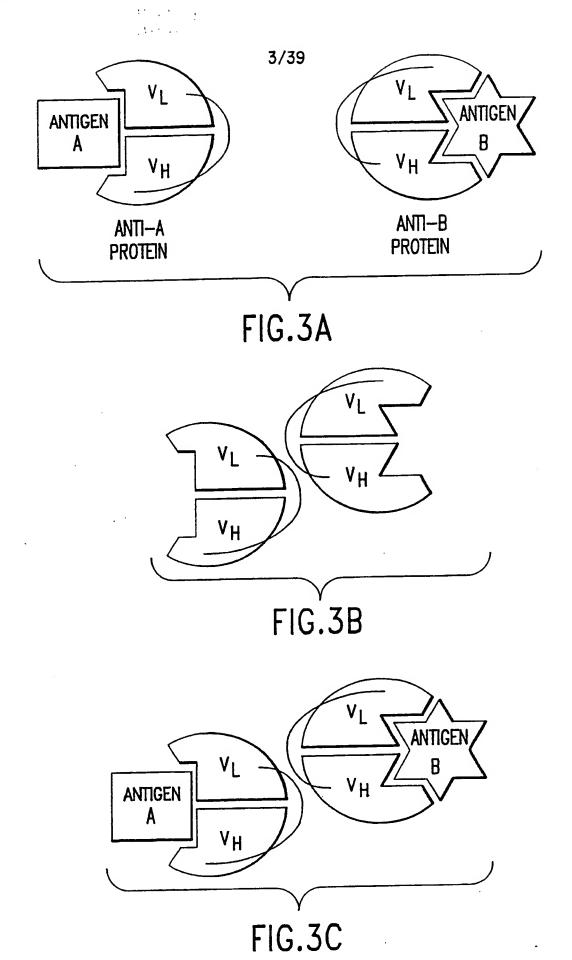


FIG.1C

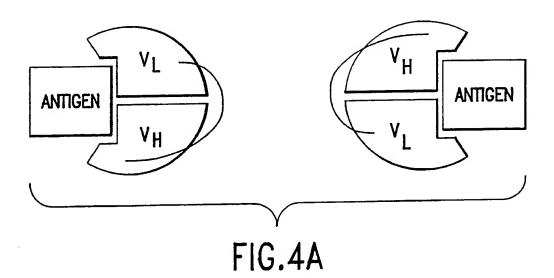
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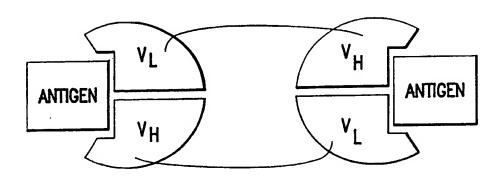
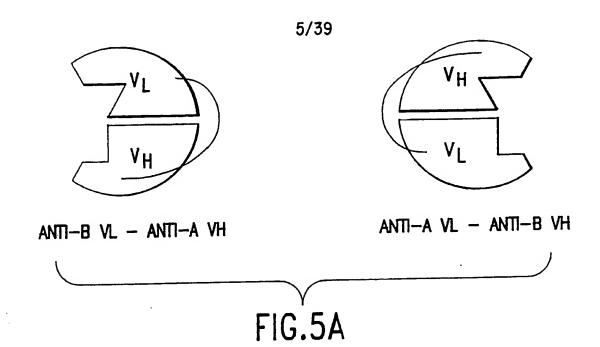


FIG.4B

WO 93/11161 PCT/US92/09965



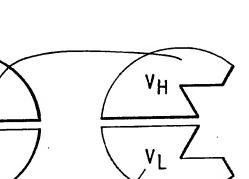
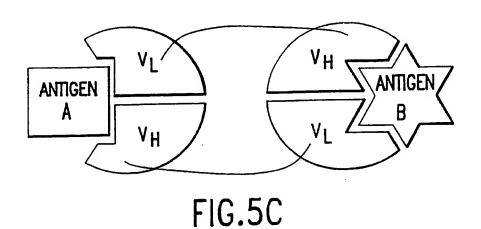


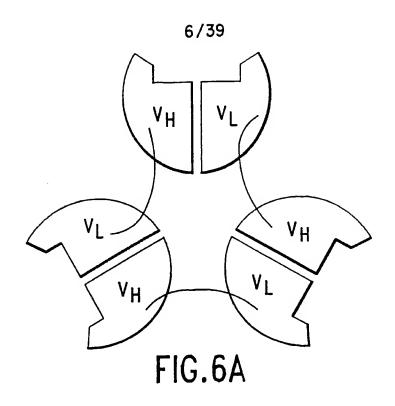
FIG.5B

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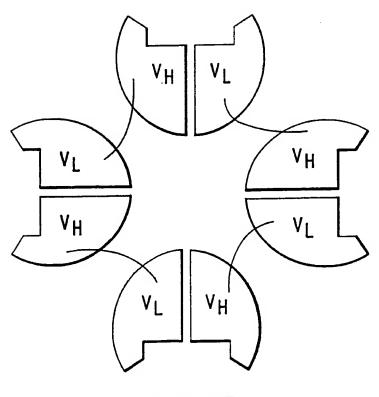
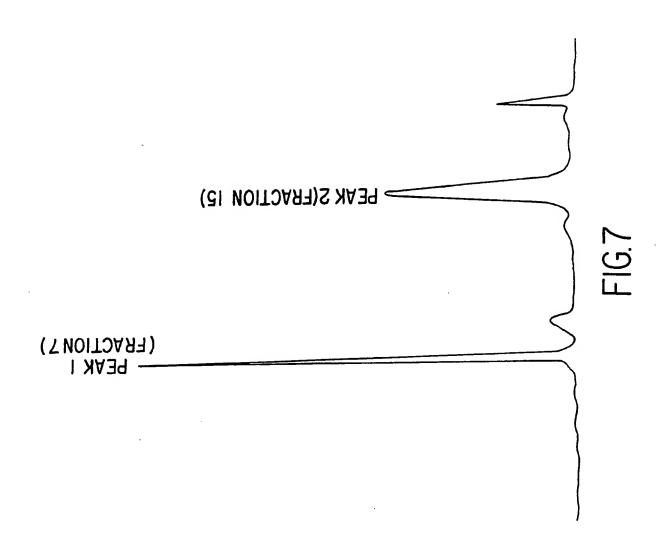
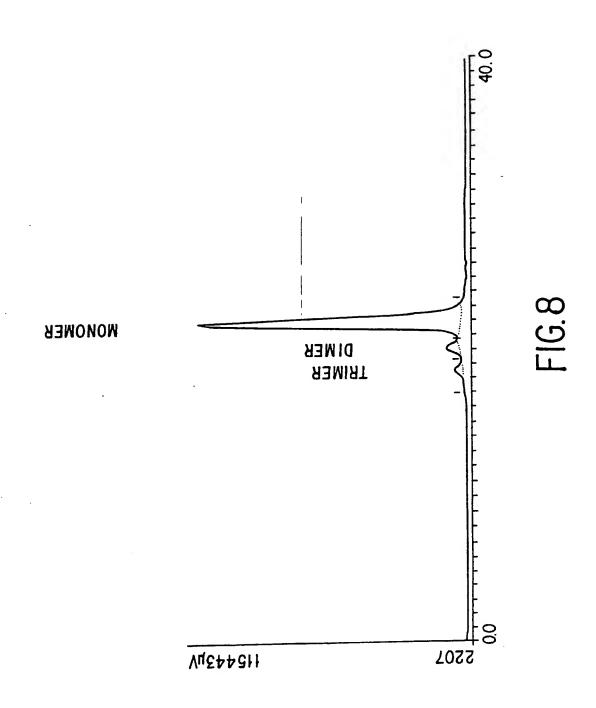


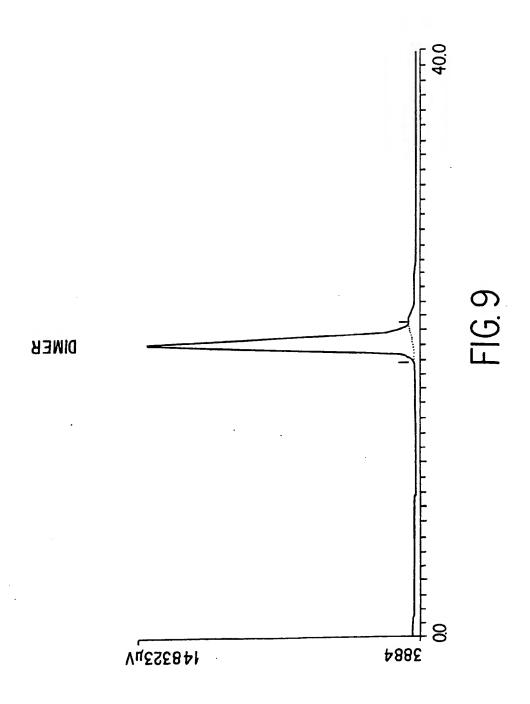
FIG.6B

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4-4-20 VL/212/CC49 VH gene

20 10 4-4-20 VI Asp Val Val Met Thr Gln Thr Pro Leu Ser Leu Pro Val Ser Leu Gly Asp Gln Ala Ser GAC GTC GTT ATG ACT CAG ACA CCA CTA TCA CTT CCT GTT AGT CTA GGT GAT CAA GCC TCC Aat II 40 30 Ile Ser Cys Arg Ser Ser Gln Ser Leu Val His Ser Asn Gly Asn Thr Tyr Leu Arg Trp ATC TCT TGC AGA TCT AGT CAG AGC CTT GTA CAC AGT AAT GGA AAC ACC TAT TTA CGT TGG 60 50 Tyr Leu Gln Lys Pro Gly Gln Ser Pro Lys Val Leu Ile Tyr Lys Val Ser Asn Arg Phe TAC CTG CAG AAG CCA GGC CAG TCT CCA AAG GTC CTG ATC TAC AAA GTT TCC AAC CGA TTT 80 70 Ser Gly Val Pro Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile TCT GGG GTC CCA GAC AGG TTC AGT GGC AGT GGA TCA GGG ACA GAT TTC ACA CTC AAG ATC 100 90 Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Phe Cys Ser Gln Ser Thr His Val Pro AGC AGA GTG GAG GCT GAG GAT CTG GGA GTT TAT TTC TGC TCT CAA AGT ACA CAT GTT CCG 212 Linker 110 Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys Gly Ser Thr Ser Gly Ser Gly Lys TGG ACG TTC GGT GGA GGC ACC AAG CTT GAA ATC AAA GGT TCT ACC TCT GGT TCT GGT AAA Hind III CC49 VH 130 Ser Ser Glu Gly Lys Gly Gln Val Gln Leu Gln Gln Ser Asp Ala Glu Leu Val Lys Pro TCC TCT GAA GGC AAA GGT CAG GTT CAG CTG CAG CAG TCT GAC GCT GAG TTG GTG AAA CCT Pvull Pstl 160 150 Gly Ala Ser Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asp His Ala Ile GGG GCT TCA GTG AAG ATT TCC TGC AAG GCT TCT GGC TAC ACC TTC ACT GAC CAT GCA ATT 180 170 His Trp Val Lys Gln Asn Pro Glu Gln Gly Leu Glu Trp Ile Gly Tyr Phe Ser Pro Gly CAC TGG GTG AÁA CAG AAC CCT GAA CAG GGC CTG GAA TGG ATT GGA TAT TTT TCT CCC GGA

FIG.10A

Asn Asp Asp Phe Lys Tyr Asn Glu Arg Phe Lys Gly Lys Ala Thr Leu Thr Ala Asp Lys AAT GAT GAT TTT AAA TAC AAT GAG AGG TTC AAG GGC AAG GCC ACA CTG ACT GCA GAC AAA

210

Ser Ser Ser Thr Ala Tyr Val Gln Leu Asn Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr TCC TCC AGC ACT GCC TAC GTG CAG CTC AAC AGC CTG ACA TCT GAG GAT TCT GCA GTG TAT

TTC TGT ACA AGA TCC CTG AAT ATG GCC TAC TGG GGT CAA GGA ACC TCA GTC ACC GTC TCC

TTC TGT ACA AGA TCC CIG AAI AIG GCL TAC 100 GGT CAA GGA ACC TCA GTC ACC GTC TCC
Phe Cys Thr Arg Ser Leu Asn Met Ala Tyr Trp Gly Gln Gly Thr Ser Val Thr Val Ser

*** *** Asp TAA TAG GAT CC Bam H1

4-4-20 V_I /212/CC49 V_H gene

FIG.10A(CONT.)

CC49 VL/212/4-4-20 VH gene

CC49 Asp GAC Aat	Va T GTC	Va I GTG	Met ATG	Ser TCA	Gln CAG	Ser TCT	Pro CCA	Ser TCC	10 Ser TCC	Leu CTA	Pro CCT	Val GTG	Ser TCA	Val GTT	G l y GGC	Glu GAG	Lys AAG	Val GTT	20 Thr ACT
ا ام	Spr	Cys TGC	Lys AAG	Ser TCC	Ser AGT	Gln CAG	Ser AGC	Leu CTT	30 Leu TTA	Tyr TAT	Ser AGT	Gly GGT	Asn AAT	Gln CAA	Lys AAG	Asn AAC	Tyr TAC	Leu TTG	40 Ala GCC
Trp TGG	Tyr TAC	Gln CAG	Gln CAG	Lys AAA	Pro CCA	Gly GGG	Gln CAG	Ser TCT	50 Pro CCT	Lys AAA	Leu CTG	Leu CTG	lle ATT	Tyr TAC	Trp TGG	Ala GCA	Ser TCC	Ala GCT	60 Arg AGG
Glu GAA	Ser TCT	Gly GGG	Val GTC	Pro CCT	Asp GAT	Arg CGC	Phe TTC	Thr ACA	70 Gly GGC	Ser AGT	Gly GGA	Ser TCT	Gly GGG	Thr ACA	Asp GAT	Phe TTC	Thr ACT	Leu CTC	80 Ser TCC
I le ATC	Ser AGC	Ser AGT	Val GTG	Lys AAG	Thr ACT	Glu GAA	Asp GAC	Leu CTG	90 Ala GCA	Val GTT	Tyr TAT	Tyr TAC	Cys TGT	Gln CAG	Gln CAG	Tyr TAT	Tyr TAT	Ser AGC	100 Tyr TAT
Pro CCC	Leu CTC	Thr ACG	Phe TTC	Gly GGT	Ala GCT	Gly GGG	Thr ACC	AAG	110 Leu CTT	Va I GTG	Leu CTG	Lys AAA	Gly	Link Ser TCT	Thr	Ser	Gly GGT	Ser AGC	GGC GGC
Lys AAA	Ser TCT	Ser TCT	Glu GAA	Gly GGT	Lys AAA	G l y GGT	Glu	-20 \ Val	/H Lys	Leu	Asp GAT	Glu GAG	Thr ACT	Gly GGA	Gly GGA	Gly GGC	Leu TTG	Val GTG	140 Gln CAA
Pro CCT	Gly GGG	Arg AGG	Pro CCC	Met ATG	Lys AAA	Leu CTC	Ser TCC	Cys TGT	150 Val GTT	Ala	Ser TCT	Gly GGA	Phe TTC	Thr ACT	Phe TTT	Ser AGT	Asp GAC	Tyr TAC	160 Trp TGG
Met ATG	Asn AAC	Trp TGG	Va l GTC	Arg CGC	Gln CAG	Ser TCT	Pro CCA	Glu GAG	170 Lys AAA	Glv	Leu CTG	Glu GAG	Trp TGG	Val GTA	Ala GCA	Gln CAA	Ile ATT	Arg AGA	180 Asn AAC

FIG.10B

Lys Pro Tyr Asn Tyr Glu Thr Tyr Tyr Ser Asp Ser Val Lys Gly Arg Phe Thr Ile Ser AAA CCT TAT AAT TAT GAA ACA TAT TAT TCA GAT TCT GTG AAA GGC AGA TTC ACC ATC TCA

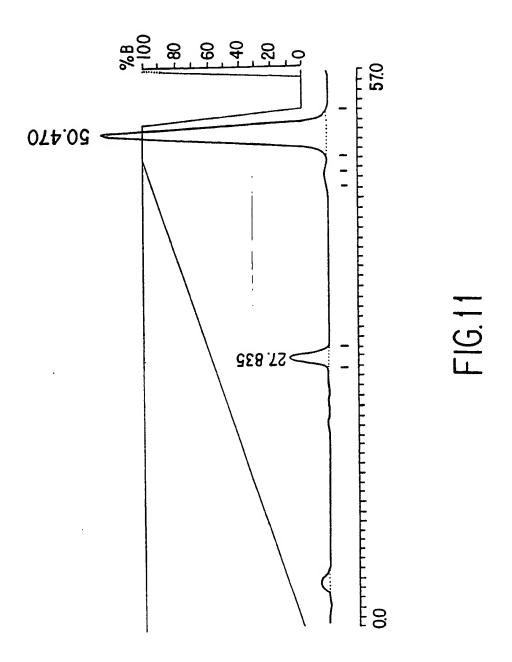
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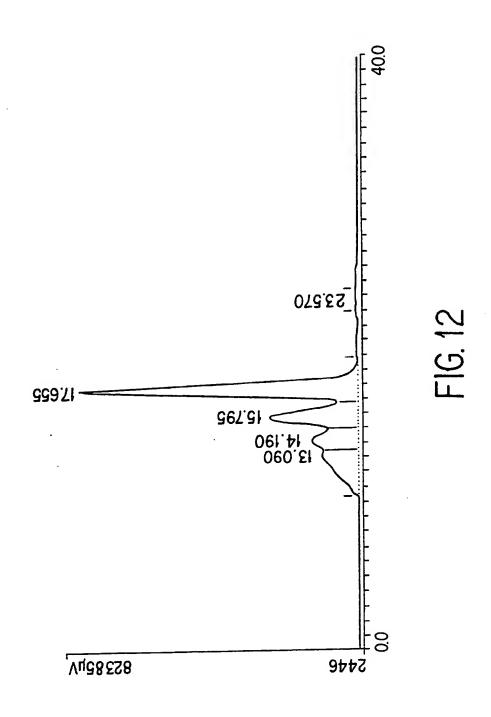
Gly Ile Tyr Tyr Cys Thr Gly Ser Tyr Tyr Gly Met Asp Tyr Trp Gly Gln Gly Thr Ser GGT ATC TAT TAC TGT ACG GGT TCT TAC TAT GGT ATG GAC TAC TGG GGT CAA GGA ACC TCA

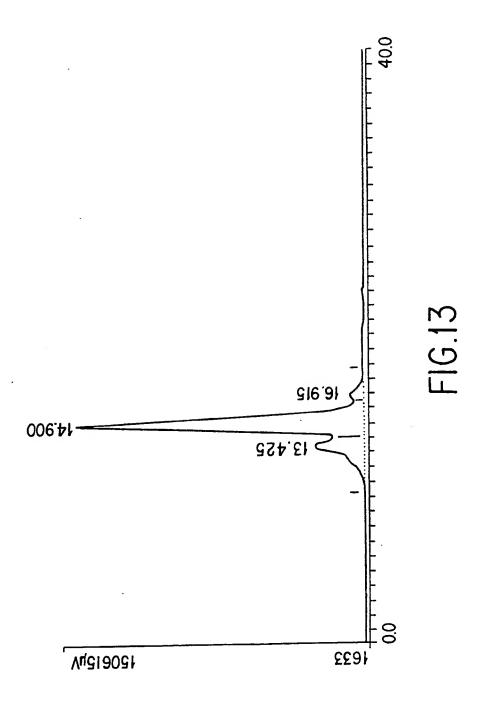
FIG.10B(CONT.)

Val Thr Val Ser * * Gly Ser GTC ACC GTC TCC TAA TAA GGA TCC

Bam H1







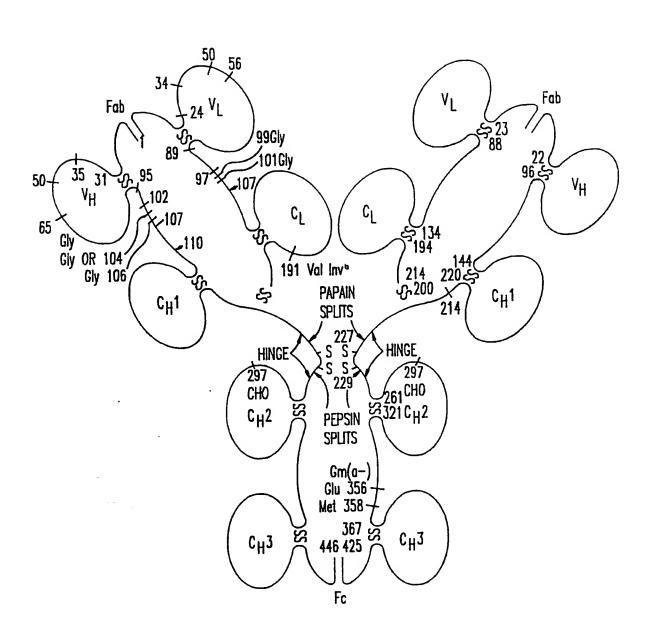


FIG.14

4-4-20/212

protein with single cysteine hinge

Asp GAC	GTC	Val	Met ATG	Thr ACT	Gln CAG	Thr ACA	Pro CCA	Leu CTA	10 Ser TCA	Leu CTT	Pro CCT	Val GTT	Ser AGT	Leu CTA	Gly GGT	Asp GAT	Gln CAA	A la GCC	20 Ser TCC
Aat Ile ATC	Ser	Cys TGC	Arg AGA	Ser TCT	Ser AGT	Gln CAG	Ser AGC	Leu CTT	30 Val GTA	H1s CAC	Ser AGT	Asn AAT	Gly GGA	Asn AAC	Thr ACC	Tyr TAT	Leu TTA	Arg CGT	40 Trp TGG
Tyr TAC	Leu CTG	G In CAG	Lys AAG	Pro CCA	Gly GGC	Gln CAG	Ser TCT	Pro CCA	50 Lys AAG	Val GTC	Leu CTG	Ile ATC	Tyr TAC	Lys AAA	Val GTT	Ser TCC	Asn AAC	Arg CGA	60 Phe TTT
Ser TCT	Gly GGG	Va I GTC	Pro CCA	Asp GAC	Arg AGG	Phe TTC	Ser AGT	Gly GGC	70 Ser AGT	Gly GGA	Ser TCA	Gly GGG	Thr ACA	Asp GAT	Phe TTC	Thr ACA	Leu CTC	Lys AAG	08 Ile ATC
Ser AGC	Arg AGA	Va I GTG	Glu GAG	Ala GCT	Glu GAG	A sp GAT	Leu CTG	Gly GGA	90 Val GTT	Tyr TAT	Phe TTC	Cys TGC	Ser TCT	Gln CAA	Ser AGT	Thr ACA	His CAT	Va l GTT	100 Pro CCG
Trp TGG	Thr ACG	Phe TTC	Gly GGT	Gly GGA	Gly GGC	Thr ACC	AAG	CTT	GAA	Ile ATC	Lys AAA	Gly	Lini Ser TCT	Thr	Ser	Gly	Ser TCT	Gly GGT	120 Lys AAA
Ser TCT	Ser TCT	Glu GAA	Gly GGT	Lys AAA	Gly GGT	Glu	1-20 Val	Lys	Leu	Asp GAT	Glu GAG	Thr ACT	G l y GGA	Gly GGA	Gly GGC	Leu TTG	Val GTG	Gln CAA	140 Pro CCT
Gly GGG	Arg AGG	Pro CCC	Met ATG	Lys AAA	Leu CTC	Ser TCC	Cys TGT	Val GTT	150 Ala GCC	Ser TCT	Gly GGA	Phe TTC	Thr ACT	Phe TTT	Ser AGT	Asp GAC	Tyr TAC	Trp TGG	160 Met ATG
Asn AAC	Trp TGG	Val GTC	Arg CGC	Gln CAG	Ser TCT	Pro CCA	Glu GAG	Lys AAA	170 Gly GGA	Leu CTG	Glu GAG	Trp TGG	Va l GTA	Ala GCA	Gln CAA	Ile ATT	Arg AGA	Asn AAC	180 Lys AAA

FIG.15A

4-4-20/212

protein with single cysteine hinge

200 190 Pro Tyr Asn Tyr Glu Thr Tyr Tyr Ser Asp Ser Val Lys Gly Arg Phe Thr Ile Ser Arg CCT TAT AAT TAT GAA ACA TAT TAT TCA GAT TCT GTG AAA GGC AGA TTC ACC ATC TCA AGA 220 210 Asp Asp Ser Lys Ser Ser Val Tyr Leu Gin Met Asn Asn Leu Arg Val Glu Asp Met Gly GAT GAT TCC AAA AGT AGT GTC TAC CTG CAA ATG AAC AAC TTA AGA GTT GAA GAC ATG GGT 240 530 Ile Tyr Tyr Cys Thr Gly Ser Tyr Tyr Gly Met Asp Tyr Trp Gly Gln Gly Thr Ser Val ATC TAT TAC TGT ACG GGT TCT TAC TAT GGT ATG GAC TAC TGG GGT CAA GGA ACC TCG GTC Bst Ell 250 Hinge Thr Val Ser Ser Asp Lys Thr His Thr Cys *** *** ACC GTC TCC AGT GAT AAG ACC CAT ACA TGC TAA TAG GAT CC Ban Hi pGx 5532, Gx 8932

FIG.15A(CONT.)

4-4-20/212 protein with two cysteine hinge

4-4- Asp	Val	Val	Met	Thr	Gln CAG	Thr ACA	Pro CCA	Leu CTA	10 Ser TCA	Leu CTT	Pro CCT	Val GTT	Ser AGT	Leu CTA	Gly GGT	Asp GAT	Gln CAA	Ala GCC	20 Ser TCC
Aat	II	Cue	۸۸۵	San	Sar.	Gla	Cor	l eu	30 Vo I	His	Ser	Asn	Glv	Asn	Thr	Tyr	Leu	Arg	40 Trp
T	امدا	Cla	Luc	Dno	Glv.	Cln.	cor.	Pro	50 I vs	CAC	l eu	Ile	Tvr	Lvs	Val	Ser	Asn	Arg	60 Phe
TAC	CTG	CAG	AAG	CCA	GGC	CAG	TCT	CCA	AAU 70	GIC	UIU	AIC	IAL	AAA	uII	166	MHL	LUA	80
Ser TCT	Gly GGG	Va l GTC	Pro CCA	Asp GAC	Arg AGG	Phe TTC	Ser AGT	Gly	Aul	¦Gly GGA 	Ser TCA	Gly GGG	Thr ACA	Asp GAT	Phe TTC	Thr ACA	Leu CTC	Lys AAG	Ile ATC 100
Ser AGC	Arg AGA	Val GTG	Glu GĄG	Ala GCT	Glu GAG	Asp GAT	Leu CTG	Gly GGA	90 Val GTT	Tyr TAT	Phe TTC	Cys TGC	Ser TCT	Gln CAA	Ser AGT	Thr ACA	His CAT	Val GTT	Pro
Ton	Thn	Pho	Glv	Glv	Glv	Thr	l vs	l eu	110 Glu	Ile	Lvs	212 Gly	Linl Ser	ker Thr	Ser	Gly	Ser	Gly	120 Lys
TGG	ACG	TTC	GGT	GGA	GGC	ACC	AAG	CTT	GAA	ATC	AAA	GGT	TCT	ACC	TCT	GGT	TCT	GGT	AAA
Ser TCT	Ser TCT	Glu GAA	Gly	Lys AAA	Gly	Glu	-20 Val	Y _H Lvs	130 Leu	Asp	Glu GAG	Thr ACT	Gly GGA	Gly GGA	Gly GGC	Leu TTG	Val GTG	Gin CAA	Pro CCT
Gly	Arg	Pro	Met	Lys	Leu	Ser	Cys TGT	Val	150 Ala	Ser	Gly	Phe TTC	Thr ACT	Phe TTT	Ser AGT	Asp GAC	Tyr TAC	Trp TGG	160 Met ATG
۸cn	Twn	Val	Δνα	Gln	Sor	Pro	նև	l vs	170 Glv	Leu	Glu	Trp	Yal	Ala	Gln	Ile	Arg	Asn	180 Lys
AAC	TGG	GTC	CGC	CAG	TCT	CCA	GAG	AAA	GGA	CTG	GAG	TGG	GTA	GCA	CAA	ATT	AGĀ	AAC	AAA

FIG.15B

4-4-20/212 protein with two cysteine hinge

Pro CCT	Tyr TAT	Asn AAT	Tyr TAT	Glu GAA	Thr ACA	Tyr TAT	Tyr TAT	Ser TCA	190 Asp GAT	Ser TCT	Val GTG	Lys AAA	Gly GGC	Arg AGA	Phe TTC	Thr ACC	Ile ATC	Ser TCA	200 Arg AGA
Asp GAT	Asp GAT	Ser TCC	Lys AAA	Ser AGT	Ser AGT	Val GTC	Tyr TAC	Leu CTG	210 Gln CAA	Met ATG	Asn AAC	Asn AAC	Leu TTA	Arg AGA	Val GTT	Glu GAA	Asp GAC	Met ATG	220 Gly GGT
Ile ATC	Tyr TAT	Tyr TAC	Cys TGT	Thr ACG	Gly GGT	Ser TCT	Tyr TAC	Tyr TAT	230 Gly GGT	Met ATG	Asp GAC	Tyr TAC	Trp TGG	Gly GGT	Gln CAA	Gly GGA	Thr ACC	Ser TCG Bst	240 Val GTC E11
The	Val	San.	Hing	ge Asn	Lve	The	Hic		250 Cvs	Pro	Pro	Суѕ	***	¥¥¥					
ACC	GTC	TCC	AGT	GAT	AAG	ACC	CAT	ACA	TGC	CCT	CCA	TGC	TAA	TAG	GAT	<u>CC</u>			
рбх	5533	3, G	8 93	33										В	am Hi				

FIG.15B(CONT.)

CC49/212 SCATM protein genetic dimer

CC49 VL Asp Val Val M GAC GTC GTG A	let Ser Gli ITG_TCA CA	Ser Pro	Ser Ser TCC TCC	Leu Pro	Val Ser GTG TCA	Val Gly GTT GGC	Glu Lys GAG AAC	20 Val Thr GTT ACT
Aat II Leu Ser Cys L TTG AGC TGC A	.ys Ser Se AG TCC AG	Gln Ser CAG AGC	30 Leu Leu CTT TTA	Tvr Ser	Gly Asr GGT AA1	Gln Lys CAA AAG	Asn Tyr AAC TAC	40 Leu Ala TTG GCC
Trp Tyr Gln G TGG TAC CAG C	iln Lys Pr CAG AAA CC	o Gly Gln A GGG CAG	50 Ser Pro TCT CCT	Lvs Leu	Leu Ile CTG ATT	Tyr Trp	Ala Ser GCA TCI	60 Ala Arg GCT AGG
Glu Ser Gly V GAA TCT GGG G	ol Pro As TC CCT GA	Arg Phe CGC TTC	70 Thr Gly ACA GGC	Ser Glv	Ser Gly	/ Thr Asp G ACA GAT	Phe The	80 Leu Ser I CTC TCC
Ile Ser Ser V ATC AGC AGT G	/al Lys Th GTG AAG AC	~ Glu Asp T GAA GAC	90 Leu Ala CTG GCA	Val Tvr	Tyr Cys	s Gln Gln F CAG CAC	Tyr Tyi i TAT TA	100 Ser Tyr I AGC TAT
Pro Leu Thr F	Phe Gly Al	a Gly Thr T GGG ACC	Lys Leu AAG CTT Hind II	Val Leu GTG CTG	Lys Gly	2 Linker / Ser Thr C TCT ACT	Ser Gl	120 y Ser Gly T AGC GGC
Lys Ser Ser C AAA TCC TÇT C	ilu Gly Ly IAA GGC AA	s Glv Gln	9 VH Val Gln GTT <u>CAG</u>	Leu Gln	Gln Ser CAG TC	r Asp Alo r GAC GC1	Glu Lei GAG TT	140 u Val Lys G GTG AAA
Pro Gly Ala S CCT GGG GCT 1	Ser Val Ly ICA GTG AA	s Ile Ser G ATT TCC	150 Cvs Lvs	l : Ala Ser	Gly Tyr	r Thr Phe C ACC TTO	Thr Asp C ACT GA	160 HIS Ala C CAT GCA
Ile His Trp \ ATT CAC TGG (Val Lys Gl GTG AAA CA	n Asn Pro G AAC CCT	170 Glu Gln GAA CAG	Glv Leu	Glu Tri GAA TG	o Ile Gly G ATT GG	y Tyr Ph A TAT TT	180 e Ser Pro T TCT CCC

FIG.16A

CC49/212 SCATM protein genetic dimer 200 190 Gly Asn Asp Asp Phe Lys Tyr Asn Glu Arg Phe Lys Gly Lys Ala Thr Leu Thr Ala Asp GGA AAT GAT GAT TTT AAA TAC AAT GAG AGG TTC AAG GGC AAG GCC ACA CTG ACT GCA GAC 550 210 Lys Ser Ser Ser Thr Ala Tyr Val Gln Leu Asn Ser Leu Thr Ser Glu Asp Ser Ala Val AAA TCC TCC AGC ACT GCC TAC GTG CAG CTC AAC AGC CTG ACA TCT GAG GAT TCT GCA GTG 530 Tyr Phe Cys Thr Arg Ser Leu Asn Met Ala Tyr Trp Gly Gln Gly Thr Ser Val Thr Val TAT TTC TGT ACA AGĂ TCC CTG AAT ATG GCC TAC TGG GGT CAA GGA ACC TCA GTC ACC GTC 260 250 CC49 VL Ser Ser Asp Val Val Met Ser Gln Ser Pro Ser Ser Leu Pro Val Ser Val Gly Glu Lys TCC TCA GAC GTC GTG ATG TCA CAG TCT CCA TCC TCC CTA CCT GTG TCA GTT GGC GAG AAG Aat II 280 270 Val Thr Leu Ser Cys Lys Ser Ser Gln Ser Leu Leu Tyr Ser Gly Asn Gln Lys Asn Tyr GTT ACT TTG AGC TGC AAG TCC AGT CAG AGC CTT TTA TAT AGT GGT AAT CAA AAG AAC TAC 300 290 Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln Ser Pro Lys Leu Leu Ile Tyr Trp Ala Ser TTG GCC TGG TAC CAG CAG AAA CCA GGG CAG TCT CCT AAA CTG CTG ATT TAC TGG GCA TCC 350 310 Ala Arg Glu Ser Gly Val Pro Asp Arg Phe Thr Gly Ser Gly Ser Gly Thr Asp Phe Thr GCT AGG GAA TCT GGG GTC CCT GAT CGC TTC ACA GGC AGT GGA TCT GGG ACA GAT TTC ACT 340 330 Leu Ser Ile Ser Ser Val Lys Thr Glu Asp Leu Ala Val Tyr Tyr Cys Gln Gin Tyr Tyr CTC TCC ATC AGC AGT GTG AAG ACT GAA GAC CTG GCA GTT TAT TAC TGT CAG CAG TAT TAT 360 212 Linker 350 Ser Tyr Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Val Leu Lys Gly Ser Thr Ser Gly AGC TAT CCC CTC ACG TTC GGT GCT GGG ACC AAG CTT GTG CTG AAA GGC TCT ACT TCC GGT Hind III 380 CC49 VH Ser Gly Lys Ser Ser Glu Gly Lys Gly Gln Val Gln Leu Gln Gln Ser Asp Ala Glu Leu AGC GGC AAA TCC TCT GAA GGC AAA GGT CAG GTT CAG CTG CAG CAG TCT GAC GCT GAG TTG

FIG.16B

PvuII PstI

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CC49/212 SCATM protein genetic dimer

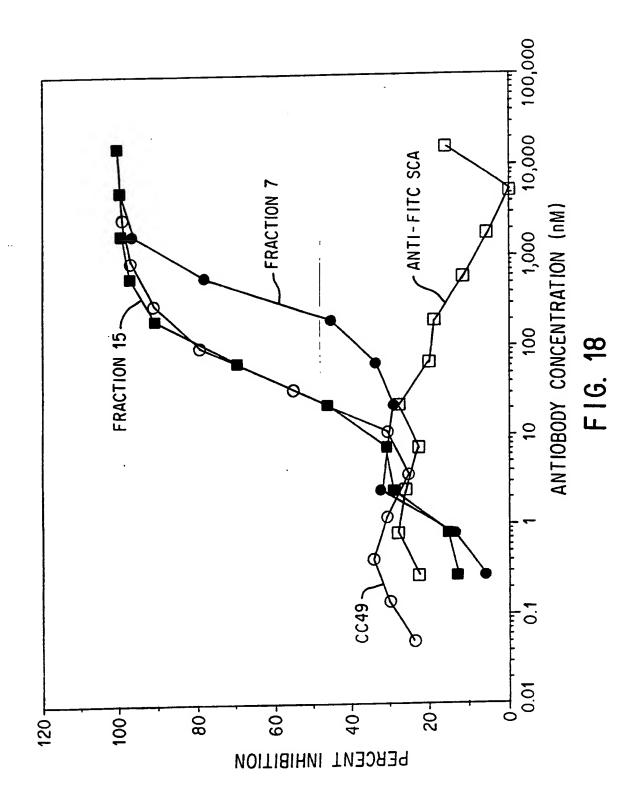
			•			-													
Val GTG	Lys AAA	Pro CCT	G l y GGG	Ala GCT	Ser TCA	Va I GTG	Lys AAG	Ile ATT	390 Ser TCC	Cys TGC	Lys AAG	Ala GCT	Ser TCT	Gly GGC	Tyr TAC	Thr ACC	Phe TTC	Thr ACT	400 Asp GAC
His CAT	Ala GCA	Ile ATT	HIS CAC	Trp TGG	Val GTG	Lys AAA	Gln CAG	Asn AAC	410 Pro CCT	Glu GAA	Gln CAG	Gly GGC	Leu CTG	Glu GAA	Trp TGG	Ile ATT	Gly GGA	Tyr TAT	420 Phe TTT
Ser TCT	Pro CCC	Gly GGA	Asn AAT	Asp GAT	Asp GAT	Phe TTT	Lys AAA	Tyr TAC	430 Asn AAT	Glu GAG	Arg AGG	Phe TTC	Lys AAG	Gly GGC	Lys AAG	Ala GCC	Thr ACA	Leu CTG	440 Thr ACT
Ala GCA	Asp GAC	Lys AAA	Ser TCC	Ser TCC	Ser AGC	Thr ACT	Ala GCC	Tyr TAC	450 Val GTG	G I n CAG	Leu CTC	Asn AAC	Ser AGC	Leu CTG	Thr ACA	Ser TCT	Glu GAG	Asp GAT	460 Ser TCT
Ala GCA	Val GTG	Tyr TAT	Phe TTC	Cys TGT	Thr ACA	Arg AGA	Ser TCC	Leu CTG	470 Asn AAT	Met ATG	Ala GCC	Tyr TAC	Trp TGG	Gly GGT	Gln CAA	Gly GGA	Thr ACC	Ser TCA	480 Val GTC

Thr Val Ser *** *** Asp ACC GTC TCC TAA TA<u>G GAT CC</u> Ban H1

FIG.16C

1 2 3

FIG. 17



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4-4-20 VL/217/CC49 VH gene

Asp GAC	GTC	Val	Met ATG	Thr ACT	Gln CAG	Thr ACA	Pro CCA	Leu CTA	10 Ser TCA	Leu CTT	Pro CCT	Val GTT	Ser AGT	Leu CTA	Gly GGT	Asp GAT	Gln CAA	Ala GCC	20 Ser TCC
Aat Ile ATC	Ser	Cys TGC	Arg AGA	Ser TCT	Ser AGT	Gln CAG	Ser AGC	Leu CTT	30 Val GTA	His	Ser AGT	Asn AAT	Gly GGA	Asn AAC	Thr ACC	Tyr TAT	Leu TTA	Arg CGT	40 Trp TGG
Tyr TAC	Leu CTG	Gln CAG	Lys AAG	Pro CCA	Gly GGC	Gln CAG	Ser TCT	Pro CCA	50 Lys AAG	Val GTC	Leu CTG	Ile ATC	Tyr TAC	Lys AAA	Val GTT	Ser TCC	Asn AAC	Arg CGA	60 Phe TTT
Ser TCT	Gly GGG	Val GTC	Pro CCA	Asp GAC	Arg AGG	Phe TTC	Ser AGT	Gly GGC	70 Ser AGT	Gly GGA	Ser TCA	Gly GGG	Thr ACA	Asp GAT	Phe TTC	Thr ACA	Leu CTC	Lys AAG	08 Ile OTA
Ser AGC	Arg AGA	Va l GTG	Glu GAG	A la GCT	Glu GAG	Asp GAT	Leu CTG	Gly GGA	90 Val GTT	Tyr TAT	Phe TTC	Cys TGC	Ser TCT	Gln CAA	Ser AGT	Thr ACA	His CAT	Val GTT	100 Pro CCG
Trp TGG	Thr ACG	Phe TTC	Gly GGT	Gly GGA	Gly GGC	Thr ACC	AAG	Leu TTO	GAA	I le ATC	Lys AAA	217 Gly GGT	Ser	Thr	Ser TCT	Gly GGT	Lys AAA	Pro CCA	120 Ser TCT
Glu GAA	Gly	Lys AAA	Gly GGT	film	Val GTT	Gln CAG	Leu	Gln CAG	130 Gln	Ser TCT	Asp GAC	Ala GCT	Glu GAG	Leu TTG	Val GTG	Lys AAA	Pro CCT	Gly GGG	140 Ala GCT
Ser TCA	Va I GTG	Lys AAG	I le ATT	Ser TCC	Cys TGC	Lvs	Ala	Ser	150 Gly GGC	Tyr TAC	Thr ACC	Phe TTC	Thr ACT	Asp GAC	His CAT	Ala GCA	Ile ATT	His CAC	160 Trp TGG
Val GTG	Lys AAA	Gln CAG	Asn AAC	Pro CCT	Glu GAA	Gln CAG	Gly GGC	Leu CTG	170 Glu GAA	Trp TGG	Ile ATT	Gly GGA	Tyr TAT	Phe TTT	Ser TCT	Pro CCC	Gly GGA	Asn AAT	180 Asp GAT

FIG.19A

Asp Phe Lys Tyr Asn Glu Arg Phe Lys Gly Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser GAT TTT AAA TAC AAT GAG AGG TTC AAG GGC AAG GCC ACA CTG ACT GCA GAC AAA TCC TCC

Ser Thr Ala Tyr Val Gln Leu Asn Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Phe Cys AGC ACT GCC, TAC GTG CAG CTC AAC AGC CTG ACA TCT GAG GAT TCT GCA GTG TAT TTC TGT

230

Thr Arg Ser Leu Asn Met Ala Tyr Trp Gly Gln Gly Thr Ser Val Thr Val Ser ****

ACA AGA TCC CTG AAT ATG GCC TAC TGG GGT CAA GGA ACC TCA GTC ACC GTC TCC TAA TAG

Asp GAT CC Bom H1

FIG.19A(CONT.)

CC49 $V_L/217/4-4-20 V_H$ gene

10																				
Leu Ser Cys Lys Ser Ser Gln Ser Leu Leu Tyr Ser Gly Asn Gln Lys Asn Tyr Leu Ala TTG AGC TGC AAG TCC AGT CAG AGC CTT TTA TAT AGT GGT AAT CAA AAG AAC TAC TTG GCC 50 Trp Tyr Gln Gln Lys Pro Gly Gln Ser Pro Lys Leu Leu I le Tyr Trp Ala Ser Ala Arg TGG TAC CAG CAG CAG AAA CCA GGG CAG TCT CCT AAA CTG CTG ATT TAC TGG GCA TCC GCT AGG Glu Ser Gly Val Pro Asp Arg Phe Thr Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Ser GAA TCT GGG GTC CCT GAT CGC TTC ACA GGC AGT GGA TCT GGG ACA GAT TTC ACT CTC TCC 90 110 11e Ser Ser Val Lys Thr Glu Asp Leu Ala Val Tyr Tyr Cys Gln Gln Tyr Tyr Ser Tyr ATC AGC AGT GTG AAG ACT GAA GAC CTG GCA GTT TAT TAC TGT CAG CAG TAT TAT AGC TAT Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Val Leu Lys Gly Ser Thr Ser Gly Lys Pro CCC CTC ACG TTC GGT GGT GCT GGG ACC AAG CTT GTG CTG AAA GGC TTG GGT CAA CCA HIND III 4-4-20 Vh 130 140 Ser Glu Gly Lys Gly Glu Val Lys Leu Asp Glu Thr Gly Gly Gly Leu Val Gln Pro Gly TCT GAA GGT AAA GGT AAA GGT GAA GTT GAA GGT GAA GTT GAA GGT GAA GTT GAA GGT AAA CCA GGT AAA GGT GAA GGT GAA GGT GAA GTT GGT GG	Asp GAC	Va l GTC	Va I GTG	Met ATG	Ser TCA	G In CAG	Ser TCT	Pro CCA	Ser TCC	Ser	Leu CTA	Pro CCT	Val GTG	Ser TCA	Val GTT	G l y GGC	Glu GAG	Lys AAG	Val GTT	Thr
Trp Tyr Gin Gin Lys Pro Gly Gin Ser Pro Lys Leu Leu Ile Tyr Trp Ala Ser Ala Arg TGG TAC CAG CAG AAA CCA GGG CAG TCT CCT AAA CTG CTG ATT TAC TGG GCA TCC GCT AGG Glu Ser Gly Val Pro Asp Arg Phe Thr Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Ser GAA TCT GGG GTC CCT GAT CGC TTC ACA GGC AGT GGA TCT GGG ACA GAT TTC ACT CTC TCC 90 100 Ile Ser Ser Val Lys Thr Glu Asp Leu Ala Val Tyr Tyr Cys Gin Gin Tyr Tyr Ser Tyr ATC AGC AGT GTG AAG ACT GAA GAC CTG GCA GTT TAT TAC TGT CAG CAG TAT TAT AGC TAT ATC AGC AGT GTG AAG ACT GAA GAC CTG GCA GTT TAT TAC TGT CAG CAG TAT TAT AGC TAT Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Val Leu Lys Giy Ser Thr Ser Gly Lys Pro CCC CTC ACG TTC GGT GCT GGG ACC AAG CTT GTG CTG AAA GGC TCT ACT TCC GGT AAA CCA Hind III 4-4-20 VH 130 Ser Glu Gly Lys Gly Glu Val Lys Leu Asp Glu Thr Gly Gly Gly Leu Val Gin Pro Gly TCT GAA GGT AAA GGT GAA GTT AAA CTG GAT GAG ACT GGA GGA GGC TTG GTG CAA CCT GGG Arg Pro Met Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asp Tyr Trp Met Ash AGG CCC ATG AAA CTC TCC TGT GTT GCC TCT GGA TTC ACT TTT AGT GAC TAC TGG ATG AAC 170 180 Trp Val Arg Gin Ser Pro Glu Lys Gly Leu Glu Trp Val Ala Gln Ile Arg Asn Lys Pro	Lou	20r	Cys TGC	Lys AAG	Ser TCC	Ser AGT	Gln CAG	Ser AGC	Leu CTT	Leu	Tyr TAT	Ser AGT	Gly GGT	Asn AAT	Gln CAA	Lys AAG	Asn AAC	Tyr TAC	Leu TTG	Ala
Glu Ser Gly Val Pro Asp Arg Phe Thr Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Ser GAA TCT GGG GTC CCT GAT CGC TTC ACA GGC AGT GGA TCT GGG ACA GAT TTC ACT CTC TCC 90 100 Ile Ser Ser Val Lys Thr Glu Asp Leu Ala Val Tyr Tyr Cys Gln Gln Tyr Tyr Ser Tyr ATC AGC AGT GTG AAG ACT GAA GAC CTG GCA GTT TAT TAC TGT CAG CAG TAT TAT AGC TAT 110 217 Linker 120 Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Val Leu Lys Gly Ser Thr Ser Gly Lys Pro CCC CTC ACG TTC GGT GCT GGG ACC AAG CTT GTG CTG AAA GGC TCT ACT TCC GGT AAA CCA Hind III 4-4-20 VH 130 140 Ser Glu Gly Lys Gly Glu Val Lys Leu Asp Glu Thr Gly Gly Gly Leu Val Gln Pro Gly TCT GAA GGT AAA GGT GAA GTT AAA CTG GAT GAG ACT GGA GGA GGC TTG GTG CAA CCT GGG Arg Pro Met Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asp Tyr Trp Met Asn AGG CCC ATG AAA CTC TCC TGT GTT GCC TCT GGA TTC ACT TTT AGT GAC TAC TGG ATG AAC 170 180 Trp Val Ara Gln Ser Pro Glu Lys Gly Leu Glu Trp Val Ala Gln Ile Arg Asn Lys Pro	Trp TGG	Tyr TAC	Gln CAG	Gln CAG	Lys AAA	Pro CCA	Gly GGG	Gln CAG	Ser TCT	Pro	Lys AAA	Leu CTG	Leu CTG	Ile ATT	Tyr TAC	Trp TGG	Ala GCA	Ser TCC	Ala GCT	Arg
Ile Ser Ser Val Lys Thr Glu Asp Leu Ala Val Tyr Tyr Cys Gln Gln Tyr Tyr Ser Tyr ATC AGC AGT GTG AAG ACT GAA GAC CTG GCA GTT TAT TAC TGT CAG CAG TAT TAT AGC TAT 110 217 Linker 120 Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Val Leu Lys Gly Ser Thr Ser Gly Lys Pro CCC CTC ACG TTC GGT GCT GGG ACC AAG CTT GTG CTG AAA GGC TCT ACT TCC GGT AAA CCA Hind III 4-4-20 VH 130 140 Ser Glu Gly Lys Gly Glu Val Lys Leu Asp Glu Thr Gly Gly Gly Leu Val Gln Pro Gly TCT GAA GGT AAA GGT GAA GTT AAA CTG GAT GAG ACT GGA GGC TTG GTG CAA CCT GGG Arg Pro Met Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asp Tyr Trp Met Asn AGG CCC ATG AAA CTC TCC TGT GTT GCC TCT GGA TTC ACT TTT AGT GAC TAC TGG ATG AAC 170 180 Trp Val Arg Gin Ser Pro Glu Lys Gly Leu Glu Trp Val Ala Gln Ile Arg Asn Lys Pro	Glu GAA	Ser TCT	Gly GGG	Val GTC	Pro CCT	Asp GAT	Arg CGC	Phe TTC	Thr ACA	filv	Ser AGT	Gly GGA	Ser TCT	Gly GGG	Thr ACA	Asp GAT	Phe TTC	Thr ACT	Leu CTC	Ser
Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Val Leu Lys Gly Ser Thr Ser Gly Lys Pro CCC CTC ACG TTC GGT GCT GGG ACC AAG CTT GTG CTG AAA GGC TCT ACT TCC GGT AAA CCA Hind III 4-4-20 VH 130 140 Ser Glu Gly Lys Gly Glu Val Lys Leu Asp Glu Thr Gly Gly Gly Leu Val Gln Pro Gly TCT GAA GGT AAA GGT GAA GTT AAA CTG GAT GAG ACT GGA GGA GGC TTG GTG CAA CCT GGG Arg Pro Met Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asp Tyr Trp Met Asn AGG CCC ATG AAA CTC TCC TGT GTT GCC TCT GGA TTC ACT TTT AGT GAC TAC TGG ATG AAC 170 180 Trp Val Arg Gln Ser Pro Glu Lys Gly Leu Glu Trp Val Ala Gln Ile Arg Asn Lys Pro	I le ATC	Ser AGC	Ser AGT	Val GTG	Lys AAG	Thr ACT	Glu GAA	Asp GAC	Leu CTG	Ala	Val GTT	Tyr TAT	Tyr TAC	Cys TGT	Gln CAG	Gln CAG	Tyr TAT	Tyr TAT	Ser AGC	Tyr
Ser Glu Gly Lys Gly Glu Val Lys Leu Asp Glu Thr Gly Gly Gly Leu Val Gln Pro Gly TCT GAA GGT AAA GGT GAA GTT AAA CTG GAT GAG ACT GGA GGA GGC TTG GTG CAA CCT GGG Arg Pro Met Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asp Tyr Trp Met Asn AGG CCC ATG AAA CTC TCC TGT GTT GCC TCT GGA TTC ACT TTT AGT GAC TAC TGG ATG AAC 170 180 Trp Val Arg Gln Ser Pro Glu Lys Gly Leu Glu Trp Val Ala Gln Ile Arg Asn Lys Pro	Pro CCC	Leu CTC	Thr ACG	Phe TTC	G ly GGT	Ala GCT	Gly GGG	Thr ACC	AAG	Leu CTT	Val GTG	Leu CTG	Lys	Gly	Ser	Thr	Ser TCC	Gly GGT	Lys AAA	Pro
Arg Pro Met Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asp Tyr Trp Met Asn AGG CCC ATG AAA CTC TCC TGT GTT GCC TCT GGA TTC ACT TTT AGT GAC TAC TGG ATG AAC 170 180 180 180 180	Ser TCT	Glu GAA	Gly	Lys AAA	Gly	Glu	Val	Lvs	Leu	130 Asp	Glu	Thr ACT	Gly GGA	Gly GGA	Gly GGC	Leu TTG	Val GTG	Gln CAA	Pro CCT	Gly
Tro Val Ara Gla Ser Pro Glu Lys Gly Leu Glu Tro Val Ala Gla Ile Arg Asa Lys Pro	Arg AGG	Pro CCC	Met ATG	Lys AAA	Leu CTC	Ser TCC	Cys TGT	Val GTT	Ala GCC	Ser	Glv	Phe TTC	Thr ACT	Phe TTT	Ser AGT	Asp GAC	Tyr TAC	Trp TGG	Met ATG	Asn
	Trp TGG	Va l GTC	Arg CGC	Gln CAG	Ser TCT	Pro CCA	Glu GAG	Lys AAA	Gly GGA	Leu	Glu	Trp TGG	Va l GTA	Ala GCA	Gln CAA	lle ATT	Arg AGA	Asn AAC	Lys AAA	Pro

FIG.19B

Tyr Asn Tyr Glu Thr Tyr Tyr Ser Asp Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp TAT AAT TAT GAA ACA TAT TAT TCA GAT TCT GTG AAA GGC AGA TTC ACC ATC TCA AGA GAT ASP Ser Lys Ser Ser Val Tyr Leu Gln Met Asn Asn Leu Arg Val Glu Asp Met Gly Ile GAT TCC AAA AGT AGT GTC TAC CTG CAA ATG AAC AAC TTA AGA GTT GAA GAC ATG GGT ATC ACC TYr Tyr Cys Thr Gly Ser Tyr Tyr Gly Met Asp Tyr Trp Gly Gln Gly Thr Ser Val Thr TAT TAC TGT ACG GGT TCT TAC TAT GGT ATG GAC TAC TGG GGT CAA GGA ACC TCA GTC ACC

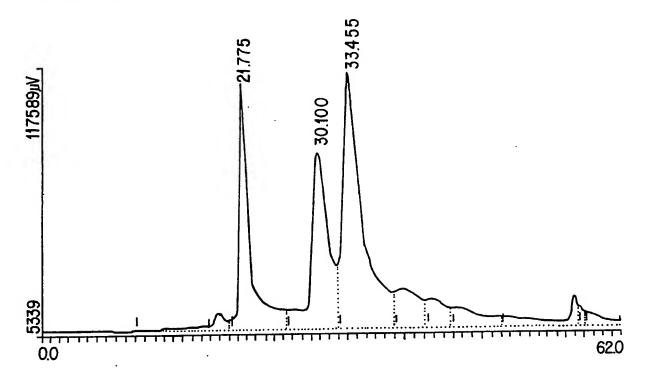
Val Ser *** *** Gly Ser
GTC TCC TAA TAA GGA TCC

Bam H1

FIG.19B(CONT.)

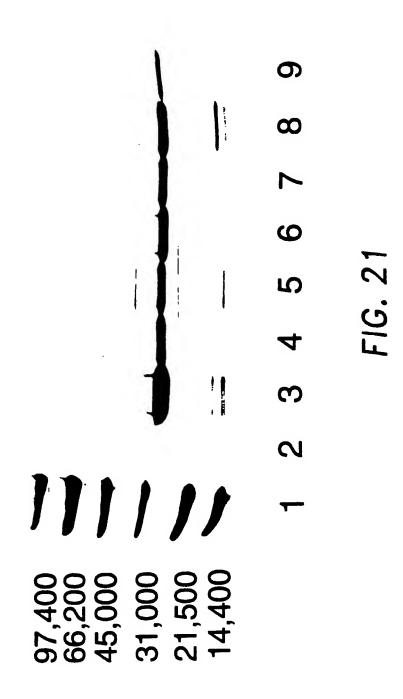
31/39

PROCESSING FILE: PolyCatA/Proc.CC-49Prep METHOD: PREP POLY CAT A#2 INJECT VOL: 44 SAMPLING INT: 0.3 SECONDS



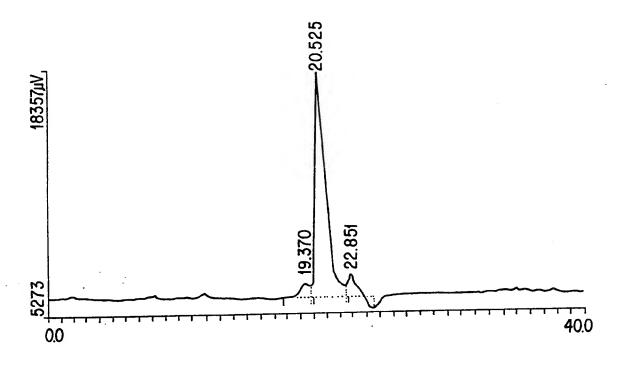
ANALYSIS:	CHANNEL A				
PEAK NO.	TIME	TYPE	HEIGHT(µV)	$AREA(\mu V-SEC)$	AREA%
1	17.090	N1	1651	348239	0.778
2	18.940	N2	8014	669441	1.496
ž	21.775	N3	104401	8617252	19.263
4	30.100	N4	74925	9753616	21.804
5	33.455	N5	106864	15749605	35.208
6	38.940	N6	17296	2833701	6.334
7	42.010	N7	12645	1637917	3.661
8	44.640	N8	9287	1968584	4.400
9	57.055	N9	13767	2012338	4.498
10	57.610	N10	9323	210914	0.471
11	58.240	X11	6824	930855	2.080
TOTAL AREA				44732462	99.993

FIG.20



PROCESSING FILE: PolyCotA/Proc.CC-49Prep METHOD: CC-49 QC SIZE-EXCLUSION INJECT VOL: .05

SAMPLING INT: 0.1 SECONDS



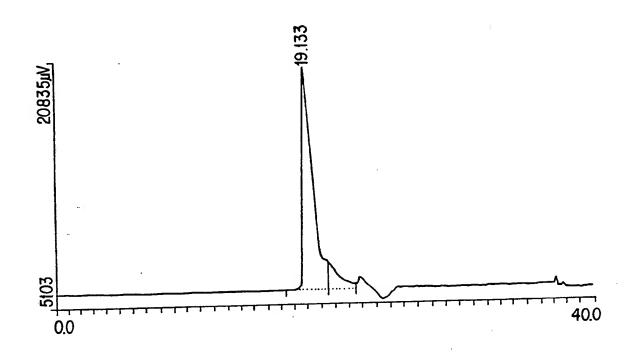
ANALYSIS:	CHANNEL A				
PEAK NO.	TIME	TYPE	HEIGHT(µV)	AREA(µV-SEC) 41706	AREA% 5.694
1 2 3	19.370 20.525 22.851	N1 N2 N3	797 11789 1227	657280 33466	89.737 4.569
TOTAL ARFA	22.001			732452	100.000

FIG.22A

PROCESSING FILE: PolyCatA/Proc.CC-49Prep METHOD: CC-49 QC SIZE-EXCLUSION

INJECT VOL: .05

SAMPLING INT: 0.1 SECONDS



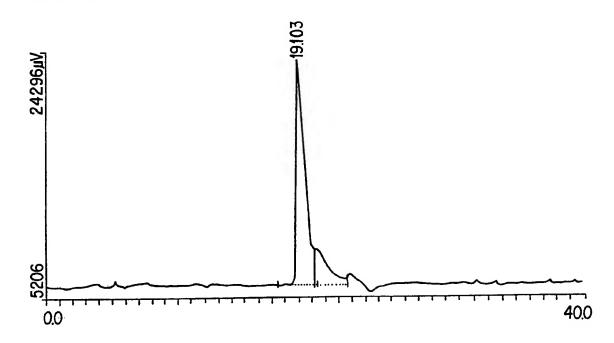
ANALYSIS:	CHANNEL A				
	THE	TYPE	HEIGHT(μV)	$AREA(\mu V-SEC)$	AREA%
PEAK NO.	TIME		** *	749671	88,214
1	19.133	N1	14211 1863	100154	11.785
2	20.538	N2	1003	849825	99.999
TOTAL AREA				043023	JJ.JJ

FIG.22B

PROCESSING FILE: PolyCatA/Proc.CC-49Prep METHOD: CC-49 QC SIZE-EXCLUSION

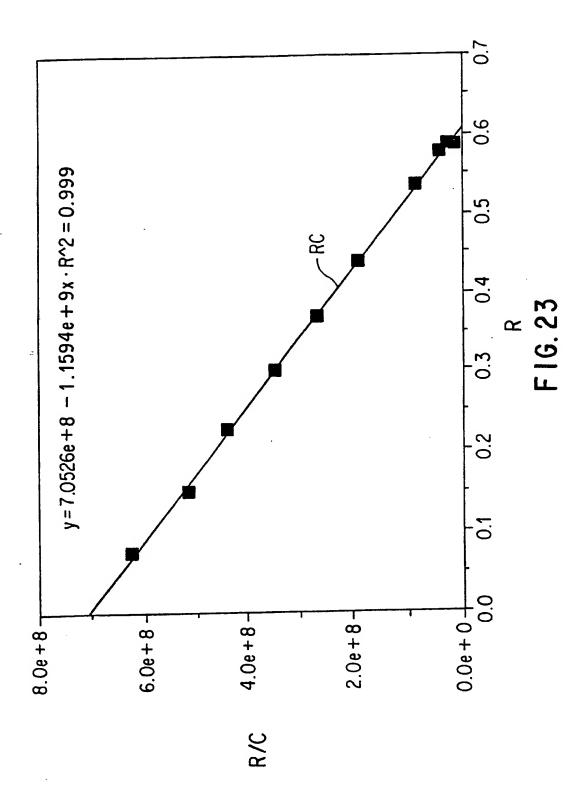
INJECT VOL: .05

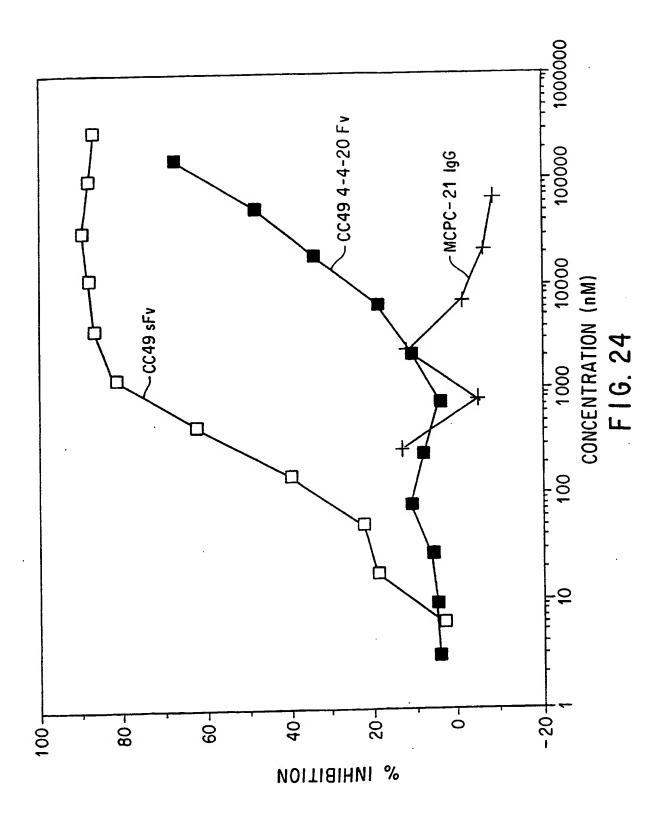
SAMPLING INT: 0.1 SECONDS



ANALYSIS:	CHANNEL A				•
PEAK NO.	TIME	TYPE	HEIGHT(μV)	AREA(µV-SEC)	AREA%
1	19.163	N1	17550	876502 17 9 029	83.039 16.961
2	20.435	N2	2981	1055531	100.000
TOTAL AREA				,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	

FIG.22C





SUBSTITUTE SHEET

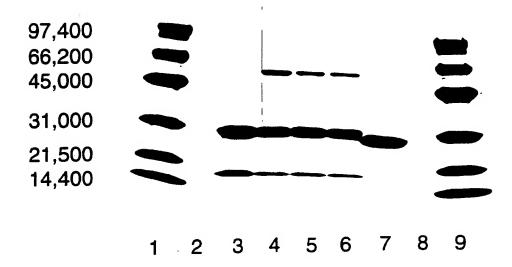


FIG. 25

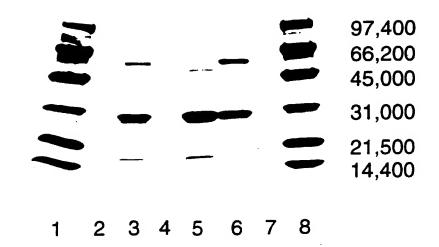


FIG. 26

INTERNATIONAL SEARCH REPORT

nucrnational application No.
PCT/US92/09965

A. CLASSIFICATION OF SUBJECT MATTER	R	TTE	ИΑ	٠.	СТ	E	BJ	U	S	OF	N	О	ITA	C_{ℓ}	FI	SI	ASS	Ľ	. C	A.
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IPC(5) :C07K 15/28, 3/20; C07H 21/04; C12P 21/08; C12N 15/00, 15/03;

US CL :Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC.

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S.: 530/387.3, 413; 435/7.92, 7.93, 7.94, 69.6, 69.7, 70.21, 172.2, 172.3, 240.27, 252.3, 320.1; 424/1.1, 85.8; 526/32.4, 23.53

536/23.4, 23.53

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
DIALOG (FILES 5, 73, 155, 351); U.S. AUTOMATED PATENT SYSTEM (FILE USPAT, 1971-PRESENT).

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X Y	WO 88/09344 (HUSTON et al.) 01 DECEMBER 1988, see entire document.	1-33,38-39,44-63 34-37,40-43
<u>X</u> Y	US, A, 4,946,778 (LADNER et al.) 07 AUGUST 1990, see entire document.	32-33,38-39 1-31,34-37,40-63
Y	CANCER RESEARCH, Vol. 48, issued 15 AUGUST 1988, Muraro et al., "Generation and Characterization of B72.3 Second Generation Monoclonal Antibodies Reactive with the Tumor-associated Glycoprotein 72 Antigen," pages 4588-4596, see entire document.	34-37,40-43
Y	SCIENCE, Vol. 242, issued 21 OCTOBER 1988, Bird et al., "Single-Chain Antigen-Binding Proteins," pages 423-426. See entire document.	32-42
Y	JOURNAL OF BIOLOGICAL CHEMISTRY, Vol. 265, No. 30, issued 25 OCTOBER 1990, Bedzyk et al., "Immunological and Structural Characterization of a High Affinity Anti-fluorescein Single-chain Antibody," pages 18615-18620. See entire document.	32-42

	Further documents are listed in the continuation of Box C	: Ц	See patent family annex.			
•	Special categories of cited documents:	·T-	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the			
,V.	document defining the general state of the art which is not considered to be part of particular relevance		principle or theory underlying the invention			
•Е•	·		document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step			
.r.	document which may throw doubts on priority claim(s) or which is		when the document is taken alone			
	cited to establish the publication date of another citation or other special reason (as specified)	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is			
.0.	document referring to an oral disclosure, use, exhibition or other means		combined with one or more other such documents, such combination being obvious to a person skilled in the art			
•P•	document published prior to the international filing date but later than the priority date claimed	.%.	document member of the same patent family			
Date of the actual completion of the international search			Date of mailing of the international search report			
23 FEBRUARY 1993			05 MAR 1993			
Name and mailing address of the ISA/ Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231			Authorized officer ROBERT D. BUDENS AMARINA KIY JA Telephone No. (703) 308-0196			
	nile No. NOT APPLICABLE	Telephone No. (703) 308-0196				

INTERNATIONAL SEARCH REPORT

In...ational application No. PCT/US92/09965

A. CLASSIFICATION OF SUBJECT MATTER: US CL :

530/387.3, 413; 435/7.92, 7.93, 7.94, 69.6, 69.7, 172.3, 252.3, 320.1; 424/1.1, 85.8; 536/23.53

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING This ISA found multiple inventions as follows:

- I. Claims 1-14, 19-29, 31, 44-46, 51-58, a first product, method of making and method of using, drawn to multivalent antigen-binding proteins, compositions, methods of making multivalent proteins and method of using multivalent proteins to detect antigens, classified in Class 530, subclass 387.3 and Class 435, subclasses 7.1, 69.7, 172.3, 320.1, 252.3.
- II. Claims 15-18, a second product, drawn to compositions containing multivalent proteins and single chain proteins, classified in Class 530, subclass 387.3.
- III. Claim 30, a second method of use, directed to a method of imaging, classified in Class 424, subclass 85.8.
- IV. Claims 32-37, a third product, drawn to single chain proteins classified in Class 530, subclass 387.3.
- V. Claims 38-43 and 47-50, a fourth product, drawn to genetic sequences, vectors and hosts, classified in Class 536, subclass 23.53, Class 435, subclasses 320.1 and 252.3.
- VI. Claims 59-61, a third method of use, drawn to immunoassay methods, classified in Class 435, subclasses 7.92, 7.93 and 7.94.
- VII. Claim 62, a fourth method of use, drawn to a method of immunotherapy using multivalent proteins, classified in Class 424, subclass 85.8.
- VIII. Claim 63, a fifth method of use, drawn to a method of ununoaffinity purification using multivalent proteins, classified in Class 530, subclass 413.

The inventions of Groups I-II and IV-V are directed to multiple products which differ in their physical properties such as primary sequence, molecular weights and chemical composition and are not so linked as to have a common special technical feature.

Further, the methods of Groups I, III and VI-VIII differ in their utilities, reagents and method steps and are not so linked as to have a common special technical feature.

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